

REVIEW

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Recent advances of enterovirus 71 3C^{pro} targeting Inhibitors

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Abstract

With CA16, enterovirus-71 is the causative agent of hand foot and mouth disease (HFMD) which occurs mostly in children under 5 years-old and responsible of several outbreaks since a decade. Most of the time, HFMD is a mild disease but can progress to severe complications such as meningitis, brain stem encephalitis, acute flaccid paralysis (AFP) and even death; EV71 has been identified in all severe cases. Therefore, it is actually one of the most public health issues that threatens children's life. 3C^{pro} is a protease which plays important functions in EV71 infection. To date, a lot of 3C^{pro} inhibitors have been tested but none of them has been approved yet. Therefore, a drug screening is still an utmost importance in order to treat and/or prevent EV71 infections. This work highlights the EV71 life cycle, 3C^{pro} functions and 3C^{pro} inhibitors recently screened. It permits to well understand all mechanisms about 3C^{pro} and consequently allow further development of drugs targeting 3C^{pro}. Thus, this review is helpful for screening of more new 3C^{pro} inhibitors or for designing analogues of well known 3C^{pro} inhibitors in order to improve its antiviral activity.

Keywords: Enterovirus 71, Enterovirus 71 life cycle, 3C^{pro} functions, 3C^{pro} inhibitors, EV71 drugs screening

Background

Enterovirus 71, belongs to human enterovirus A species, *Picornaviridae* family, was discovered in a patient with central nervous system (CNS), in California, 1969 [1]. In term of structure, EV71 is a non-enveloped virus with a capsid made up of 60 protomers of envelop proteins and contains a single-stranded RNA positive [2, 3]. Each protomer contains four envelop proteins: VP1–VP2–VP3, located in the external part and are exposed to the host antibodies and cell receptors; and VP4 which is completely hidden in the internal part. The RNA genome is small (7.5 kb) and constituted by 3 parts: P1, P2 and P3, flanked by 2 UTRs (non-translated regions) located in 5' and 3' [4]. Several outbreaks and fatal cases, caused by this virus, make it a major public health issue mainly in

the Asia-Pacific region. Indeed, China has experienced the latest and largest outbreaks with more than 1.7 million cases, 27.000 patients with severe neurological complications and 905 deaths, in 2010 [5]; while a cyclical and seasonal pattern occurs in Sarawak, Japan, Taiwan and Vietnam [6–9]. To manage such infections and epidemics is primordial, and the best way to eradicate this infection is the combination of a valuable vaccine and drugs [10]. Nevertheless, vaccine research has progressed more than drugs discovery because to date there is no approved drug against EV71 while 3 vaccines have completed their clinical trials III and are in following-up stage [11]. For this reason, the treatment is only symptomatic along with public surveillance systems [12]. Many plant extracts and chemical compounds have been discovered as having a potential effects against the virus and might be used as drugs against enterovirus 71 infections but none of them has been approved yet [13]. Thus, the finding of an approved and valuable drug is still an utmost importance. 3C^{pro} represent a valuable target because it has primordial functions in both virulence and virus-host

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interactions. This review highlights the important functions and recent progress of 3C^{pro} inhibitors and permit to acknowledge that 3C^{pro} is a valuable target for EV71 drug development, which should be deeply investigated.

Review on EV-71 life cycle

The EV71 life cycle goes through an attachment and entry, via a recognition and binding of surface protein to the cell receptors (SCARB2, PSGL-1, Anx2, Heparan Sulfate, Sialylated glycan) [14], to the release of the new virions by cell lysis (Fig. 1a). The mechanism of entry is known as through clathrin-mediated endocytosis (real events remain unclear) but recent investigation has showed that multiple pathways may be used by EV71 to enter the host cells [15, 16]. Then, a series of conformational changes occurs at low pH and let the virus to leave his icosahedral capsid structure to an A-particle: loss of VP4 and formation of a channel followed by a release of RNA in cell cytoplasm [17]. Once the RNA is located in the cytoplasm, the viral genome, as a positive sense, act as an mRNA, so directly translated into a polyprotein (P1, P2, P3) of 2193 AA. The polyprotein processing is assured by two main proteins 2A^{pro} and 3C^{pro}. Thus, 2A^{pro} and 3C^{pro} cleaved the polyprotein into VP1–VP4 (structural protein) and 2A–2C, 3A–3D (non-structural protein) [18]. When a considerable number of the 11 mature proteins are synthesized, the RNA replication take place after the interactions of IRES-specific-trans-acting factors (ITAFs), which are translocated from the nucleus to the cytoplasm, with the internal ribosome site (IRES) at its stem-loop [19, 20]. A negative-RNA is first synthesis within using the viral genome as template, and then followed by synthesis of numerous positive-strands using in turn the negative-strand as template. RNA-dependent RNA-polymerase (RdRp) or 3D^{P01} is the viral enzyme responsible of the RNA synthesis [18]. Finally, the structural proteins and the genome is encapsidated to form a new virion which is released during lysis of the cell (apoptosis).

3C^{pro} functions

In addition to its polyprotein processing activity, the non-structural protein 3C plays a role in numerous biological mechanisms. Recent discovery of the 3C crystal structure has permit to identify the sites of its substrat binding affinity (between 2 similar β -ribbon) and confirmed its cleavage activity of the viral polyprotein but also several host proteins in order to optimize viral replication and spreading [21]. EV71 infection symptoms range from mild to severe diseases which depend on both the viral genetic sequence and the host immune system. In fact, the relationship between 3C genome sequence and the corresponding clinical symptoms (mild or severe)

revealed that the 79th residue is the responsible sequence that leads to severe diseases [22]. Besides, Li et al. [23] have found another residue associated with the virulence of EV71, their finding suggests that the 69th residue is the virulent determinant because a single mutation of the hydrogen bond between Asn69 and Glu71 causes a significant decrease in the EV71 infection. The same result was found during the study of NK-1.8k compound where the substitution of asparagine at 69th residue by serine has decreased the fitness of the virus but on the other hand causes total resistance towards the tested compound. Indeed, the 69th residue plays an important role in 3C^{pro} functions even if it is not directly part of the active site according to the crystal structure [24]. EV71 interacts with the innate immune system through PRRs (Pattern-recognition receptors) such as TLRs which is involved in IFN – I production, RLRs responsible for detection of RNA virus infection and NLRs which function is to form cytosolic inflammasome [25]. In fact, concomitantly with the virus invasion, different host-immune responses occur such as production of type I interferon (IFN(α/β)) ; then to escape and to impair the immunity, the virus uses the proteolytic activity of 3C^{pro} by cleaving numerous needed host proteins: KPNA-I in order to suppress the signaling pathway STAT/KPNA-I [26], TAK1/TAB1/TAB2/TAB3 complex [27], TRIF, shut-off IR3/7 [28] and consequently block the production of IFN(α/β). Likewise, to permit the release and spread of virus progeny, 3C induced apoptosis of host cells through the caspase-3 pathway [29], cleavage of hnRNPA1 [30] and PinX1 [31]. Finally, 3C is able to enter the nuclei through its precursor 3CD [32] and cleaves the polyadenylation factor CstF-64. As a result, the host mRNA 3' polyadenylation ,which is essential for its translocation, stability and translation, is shut off [33] (Fig. 1b). Due to such functions, 3C is definitely an excellent target for drug screening.

3C^{pro} inhibitors

3C^{pro} is an important target to block EV71 replication. Indeed, several 3C^{pro} inhibitors have been deeply investigated (Table 1, Fig. 1b)

Peptidomimetic compounds

- (a) Rupintrivir and analogues: Rupintrivir (AG7088) is probably the well-known 3C^{pro} inhibitors to date. More than being a safe compound for the cells, it is able to bind to the active site of 3C^{pro} [21]. It was firstly identified as 3C Human Rhinovirus (HRV) inhibitors, later Zhang et al. [34] shown that it also had a strong antiviral activity against EV71 3C^{pro} in both cell lines and animal models. In fact, AG7088

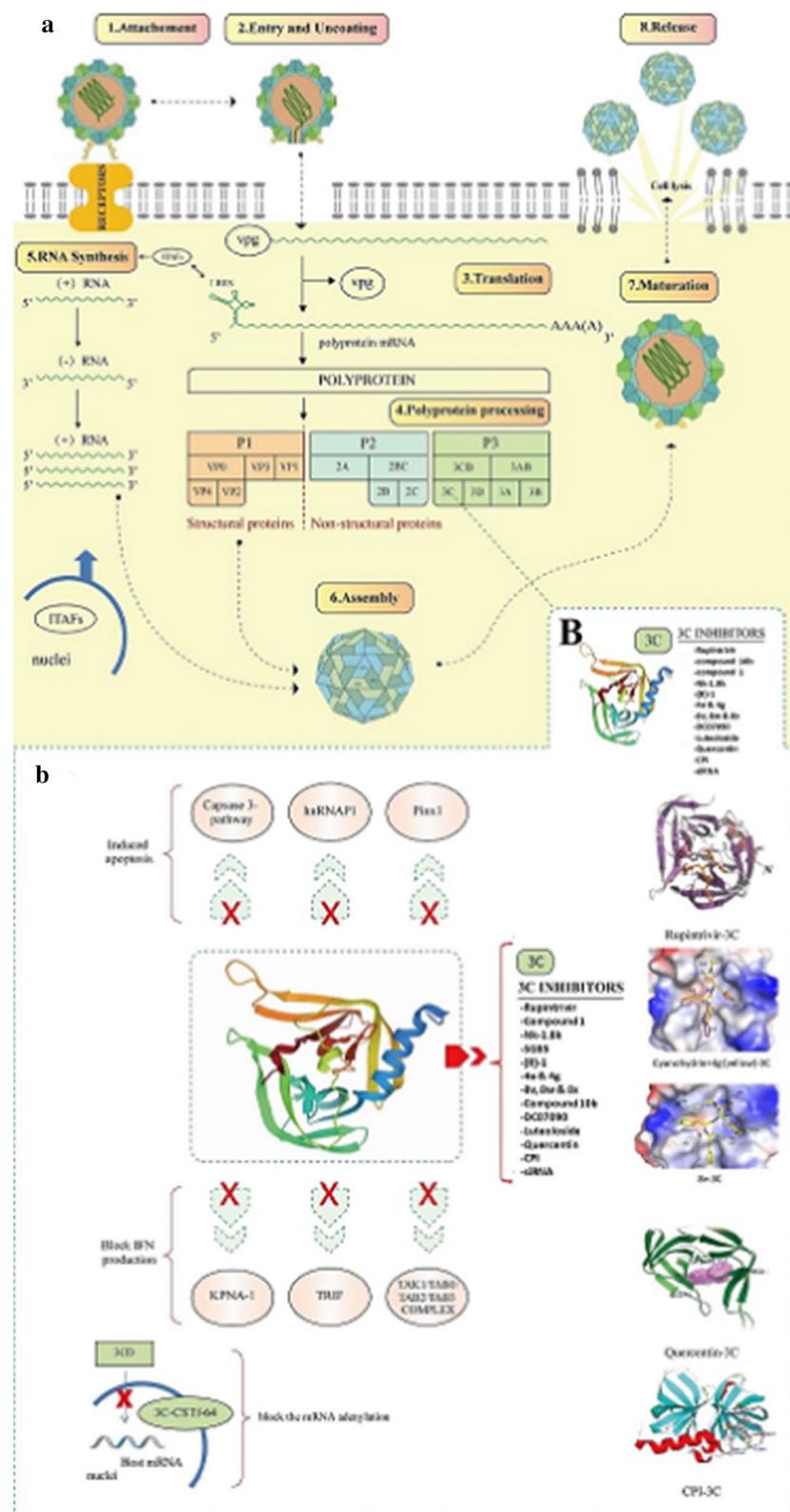


Fig. 1 Illustration of EV71 life cycle and virus-host interactions. EV71 replication steps: from attachment to release (a). 3C-host proteins interactions are blocked by 3C^{pro} inhibitors (b)

inhibits the antiviral activity at $EC_{50} = 0.01 \mu M$ and protease activity at $IC_{50} = 2.5 \pm 0.5 \mu M$ with $CC_{50} = 1000 \mu M$; in-vivo a low dose of 0.1 mg/kg prevent severe symptoms in suckling mice. Since the discovery of this compound, several analogues have been designed in order to increase its efficiency against EV71 infection [21]. To improve the anti-EV71 activity of rupintrivir, Kuo et al. has

designed several inhibitor analogues (compound 1 to 10b) by replacing the P3 group of AG7088 with a series of cinnamoyl derivates. The compound 10b seemed to be potentially effective against EV71 among all the analogues, with an EC_{50} and CC_{50} of $0.018 \mu M$ and $> 25 \mu M$ respectively [35]. Then later Shang et al. [36] replaced the cinnamoyl of compound 1 to 2-chloride-phenylacetyl and noticed

Table 1 Detailed list and classification of 3C^{Pro} inhibitors: chemical structure, classes, effectivity, test in cell lines and animal models

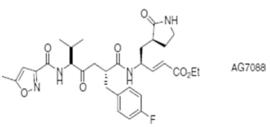
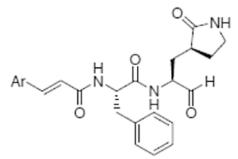
Chemical structures	Classes	Compound's names	IC ₅₀ /EC ₅₀	Cell lines and animal models	References
  10b Ar = 4-Me ₂ NC ₆ H ₄	Peptidomimetic compounds	AG7088	0.01 μM	RD, 2 days suckling mice	[21]
		Compound 10b	0.018 μM	RD	[35]
		Compound 1 (with 2-chloride-phenylacetyl)	1.89±0.25 μM	RD	[36]
		NK-1.8k	0.108 μM and 2.41 μM	RD, T293, Vero	[24]
		SG85	180 nM to 0.200 μM	RD, Huh7, Vero, BGM, HeLa	[38,39]
		(R)-1	0.088±0.006 μM	RD, 293T	[40]

Table 1 (continued)

<p>4e</p> <p>4g</p> <p>R</p> <p>R'</p> <p>8v</p> <p>8w</p> <p>8x</p> <p>R'</p>		4e and 4g 0.21 ± 0.05 and $0.033 \pm 0.008 \mu\text{M}$ respectively	Adult male mice	[41]	
	Non-peptidyl compound	DC07090	$22.09 \pm 1.07 \mu\text{M}$	RD	[43]
<p>Luteolin</p> <p>Quercetin</p> <p>CPI</p>	Flavonoids	Luteoloside	$0.36 \text{ mM}/0.43 \text{ m M}$	RD	[45]
		Quercetin	$8.8 \mu\text{M} / 12.1 \mu\text{M}$	RD/ Vero	[46]
		CPI	$4.03 \mu\text{M}$	RD	[47]
	RNA interference	siRNA		RD/ suckling mice	[48,49]

- that the efficiency of its antiviral activity has been increased twice $IC_{50} = 1.89 \pm 0.25 \mu\text{M}$. Another method to further improve rupintrivir action is to combine it with IFN(α/β). In fact, it was proved that rupintrivir and Interferon had an synergistic inhibition against EV71 infection [37].
- (b) NK-1.8k: is a peptidyl aldehyde discovered to have strong anti-viral activity against not only EV71 but also the Enterovirus 68. The mechanism of action is known as the same as rupintrivir which targeted the 3C^{pro} EV71 in dependent-concentration manner. However, structurally, they are different because NK-1.8k is a dipeptide with six-member-ring lactam and rupintrivir, a tripeptide with five-member-ring lactam. Thus, its structure confers to NK-1.8k a better stability and drug features than rupintrivir which is always taken as reference. Indeed, NK-1.8k decrease the viral RNA production at $EC_{50} = 34.5 \text{ nM}$. Moreover, it is potent in all the 3 genotypes of EV71 in different cell lines (RD and T293 $EC_{50} = 0.108 \mu\text{M}$; Vero $EC_{50} = 2.41 \mu\text{M}$) [24]. NK-1.8k represents a new peptidomimetic compound which might take the place of rupintrivir as an archetype in EV71 drug screening.
 - (c) SG85: the 3C^{pro} inhibitors SG85 is a peptidic Michael acceptor compound. It has been tested against Enterovirus 68, EV71, echovirus 11 and various rhinovirus serotypes. However, it was found to be more potent against HRV11 and EV71 with $EC_{50} = 60 \text{ nM}$, $EC_{50} = 180 \text{ nM}$ respectively [38]. Furthermore, it has screened to have strong antiviral activity against all the 11 EV71 strains with EC_{50} between 0.039 and 0.200 μM [39]. Deep study of SG85 is needed in order to progress the drug discovery of EV71.
 - (d) (R)-1: is proved to be one of the most efficient 3C^{pro} inhibitors screened to date with an $EC_{50} = 0.088 \pm 0.006 \mu\text{M}$. However, the presence of cyanohydrins, which is labile, gives it unstable and toxic properties [40].
 - (e) 4e and 4g: are compounds resulted from improvement of (R)-1. In fact, acyl cyanohydrins which make unstable (R)-1 have been replaced by 4-iminoazolidin-2-one. After a series of test, 4e and 4g were the compound having the most potent antiviral activity with $EC_{50} = 0.21 \pm 0.005$ and $0.033 \pm 0.008 \mu\text{M}$ respectively. Moreover, those compounds are safe towards the cell ($CC_{50} > 100 \mu\text{M}$). Thus, they can be used as base for EV71 drug therapy [41].
 - (f) 8v, 8w and 8x: are alpha-keto-amid inhibitors against EV71 3C^{pro}. Zeng et al. noticed that the pivotal function of 3C^{pro} makes it the ideal target to fight against EV71 infection. Then, they synthesized several alpha-keto-amids as 3C inhibitors via Passerini reaction. Hence, the compounds 8v, 8w and 8x were exhibiting the most potent antiviral activity against enterovirus 71 with $EC_{50} = 1.32 \pm 0.26$, 1.88 ± 0.35 and $1.52 \pm 0.31 \mu\text{M}$ respectively. Nevertheless, those compounds should be more improved and studied in order to contribute for EV71 drug discovery which is currently in need [42].
- #### Non-peptidyl compound: DC07090
- Recently identified as novel small potent molecule 3C inhibitor, it is a non-peptidyl compound designed by docking-based virtual screening and able to bind with 3C through its binding site and reversible inhibits its protease activity at $EC_{50} = 22.09 \pm 1.07 \mu\text{M}$. Besides, DC07090 has a very low cytotoxicity rate ($CC_{50} > 200 \mu\text{M}$) which makes it an attractive compound for further drug development [43].
- #### Flavonoids
- Flavonoids, originally synthesized by the plants as abiotic stresses: in order to protect themselves against ultraviolet radiation, pathogens and herbivores are a group of natural compounds largely distributed in fruits, vegetables, tea, soy foods and herbs. Most importantly, they have huge therapeutic bioactivities: anti-oxidative, anti-inflammatory and antiviral properties. Researchers used them as a base of drug and dietary supplement in several diseases [44]. They present an attractive therapy for Enterovirus 71 due to their low toxicity towards host cells and their strong antiviral activity.
- (a) Luteoloside: is a flavonoid distributed mainly in *Lonicera japonica*, plant used in traditional Chinese medicine, and has got broad activities such as anti-microbial, anti-cancer and antiviral activity against influenza virus, human rhinovirus, coxsackievirus B4 and enterovirus 71. The real mechanisms against EV71 remain unknown and need further deep to elucidate but it is sure that it blocked the pathway at 3C protease activity stage, $IC_{50} = 0.36 \text{ mM}$ with a selectivity index of 5.3 according to the investigation of Cao et al. Therefore, it is an excellent candidate for drug development [45].
 - (b) Quercetin: is a member of the flavonol subgroup of flavonoid found in many plants, fruits, grains and vegetables with anti-inflammatory, anti-cancer and anti-viral properties. It is probably one of the latest 3C inhibitor tested. Without toxicity towards

the cells, our group's recent finding reveals that quercetin exhibits a prominent effectiveness against the protein 3C of enterovirus 71 by binding its substrate-binding pocket. Moreover, quercetin seems to have a preventive action. Indeed, cells pre-treated by quercetin present a high survival rate when infected by EV71 virus. Consequently, quercetin may be used both in preventive and in therapeutic application [46]. Therewith, a drug library composed of 1430 FDA approved drugs were previously screened from our laboratory. Interestingly, we found that the compound 3 had significantly anti-EV71 effect among them. Further mechanism study revealed that it targeted viral 3C protease and block viral replication (unpublished data).

- (c) Diisopropyl Chrysin-7-i1 Phosphate (CPI): is a phosphate ester of chrysin, a natural flavonoid found in many plants. CPI is able to bind in the pocket site of hydrophobic and polar residue of 3C protease like LEU4- 8, SER-I III, MET-112, PHE-113 and PRO-115 and inhibits the protease activity at EC₅₀ = 4.03 mM. Indeed, 3C^{Pro} is unable to cleave human interferon regulator factor 9(IRF9) in the presence of CPI [47].

siRNA

siRNA is a powerful tool which can be used to target a specific gene in order to suppress it. Small interfering RNA therapeutics has been explored against several human viral infections including Enterovirus due to its specificity and promising effect both in-vitro and in-vivo [48]. Indeed, siRNA recognize, bind and degrade the target mRNA. It is a challenging strategy by the potential risk of mutation, inflammation or immune responses. However, Yang et al. showed that there is any toxicity of the siRNA targeted 3C^{Pro} and 3D^{Pol} during their investigation. They have designed a novel minicircle vector through 3C^{Pro} and 3D^{Pol} sequence available in Genbank. In fact, the siRNA did not affect the growth and viability of the cell. Moreover, it has reduced the protein levels to 10.8 ± 6.7%, the viral mRNAs to 12.4 ± 1.75% and the progeny virion production to 15% in infected cells. More importantly, it has protected the infected-suckling mice of a significant weight loss and hind limbs paralysis. Hence, further investigation must be conducted about silencing gene strategy within using 3C^{Pro} as target [49].

Discussion

The unavailable of approved clinical drug makes the finding of a potent compound against EV71 really important. 3C^{Pro} is an essential protein for EV71 life cycle and infection, moreover, it has strict substrate and does not

have a lot of homologues in mammalian cells [35]. Thus, it is an excellent and attractive target for development of potent drugs. In this review, we summarized several classes of compound recently screened and also rupintrivir which is the drug of reference against 3C^{Pro}. Actually, rupintrivir and analogues are considered as the most potent 3C^{Pro} inhibitors. However, NK-1.8k has almost the same potency and efficiency as rupintrivir (Table 1), and as more stable, it can take the place of rupintrivir as archetype of 3C^{Pro} inhibitors. In fact, peptidomimetic compounds represent the most potent class with the minimal effective concentration (180 nM to 2.89 μM, Table 1). It might be due to the fact that they are synthetically designed to fit in the 3C^{Pro} active pocket. Nevertheless, flavonoids class, which is composed of active compounds from plants, has satisfactory antiviral activity as well. Indeed, nowadays, the trend of using bioactive compounds as drug candidates is done more and more, because of their broad biological and pharmacological activities, their availability and safety towards the host cells. Besides, the screening of non-peptidyl compound has been tempted but only DC07090 among 50 other compounds has given a satisfactory result [43]. Peptidomimetic compounds might be more potent and interesting than non-peptidyl-compounds. Hence, deep investigation, mainly in an appropriate animal model, should be done for luteoloside, quercetin and CPI which could be approved as EV71 therapy; while more and more peptidomimetic compounds should be designed and/or improved by using the revelation of 3C^{Pro} structure as reference. Following the drug screening work, the 69th residue of 3C^{Pro}, which plays important role in conferring EV71 resistance, could be investigated in order to make sure that the virus will not develop a resistance mutation toward the potent drug as investigated by Wang et al. [24]. Finally, the last recent strategy is the use of RNAi. In fact, there are few investigation about siRNA as therapy against EV71 infection; however, it has been successful against a wide range of viruses: Human immunodeficiency virus, hepatitis B/C virus, Influenza virus [50–53]. Therefore, even if it is a challenging technique, investigating this strategy is worth it.

Conclusion

Coupling an effective vaccine and drugs against Enterovirus 71 is the most prominent manner to eradicate EV71 infection. The prevention will be secure by the vaccine and the treatment by an effective drug. However, the drug progress has not been as developed as for vaccines. In fact, currently only a surveillance is set up to control the disease. EV71 is a threat for children's life; therefore, the screening of an effective drug is quite indispensable as soon as possible. For that, 3C^{Pro} represent an excellent target due to

the several key functions that it plays in both virulence and interaction of the virus to the host. More 3C^{pro} inhibitors should be exploited. Besides, as 3C^{pro} and 2A^{pro} play role in early stage of the viral replication through cleaving the EV71 polyprotein, a combination of 2A^{pro} and 3C^{pro} inhibitors in order to act in a synergistic manner may represent a valuable strategy. Indeed, the 3C X-ray structure is already defined so it would promote further studies of its protease activity inhibitions by a compound. Meanwhile, all drugs screening must be tested in an appropriate animal model which will be compared to the in-vitro screening in order to achieve the goals of using it as treatment against EV71 infections.

Abbreviations

EV71: Enterovirus 71; CA16: Coxsackievirus A16; HFMD: Hand, foot and mouth disease; AFP: Acute flaccid paralysis; 3C^{pro}: 3C protease; CNS: Central Nervous System; RNA: Ribonucleic acid; UTRs: non-translated regions; SCARB-2: Scavenger receptor class B member 2; PSGL-1: P-selectin glycoprotein ligand-1; Anx2: Annexin-2; 2A^{pro}: 2A protease; 3D^{pol}: 3D polymerase; ITAFs: IRES-specific-trans-acting factors; IRES: Internal ribosome site; RdRp: RNA-dependent RNA-polymerase; PRRs: Pattern-recognition receptors; TLRs: Toll-like receptors; IFN- λ : Interferon type I; RLRs: RIG-I-like receptors; NLRs: NOD-like receptors; IFNa/ β : Interferon alpha/beta; KPNA-1: Karyopherin subunit Alpha-1; STAT: Signal transducer and activator of transcription; TAK1: Transforming growth factor beta activated kinase; TAB1/2/3: TGF-beta activated kinase 1/2/3; IR3/7: Interferon regulator 3/7; hnRNP: heterogeneous nuclear ribonucleoprotein; PinX1: PIN2 interacting telomerase inhibitor-1; mRNA: messenger Ribonucleic acid; HRV: Human Rhinovirus; EC₅₀: 50% Effective Concentration; IC₅₀: 50% Inhibition concentration; CC₅₀: 50% Cytotoxic Concentration; FDA: Food and Drug Administration; CPI: Chrysanthemum 7-Il Phosphate; LEU: Leucine; SER: Serine; MET: Methionine; PHE: Phenylalanine; PRO: Proline; siRNA: Small Interfering RNA; RNAi: RNA interference; RD: Rhabdomyosarcoma; Vero: Verda Reno; BGM: Buffalo green monkey kidney cells; Hela: Henrietta Lack.

Acknowledgements

Not applicable.

Author's contributions

RD wrote the review under the lead, supervision and correction of HK. All the authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

Not applicable.

Ethics approval and consent for participation

Not applicable.

Consent for publication

Not applicable.

Competing interests

Not applicable.

Received: 11 August 2020 Accepted: 7 October 2020

Published online: 11 November 2020

References

- Schmidt NJ, Lennette EH, Ho HH. An apparently new enterovirus isolated from patients with disease of the central nervous system. *J Infect Dis*. 1974;129(3):304–9. <https://doi.org/10.1093/infdis/129.3.304>.
- Putnak JR, Phillips BA. Picornaviral structure and assembly. *Microbiol Rev*. 1981;45(2):287–315. <https://doi.org/10.1128/mmbr.45.2.287-315.1981>.
- Yuan J, et al. Enterovirus A71 proteins: structure and function. *Front Microbiol*. 2018;9:286. <https://doi.org/10.3389/fmicb.2018.00286>.
- Solomon T, Lewthwaite P, Perera D, Cardosa MJ, McMinn P, Ooi MH. Virology, epidemiology, pathogenesis, and control of enterovirus 71. *Lancet Infect Dis*. 2010;10(11):778–90. [https://doi.org/10.1016/S1473-3099\(10\)70194-8](https://doi.org/10.1016/S1473-3099(10)70194-8).
- Zeng M, et al. Seroepidemiology of Enterovirus 71 infection prior to the 2011 season in children in Shanghai. *J Clin Virol*. 2012;53(4):285–9. <https://doi.org/10.1016/j.jcv.2011.12.025>.
- Podin Y, et al. Sentinel surveillance for human enterovirus 71 in Sarawak, Malaysia: lessons from the first 7 years. *BMC Public Health*. 2006;6:180. <https://doi.org/10.1186/1471-2458-6-180>.
- Mizuta K, et al. Molecular epidemiology of enterovirus 71 strains isolated from children in Yamagata, Japan, between 1990 and 2013. *J Med Microbiol*. 2014;63:1356–62. <https://doi.org/10.1099/jmm.0.079699-0>.
- Te Lee J, et al. Enterovirus 71 seroepidemiology in Taiwan in 2017 and comparison of those rates in 1997, 1999 and 2007. *PLoS ONE*. 2019;14(10):e0224110. <https://doi.org/10.1371/journal.pone.0224110>.
- Van Le T, Nguyen VTT, Nguyen QH, Pham DT. Molecular epidemiology analysis of enterovirus 71 strains isolated in Dak Lak, Vietnam, 2011–2016. *J Med Virol*. 2019;91(1):56–64. <https://doi.org/10.1002/jmv.25286>.
- Liang Z, Wang J. EV71 vaccine, an invaluable gift for children. *Clin Transl Immunol*. 2014;3(10):e28. <https://doi.org/10.1038/cti.2014.24>.
- Lin JY, Kung YA, Shih SR. Antivirals and vaccines for Enterovirus A71. *J Biomed Sci*. 2019;26(1):65. <https://doi.org/10.1186/s12929-019-0560-7>.
- Lin C, Chen KH, Tong Chen K. Update on enterovirus 71 infections: epidemiology, molecular epidemiology, and vaccine development. *J Infect Dis Ther*. 2018. <https://doi.org/10.4172/2332-0877.1000370>.
- Wang H, Li Y. Recent progress on functional genomics research of enterovirus 71. *Virol Sin*. 2019;34(1):9–21. <https://doi.org/10.1007/s12250-018-0071-9>.
- Yamayoshi S, Fujii K, Koike S. Receptors for enterovirus 71. *Emerg Microbes Infect*. 2014;3:e53. <https://doi.org/10.1038/emi.2014.49>.
- Xu Y, Liu Q, Zhang Z. Human EV71 invades human neuroblastoma SK-N-SH cells by clathrin-mediated endocytosis. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*. 2017;33(6):761–6.
- Yuan M, et al. Enhanced human enterovirus 71 infection by endocytosis inhibitors reveals multiple entry pathways by enterovirus causing hand-foot-and-mouth diseases. *Virol J*. 2018;15(1):1. <https://doi.org/10.1186/s12985-017-0913-3>.
- Danithi P, Tosteson M, Li Q, Chow M. Genome delivery and ion channel properties are altered in VP4 mutants of poliovirus. *J Virol*. 2003;77(9):5266–74. <https://doi.org/10.1128/jvi.77.9.5266-5274.2003>.
- Baggen J, Thibaut HJ, Strating JR, van Kuppeveld FJ. The life cycle of non-polio enteroviruses and how to target it. *Nat Rev Microbiol*. 2018;16(6):368–81. <https://doi.org/10.1038/s41579-018-0005-4>.
- Huang PN, et al. Far upstream element binding protein 1 binds the internal ribosomal entry site of enterovirus 71 and enhances viral translation and viral growth. *Nucleic Acids Res*. 2011;39(22):9633–48. <https://doi.org/10.1093/nar/gkr682>.
- Lin JY, Li ML, Shih SR. Far upstream element binding protein 2 interacts with enterovirus 71 internal ribosomal entry site and negatively regulates viral translation. *Nucleic Acids Res*. 2009;37(1):47–59. <https://doi.org/10.1093/nar/gkn901>.
- Wang J, et al. Crystal structures of enterovirus 71 3C protease complexed with rupintrivir reveal the roles of catalytically important residues. *J Virol*. 2011;85(19):10021–30. <https://doi.org/10.1128/jvi.05107-II>.
- Ma HY, et al. Association of EV71 3C polymorphisms with clinical severity. *J Microbiol Immunol Infect*. 2018;51(5):608–13. <https://doi.org/10.1016/j.jmii.2016.12.006>.
- Bingqing L, et al. A novel enterovirus 71 (EV71) virulence determinant: The 69th residue of 3C protease modulates pathogenicity. *Front Cell Infect Microbiol*. 2017;7:26. <https://doi.org/10.3389/fcimb.2017.00026>.
- Wang Y, Yang B, Zhai Y, Yin Z, Sun Y, Rao Z. Peptidyl aldehyde NK-1.8k suppresses enterovirus 71 and enterovirus 68 infection by targeting protease

- 3C. *Antimicrob Agents Chemother*. 2015;59(5):2636–46. <https://doi.org/10.1128/AAC.00049-15>.
25. Chen KR, Ling P. Interplays between enterovirus A71 and the innate immune system. *J Biomed Sci*. 2019;26(1):95. <https://doi.org/10.1186/s12929-019-0596-8>.
26. Wang C, et al. Enterovirus 71 suppresses interferon responses by blocking Janus kinase (JAK)/signal transducer and activator of transcription (STAT) signaling through inducing karyopherin- α 1 degradation. *J Biol Chem*. 2017;292(24):10262–74. <https://doi.org/10.1074/jbc.M116.745729>.
27. Lei X, Han N, Xiao X, Jin Q, He B, Wang J. Enterovirus 713C inhibits cytokine expression through cleavage of the TAK1/TAB 1/TAB 2/TAB3 complex. *J Virol*. 2014;88(17):9830–41. <https://doi.org/10.1128/jvi.04251-14>.
28. Lei X, Xiao X, Xue Q, Jin Q, He B, Wang J. Cleavage of interferon regulatory factor 7 by enterovirus 713C suppresses cellular responses. *J Virol*. 2013;87(3):1690–8. <https://doi.org/10.1128/jvi.01855-12>.
29. Li ML, et al. The 3C protease activity of enterovirus 71 induces human neural cell apoptosis. *Virology*. 2002;293(2):386–95. <https://doi.org/10.1006/viro.2001.1310>.
30. Li ML, et al. EV713C protease induces apoptosis by cleavage of hnRNP A1 to promote apaf-1 translation. *PLoS ONE*. 2019;14(9):e0221048. <https://doi.org/10.1371/journal.pone.0221048>.
31. Li J, et al. Enterovirus 713C promotes apoptosis through cleavage of PinX1, a telomere binding protein. *J Virol*. 2017. <https://doi.org/10.1128/jvi.02016-16>.
32. Sharma R, Raychaudhuri S, Dasgupta A. Nuclear entry of poliovirus protease- polymerase precursor 3CD: implications for host cell transcription shut-off. *Virology*. 2004;320(2):195–205. <https://doi.org/10.1016/j.virol.2003.10.020>.
33. Weng KF, Li ML, Hung CT, Shih SR. Enterovirus 713C protease cleaves a novel target CstF-64 and inhibits cellular polyadenylation. *PLoS Pathog*. 2009;5(9):e1000593. <https://doi.org/10.1371/journal.ppat.1000593>.
34. Zhang XN, Song ZG, Jiang T, Shi BS, Hu YW, Yuan ZH. Rupintrivir is a promising candidate for treating severe cases of Enterovirus-71 infection. *World J Gastroenterol*. 2010;16(2):201–9. <https://doi.org/10.3748/wjg.v16.i2.201>.
35. Kuo CJ, et al. Design, synthesis, and evaluation of 3C protease inhibitors as anti-enterovirus 71 agents. *Bioorgan Med Chem*. 2008;16(15):7388–98. <https://doi.org/10.1016/j.bmc.2008.06.015>.
36. Shang L, et al. Biochemical characterization of recombinant enterovirus 71 3C protease with fluorogenic model peptide substrates and development of a biochemical assay. *Antimicrob Agents Chemother*. 2015;59(4):1827–36. <https://doi.org/10.1128/AAC.04698-14>.
37. Hung HC, Wang HC, Shih SR, Teng IF, Tseng CP, Hsu JTA. Synergistic inhibition of enterovirus 71 replication by interferon and rupintrivir. *J Infect Dis*. 2011;203(12):1784–90. <https://doi.org/10.1093/infdis/jir174>.
38. Tan J, et al. 3C protease of enterovirus 68: structure-based design of Michael acceptor inhibitors and their broad-spectrum antiviral effects against picornaviruses. *J Virol*. 2013;87(8):4339–51. <https://doi.org/10.1128/jvi.01123-12>.
39. Tijsma A, et al. The capsid binder vapendavir and the novel protease inhibitor SG85 inhibit enterovirus 71 replication. *Antimicrob Agents Chemother*. 2014;58(11):6990–2. <https://doi.org/10.1128/AAC.03328-14>.
40. Y Z, et al. Cyanohydrin as an anchoring group for potent and selective inhibitors of enterovirus 713C protease. *J Med Chem*. 2015;58(23):9414–20. <https://doi.org/10.1021/acs.jmedchem.5b01013> LK-.
41. Ma Y, et al. 4-Lminooxazolidin-2-one as a Bioisostere of the cyanohydrin moiety: inhibitors of enterovirus 71 3C protease. *J Med Chem*. 2018;61(22):10333–9. <https://doi.org/10.1021/acs.jmedchem.8b01335>.
42. Zeng D, et al. Synthesis and structure-activity relationship of α -keto amides as enterovirus 713C protease inhibitors. *Bioorgan Med Chem Lett*. 2016;26(7):1762–6. <https://doi.org/10.1016/j.bmcl.2016.02.039>.
43. Ma GH, et al. Identification and biochemical characterization of DC07090 as a novel potent small molecule inhibitor against human enterovirus 71 3C protease by structure-based virtual screening. *Eur J Med Chem*. 2016;124:981–91. <https://doi.org/10.1016/j.ejmch.2016.10.019>.
44. Zakaryan H, Arabyan E, Oo A, Zandi K. Flavonoids: promising natural compounds against viral infections. *Arch Virol*. 2017;162(9):2539–51. <https://doi.org/10.1007/s00705-017-3417-y>.
45. Cao Z, et al. Luteoloside acts as 3C protease inhibitor of enterovirus 71 in vitro. *PLoS ONE*. 2016;11(2):e0148693. <https://doi.org/10.1371/journal.alpone.0148693>.
46. Yao C, et al. Inhibition of enterovirus 71 replication and viral 3C protease by quercetin. *Virol J*. 2018;15(1):116. <https://doi.org/10.1186/s12985-018-0123-6>.
47. Wang J, Zhang T, Du J, Cui S, Yang F, Jin Q. Anti-enterovirus 71 effects of chrysins and its phosphate ester. *PLoS ONE*. 2014;9(3):e89668. <https://doi.org/10.1371/journal.pone.0089668>.
48. Tan EL, Tan TMC, Chow VTK, Poh CL. Inhibition of enterovirus 71 in virus-infected mice by RNA interference. *Mol Ther*. 2007;15(11):1931–8. <https://doi.org/10.1038/sj.mt.6300287>.
49. Yang Z, Li G, Zhang Y, Liu X, Tien P. A novel minicircle vector based system for inhibiting the replication and gene expression of enterovirus 71 and Coxsackievirus A16. *Antivir Res*. 2012;96(2):234–44. <https://doi.org/10.1016/j.antiviral.2012.08.003>.
50. Subramanya S, Kim SS, Manjunath N, Shankar P. RNA interference-based therapeutics for human immunodeficiency virus HIV-I treatment: synthetic siRNA or vectorbased shRNA? *Expert Opin Biol Ther*. 2010;10(2):201–13. <https://doi.org/10.1517/14712590903448158>.
51. Ashfaq UA, Yousaf MZ, Aslam M, Ejaz R, Jahan S, Ullah O. SiRNAs: potential therapeutic agents against Hepatitis C Virus. *Virol J*. 2011;8:1–6. <https://doi.org/10.1186/1743-422X-8-276>.
52. Fujimoto Y, et al. Antiviral effects against influenza A virus infection by a short hairpin RNA targeting the non-coding terminal region of the viral nucleoprotein gene. *J Vet Med Sci*. 2019;81(3):383–8. <https://doi.org/10.1292/jvms.l8-0436>.
53. Wang W, Peng H, Li J, et al. Controllable inhibition of hepatitis B virus replication by a DR1-targeting short hairpin RNA (shRNA) expressed from a DOX-inducible lentiviral vector. *Virus Genes*. 2013;46:393–403. <https://doi.org/10.1007/s11262-013-0886-2>.

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