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Polysaccharide and extracts from *Lentinula edodes*: structural features and antiviral activity

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Abstract

Background: Lentinula edodes, known as shiitake, has been utilized as food, as well as, in popular medicine, moreover, compounds isolated from its mycelium and fruiting body have shown several therapeutic properties. The aim of this study was to determine the antiviral activity of aqueous (AqE) and ethanol (EtOHE) extracts and polysaccharide (LeP) from *Lentinula edodes* in the replication of poliovirus type 1 (PV-1) and bovine herpes virus type 1 (BoHV-1).

Methods: The time-of-addition assay was performed at the times -2, -1, 0, 1 and 2 h of the infection. The virucidal activity and the inhibition of viral adsorption were also evaluated. Plaque assay was used to monitor antiviral activity throughout.

Results: The AqE and LeP were more effective when added at 0 h of infection, however, EtOHE was more effective at the times 1 h and 2 h of the infection. AqE, EtOHE and LeP showed low virucidal activity, and the inhibition of viral adsorption was not significant.

Conclusions: The results allowed us to conclude that AqE, EtOHE and LeP act on the initial processes of the replication of both strains of virus.

Keywords: Lentinula edodes, Antiviral activity, Poliovirus, Bovine herpesvirus

Background

Currently, there are a little more than 40 drugs approved for clinical use in the treatment of viral infections [1]. Studies regarding these drugs concentrated on synthetic products up to the end of the 80 s, when natural compounds attracted attention because of their efficacy in the inhibition of viruses important for human and animal health [2]. Natural products are recognized by the pharmaceutical industry because of their wide structural diversity, as well as, variety of pharmacological activities [3]. Amongst the sources of natural compounds, the fungi, especially the basidiomycetes, have stimulated interest from investigators.

The basidiomycete *Lentinula edodes* (Berkeley) Pegler (*Lentinus edodes*), known as shiitake, an edible mushroom native of the East Asia, is valued for its nutritional

Poliovirus is a non-enveloped virus with an icosahedral capsid symmetry, and a genome consisting of a positive single-stranded RNA. The virion is classified in the genus *Enterovirus*, belonging to the family *Picornaviridae*, which includes many other pathogens of great importance to humans and other animals [15]. Despite the efforts to eradicate the virus, there were 1352 reported cases of poliomyelitis in African and Asian countries in 2010 [16]. Bovine herpesvirus (BoHV) is an

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and medicinal properties and culinary and industrial applications [4]. Shiitake is the second most popular in the world [5] and in the last decades various compounds with therapeutic properties have been isolated from its mycelium and fruiting body. Some of these have been widely studied, for example, the polysaccharide lentinan which has demonstrated high immunopotentiating and antimetastasic activities [6,7], antitumor activity [8,9], antibacterial, antifungal and antidiabetic activities [10,11], among others. However, few works have examined the antiviral activity of this basidiomycete [12-14].

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important pathogen for the cattle industry. Viron belongs to the subfamily *Alphaherpesvirinae*, family *Herpesviridae* [17], has a genome consisting of a linear double-stranded DNA within an icosahedral capsid, enclosed by an envelope. In order to reduce losses caused by BoHV, vaccines consisting of attenuated virus are being utilized with positive results. However, the control of infections is still difficult, due to the latency established by the virus after primary infection or after vaccination [18].

The basidiomycetes show various biological activities and low toxicity what make them a promising source of bioactive molecules. Therefore, the aim of this study was to determine the antiviral activity of aqueous and ethanol extracts and polysaccharide of *Lentinula edodes* in the replication of poliovirus and bovine herpesvirus.

Methods

Cells and virus

HEp-2 cell cultures (human larynx epithelial cells carcinoma–ATCC, CCL-23) were grown in Dulbecco's Modified Eagle Medium (DMEM) (Gibco-BRL, EUA*), supplemented with 10% fetal bovine serum (*) and 2 mM glutamine (Sigma Chem. Co., EUA**), 100 μg/ml streptomycin (**), 100 IU/ml penicillin (**) and 2.5 μg/ml amphotericin B (Bristol Myers-Squibb, Brazil).

The poliovirus type 1 (PV-1), vaccinal strain, was obtained from the ATCC (ATCC, VR-58) and the bovine herpesvirus type 1 (BoHV-1) was supplied by DMVP-UEL, Brazil. Both strains were propagated in HEp-2 cells, and virus titers determined by plaque assay.

Aqueous and ethanol extracts

The aqueous extract (AqE) of *Lentinula edodes* (lineage IW) was obtained as follows. Ground basideocarp was resuspended with distilled water, heated at 60° C for 1 h and centrifuged at $3000 \times g$ for 5 min. The supernatant was pre-filtered and submitted to ultrafiltration in 0.2 μ m pore size membrane, and stored at -20° C.

The ethanol extract (EtOHE) was prepared by dissolving ground basideocarp in 46% ethanol, at room temperature (\pm 25°C). The extract was centrifuged at 3,000 \times g for 5 min, and the supernatant was lyophilized. The lyophilized was resuspended in DMEM, submitted to ultrafiltration in 0.2 μ m pore size membrane, and stored at -20°C.

Polysaccharide extraction and purification

The polysaccharide from *Lentinula edodes* (LeP) was isolated as described by Gonzaga et al. [19]. Briefly, dried mushroom was dissolved at 5% (w/w) in distilled water at 100°C during 5 h. The suspension was centrifuged and clear colorless extract was neutralized to pH 7.0 with 0.1 N NaOH. One percent NaCl was added and the extract

submitted to polysaccharides precipitation with ethanol (5 vol ethanol:1 vol extract). After hydrogen peroxide/ ethanol treatment, precipitate was resubmitted to ethanol extraction. The precipitate washed with ethanol and acetone was dried at 40°C, dissolved in distilled water, clarified by centrifugation and lyophilized.

Cytotoxic assay

The cytotoxicity of the test substances was performed by dimethylthiazolyldiphenyl tetrazolium bromide (MTT) kit (**). HEp-2 cells were grown in 96-well microplates and treated with varying concentrations of AqE (0.1-100 mg/ml), EtOHE (0.1-40 mg/ml) and LeP (0.25-6.0 mg/ml). After 72 h incubation, the test was carried out according to the manufacturer's recommendation. Under the same conditions, cells without treatment were used as control. The 50% cytotoxic concentration (CC $_{50}$) was determined as the concentration capable of reducing the optical density by 50% in comparison with the control. The CC $_{50}$ was calculated by linear regression analysis of the dose-response curves generated.

Plaque reduction assay

The antiviral activity by plaque reduction assay was done according to Melo et al. [20] and used throughout. Briefly, HEp-2 cells were cultivated in 24-well plates at 37°C in 5% of CO_2 . After complete confluence, the cells were infected and treated with the substances accordingly. Cell cultures were overlaid with nutrient agarose and supplemented with antibiotic. For PV-1 experiments, nutrient agarose was added of 25 mM MgCl_2 . The plates were incubated inverted at 37°C in 5% CO₂, for 48 h. The cells were fixed with 20% formalin, stained with 0.5% crystal violet, after removal of the nutrient agarose layer. Concomitantly, mock-infected cells were used as control. The percentage of viral inhibition (% V.I.) was calculated by the formula: % V.I. = [1 - (number of plaques in test/ number of plaques in virus control)] \times 100.

The 50% inhibitory concentration (IC₅₀) was determined as the concentration capable of reducing 50% the number of plaques forming units (PFU) in relation to the controls. The IC₅₀ was determined by linear regression analysis of the curves of viral inhibition, for each treatment.

The selectivity index (SI) was calculated as the ratio of CC_{50} and IC_{50} .

Strains of PV-1 and BoHV-1 were submitted to the treatment with 1,000 U/ml and 10,000 U/ml human alfa-2 interferon (Meizler Com. Intern. SA, Brazil), respectively.

Virucidal activity

To evaluate the direct effect of the substances on viral particles, at varying concentrations, 10⁶ PFU/ml of PV-1

and 10^5 PFU/ml of BoHV-1 were mixed with equal volumes of AqE (3.1, 6.3, 12.5 and 25 mg/ml), EtOHE (0.375, 0.75, 1.5 and 3 mg/ml) and LeP (0.025, 0.05, 0.1 and 0.2 mg/ml) for 1 h at 37°C and inoculated in cell cultures.

Time-of-addition assay

The evaluation of the time-of-addition effect of the substances, at varying concentrations, was done as in Yang et al. [21]. Cells cultivated in 24-well plates were treated with concentrations of AqE (3.1, 6.3, 12.5 and 25 mg/ml), EtOHE (0.375, 0.75, 1.5 and 3 mg/ml) and LeP (0.025, 0.05, 0.1 and 0.2 mg/ml), before (-1 h and -2 h), during (0 h) and after (1 h and 2 h) infection.

Viral adsorption assay

The inhibition of the viral adsorption was carried out according to Zhu et al. [22]. Briefly, the cells cultivated in 24-well plates were infected with viral strains in the presence of the AqE (3.1, 6.3, 12.5 and 25 mg/ml), EtOHE (0.375, 0.75, 1.5 and 3 mg/ml) and LeP (0.025, 0.05, 0.1 and 0.2 mg/ml), and incubated at 4° C in 5% CO₂ for one h. The cells were washed twice with PBS, and plaque reduction assay was performed after 48 h.

Statistics

The data were analyzed by ANOVA followed by Student's t-test. Values were considered significant to $p \le 0.05$. All experiments were performed in triplicate.

Results

The polysaccharide isolated from *Lentinula edodes* contains high levels of β -D-glucan. The ^{13}C NMR spectrum is shown in Figure 1 and $\beta(1{\to}6)$ and $\alpha(1{\to}4)$ glucans configurations were identified in the spectrum. The chemical displacements characteristics of the carbons C1 to C6 of the glycosidic ring are presented in Table 1.

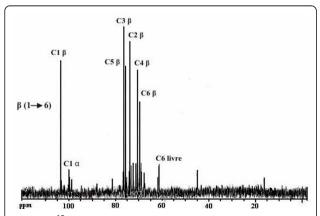


Figure 1 ¹³C NMR spectrum from polysaccharide isolated from *Lentinula edodes*.

Table 1 Chemical displacements characteristics of $\beta(1\rightarrow 6)$ and $\alpha(1\rightarrow 4)$ glucans present in the purified polysaccharides isolated from *Lentinula edodes*

Configurations	Chemical displacements (ppm)							
	C1	C2	C3	C4	C5	C6		
β(1→6)	103.5	73.7	75.6	70.4	75.6	69.7		
<u>α(1→4)</u>	99.9	72.4	76.4	81.3	71.1	61.6		

The cytotoxicity of the substances tested resulted in CC_{50} for the AqE, EtOHE and LeP of 74.0 mg/ml, 25.8 mg/ml and 4.0 mg/ml, respectively (Table 2).

The results of antiviral activity for the AqE at different times of infection for PV-1 are shown in Figure 2a. When the AqE was added, one or two hours before infection (-1 h and -2 h), at the highest concentration tested (25 mg/ ml), there was an inhibition of 5.8% and zero%, respectively. The addition of the extract at the concentrations of 3.1, 6.3, 12.5 and 25 mg/ml, at the moment of infection (0 h), resulted in a viral inhibition of 1.8, 17.5, 41.1 and 82.5%, respectively. However, for the time-of-addition of one hour post-infection (1 h), at the same concentrations, the percentages of inhibition were 9.2, 12.1, 24.5 and 60.2%, respectively. With the time-of-addition of two hours post-infection (2 h), the concentrations of 3.1 mg/ ml and 6.3 mg/ml were not effective, but, the concentrations of 12.5 mg/ml and 25 mg/ml inhibited the replication of PV-1 by 28.2 and 49.4%, respectively.

The results of the tests for virucidal activity and the inhibition of adsorption showed that the AqE inhibited the replication of PV-1 by 38.3% and 19.0%, respectively, at the highest concentration tested. Figure 2b shows the results of the antiviral activity of AqE in the replication of BoHV-1. When the cells were treated with a concentration of 12.5 mg/ml, the highest percentage of viral inhibition obtained was 19.1 and 4.2%, for the times -1 h and -2 h before the infection, respectively. At the time 0 h, for the concentrations of 3.1, 6.3, 12.5 and 25 mg/ml there was inhibition of 50.4, 43.3, 66.7 and 89.9%, respectively. However, for post-infection treatments, at the same concentrations, there was inhibition of 20.4,

Table 2 Antiviral activity of aqueous extract (AqE), ethanol extract (EtOHE) and polysaccharide (LeP) of *Lentinula edodes* for poliovirus and bovine herpesvirus, monitored by plaque assay

Substances	CC ₅₀ ^a	PV-1		BHV-1	
		IC ₅₀ ^b	SIc	IC ₅₀	SI
AqE	74.0	12.7	5.82	8.2	9.02
EtOHE	25.8	1.30	19.85	2.13	12.11
LeP	> 4.0	0.19	> 21.33	0.1	> 39.21

^a Fifty percent cytotoxic concentration (mg/ml)

^b Fifty percent inhibitory concentration (mg/ml)

 $^{^{\}mbox{\tiny c}}$ Selectivity index

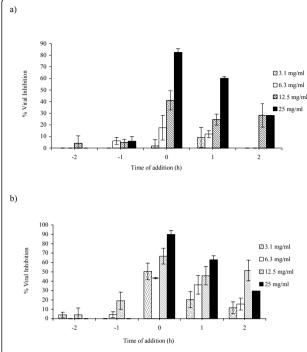


Figure 2 Effect of *Lentinula edodes* aqueous extract (AqE) on poliovirus (a) and bovine herpesvirus (b) replication, monitored by plaque reduction assay in HEp-2 cells. The extract was utilized in the indicated concentrations before (-1 and -2), during (0) and after infection (1 and 2). The experiments were carried out in triplicate, and the percent of the inhibition is represented with the respective standard deviations.

36.1, 45.8 and 63.0% for the time 1 h and 11.7, 15.5, 51.5 and 75.7% for the time 2 h.

For virucidal and inhibition of adsorption activities, there was a low inhibitory effect for the replication of BoHV-1 (32.1% and zero%, respectively) at the highest concentration of AqE.

The results of the EtOHE on the replication of PV-1 and BoHV-1 are shown in Figure 3. For PV-1, at concentrations of 0.375, 0.75, 1.5 and 3 mg/ml, the extract inhibited replication of the virus by zero, 45.7, 56.5 and 73.9%, respectively, at time 0 h of infection (Figure 3a). When the extract was added at the times 1 h and 2 h, at the indicated concentrations, the percentages of inhibition were 21.4, 24.8, 53.9 and 76.9%, and 20.7, 36.3, 70.8 and 94.6%, respectively. However, treatments at times -1 h and -2 h, showed no significant effect, with the highest inhibition of 21.7%. In the replication of BoHV-1 (Figure 3b), at the same concentrations, the extract showed inhibition of zero, 10.2, 32.4 and 70.8%, respectively, for the time 0 h. Treatments at times 1 h and 2 h resulted in inhibition of 35.4, 46.9, 56.3 and 90.6%, and, zero, 8.1, 22.3 and 47.6%, respectively. The extract did not show any virucidal activity or inhibition of viral adsorption either for both virus strains.

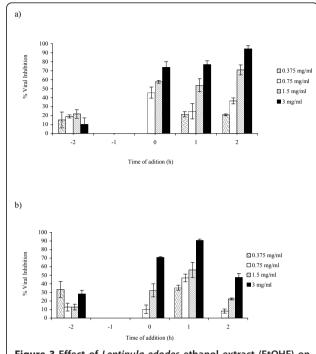


Figure 3 Effect of *Lentinula edodes* ethanol extract (EtOHE) on poliovirus (a) and bovine herpesvirus (b) replication, monitored by plaque reduction assay in HEp-2 cells. The extract was utilized in the indicated concentrations before (-1 and -2), during (0) and after infection (1 and 2). The experiments were carried out in triplicate, and the percent of the inhibition is represented with the respective standard deviations.

The antiviral activity of LeP on the replication of PV-1 and BoHV-1 are shown in Figure 4. For PV-1, at the concentrations of 0.025, 0.05, 0.1 and 0.2 mg/ml, LeP inhibited viral replication by 11.2, 21.0, 23.6 and 63.8%, respectively, at time 0 h of infection (Figure 4a). When the polysaccharide was added at time 1 h, at the same concentrations, the percentages of inhibition were zero, 4.2, 2.0 and 28.5%, respectively. However, for the treatments at the times 2 h, -1 h and -2 h, no effect was observed. In the replication of BoHV-1, at the same concentrations, LeP showed inhibition of 23.1, 29.6, 37.1 and 74.2%, respectively, for the time 0 h (Figure 4b). Treatments at the times 1 h and 2 h showed inhibition of 13.1, 17.9, 29.4 and 33.5%, and, zero, zero, 40.9 and 52.7%, respectively.

The LeP virucidal activity on the PV-1 replication, at the same concentrations, showed inhibition of 14.5, 24.4, 28.5 and 34.3%. However, LeP neither demonstrated virucidal activity for BoHV-1 nor inhibition of viral adsorption for both virus strains.

The IC_{50} and the respective SI for AqE, EtOHE and LeP, calculated for treatment at time 0 h of the infection, are shown in Table 2.

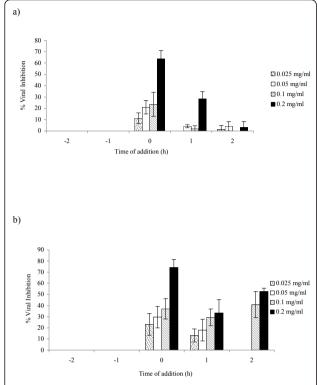


Figure 4 Effect of *Lentinula edodes* polysaccharide (LeP) on poliovirus (a) and bovine herpesvirus (b) replication, monitored by plaque reduction assay in HEp-2 cells. The extract was utilized in the indicated concentrations before (-1 and -2), during (0) and after infection (1 and 2). The experiments were carried out in triplicate, and the percent of the inhibition is represented with the respective standard deviations.

Interferon alpha-2, used as positive control, inhibited the replication of PV-1 and BoHV-1 by 100% at concentrations of 1,000 U/ml and 10,000 U/ml, respectively.

Discussion

In this work, the aqueous and ethanol extracts, and, polysaccharides of the fruiting body of *Lentinula edodes* were evaluated for antiviral activity.

The results demonstrated that both extracts, as well as, LeP, inhibited the replication of PV-1 and BoHV-1. AqE inhibited PV-1 and BoHV-1 in a dose-dependent curve and the highest percentages of inhibition were obtained at time 0 h of infection. No significant inhibitory effect was observed when the extract was added at -1 h and -2 h of the infection and for the inhibition of adsorption, for both virus. This may suggest that AqE did not influence significantly the specific binding of both virus to cell receptors. AqE did not show virucidal activity, and therefore, did not affect virus particle directly either. Sorimachi et al. [23] demonstrated that fractions of the aqueous extract of *Agaricus blazei* mycelium inhibited significantly the CPE of western

equine encephalitis (WEE) virus, herpes simplex virus and poliovirus in Vero cells. This effect was observed after infection demonstrating that extract of basidiomycetes may contain inhibitory compounds for the initial phases of replication at least for herpesvirus and enterovirus. This finding strengthens the activity that we demonstrated of AqE on PV-1 and BoHV-1 replication.

EtOHE was effective in inhibiting the replication of both viruses. The greatest percent of viral inhibition was demonstrated when the extract was added post-infection, 94.6% (2 h) and 90.6% (1 h) for PV-1 and BoHV-1, respectively. It is likely, therefore, that EtOHE acted on the initial steps of the replication, considering that AqE did not show virucidal activity or inhibition of viral adsorption either. Awadh Ali et al. [24] demonstrated the presence of antiviral activity in ethanol extracts of the fruiting body and mycelium of Inonotus hispidus against influenza virus A and B, attributing the effect to phenolic compounds. Concerning the relevance of glucan in biological activities it was demonstrated that LeP contains higher levels of β-D-glucan in comparison with an Agaricus blazei isolate [19]. In our study, the pronounced peak representative of anomeric carbon in β $(1\rightarrow 6)$ configuration is evidence of greater concentration of glucan in relation to that of the α configuration. Many studies disclose $\beta(1\rightarrow 3)$ configuration for the isolated glucan from Lentinus edodes [25,26], therefore, being different to the one disclosed in this study. Certainly, the characteristics of the cultivation region, such as climate and growth conditions, justify such behaviour. LeP, a β -glucan-protein with a predominance of β -1-6, also showed a dose-dependent inhibitory effect on BoHV-1 and PV-1, although less pronounced, nevertheless, with a higher SI and lower IC₅₀, compared to AqE and EtOHE extracts. The antiviral activity of extracts isolated from the basidiomycetes seems to be mostly attributed to the presence of polysaccharides. The anionic feature of the molecules can interfere with early stages of viral replication [27,28] and sulfated polysaccharides demonstrate higher antiviral activity on enveloped virus [29].

Conclusions

In conclusion, we suggest that AqE, EtOHE and LeP act on the initial processes of PV-1 and BoHV-1 replication. The extracts and the polysaccharide could be considered as a source of potential antiviral substances. However, further study is necessary to better understanding of the step(s) of viral replication where inhibition occurs.

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Authors' contributions

VPR participated in the studies on cytotoxicity and antiviral activity of the polysaccharide and extracts from *Lentinula edodes* and prepared the draft of the manuscript. KAY prepared the extracts and fractions and participated in the studies on antiviral activity. NMPSR and SAS participated in the extraction and purification of the polysaccharide, and participated in the manuscript. LDPM was responsible for shiitake cultivation and preparation of the extracts. RECL and CN were responsible for the design and coordination of the work, data analysis and manuscript writing and revision. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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