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The antigenic property of the H5N1 avian influenza viruses isolated in central China

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Abstract

Background: Three influenza pandemics outbroke in the last century accompanied the viral antigen shift and drift, resulting in the change of antigenic property and the low cross protective ability of the existed antibody to the newly emerged pandemic virus, and eventually the death of millions of people. The antigenic characterizations of the viruses isolated in central China in 2004 and 2006–2007 were investigated in the present study.

Results: Hemagglutinin inhibition assay and neutralization assay displayed differential antigenic characteristics of the viruses isolated in central China in two periods (2004 and 2006–2007). HA genes of the viruses mainly located in two branches in phylogeny analysis. 53 mutations of the deduced amino acids of the HA genes were divided into 4 patterns. Mutations in pattern 2 and 3 showed the main difference between viruses isolated in 2004 and 2006–2007. Meanwhile, most amino acids in pattern 2 and 3 located in the globular head of the HA protein, and some of the mutations evenly distributed at the epitope sites.

Conclusions: The study demonstrated that a major antigenic drift had occurred in the viruses isolated in central China. And monitoring the antigenic property should be the priority in preventing the potential pandemic of H5N1 avian influenza virus.

Keywords: Avian influenza virus, Antigenic epitope, Antigenic drift

Background

Three influenza pandemics in 20th century (1918 H1N1 Spanish, 1957 H2N2 Asian and 1968 H3N2 Hong Kong) and the first influenza pandemic in 21st century (H1N1/2009 Mexico) were due to the direct interspecies transmission or exchange of gene segments between avian, swine and human influenza viruses [1]. The newly emerged pandemic strains were antigenically divergent from seasonal influenza viruses circulating at that time. Vaccines efficient for the seasonal flu could not elicit any cross-reactivity in humans. Tens of thousands of people died in each pandemic due to the lack of effective cross-protection of existed antibody.

HA protein is the primary target of neutralizing antibodies and continuously accumulates mutations to escape recognition of the immune system. Alteration of

the antigenic epitopes of HA protein results in immune evasion and more rapid spread of influenza virus. The antigenic epitopes of H3 subtype influenza virus were well characterized and mapped to the three dimensional structure of the HA protein [2,3]. The epitopes of H5 avian influenza virus (AIV) were also identified through sequencing HA gene of the escape mutants selected by specific monoclonal antibodies (Mabs) [4-6]. Nearly all amino acids in epitopes located in the surface of the HA protein.

In our previous study, an H5N1 highly pathogenic AIV (HPAIV), A/duck/Hubei/hangmei01/2006 (hm/06), had been isolated from brains of dead laying ducks with severe central nervous system (CNS) dysfunction [7]. Subsequently, several HPAIV H5N1 viruses isolated from ducks and pigeons also showed neurovirulence in field ducks and pigeons. In view of the increasing virulence as well as mortality to the natural host, waterfowls, we try to elucidate whether the changed biological properties are related to the antigenicity of these H5N1 viruses isolated after 2005. Our previous study had identified the antigenicity of the viruses isolated in 2004 [8].

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The present study compared the antigenic features of the viruses isolated in 2004 and 2006–2007 in central China. Hemagglutination inhibition (HI) and neutralization assay (NT) activity, the phylogenetic tree and deduced amino acids of HA gene as well as the location of mutated sites in the HA protein crystal model were performed to reveal the molecular mechanism of the antigenic properties of the viruses isolated respectively from the two periods in central China.

Results

2.1 The HI activity of the Mabs to the 10 viruses

Before detecting the HI activity of the Mabs to the 10 viruses, western blot assays were used to identify the activity of the selected Mabs. The results displayed that all six Mabs could recognize the HA protein of virus dw/04 (figure 1). Then the Mabs were tested for their abilities to inhibit hemagglutination of chicken erythrocytes to the selected 10 viruses (table 1). Mab 2 C9 showed moderate HI activity to all the 10 viruses. But the other five Mabs displayed obviously weaker HI activity to the viruses isolated in 2006–2007 than those isolated in 2004. Mab 5E12 displayed relatively higher HI activity to all the 10 selected viruses, however, the difference in HI activity could be observed in the two periods (table 1).

2.2 The NT activity of the Mabs to the 10 viruses

The Mabs displayed different neutralizing efficiency to the viruses isolated from the two periods (table 1). Mab 2 C9 displayed extremely high neutralization activity to all the 10 viruses, which was inconsistent with the moderate HI activity. This incoordination might indicate that Mab 2 C9 did not bind to the receptor binding regions of the HA protein. The NT activity of the four Mabs (2 H4, 1 C4, 5E12 and 4 C12) is much higher to the 5 viruses isolated in 2004 than those isolated in 2006–2007. However, the Mab 2E11 displayed very low NT activity to 10 AIV H5N1 strains except the virus ewhc/04.

2.3 The phylogenetic analysis of the HA gene

Phylogenetic analysis was performed to elucidate the evolution situation of the viruses isolated in the two periods. The viruses mainly formed two different branches in the phylogenetic tree. And the viruses isolated in 2004 were closely related to the viruses isolated in

Vietnam. The HA clade except xfy/04 was 9, while the HA clade of xfy/04 was 7 (figure 2). The viruses isolated in 2006–2007 belonged to Fujian-like sublineage and the HA clade of hm/06 and d16/06 was 2.3.4 and the HA clade of zg/07, xg/07 and xn/07 was 2.3.2 (figure 2).

2.4 Analysis of the mutated amino acids in HA protein of the viruses

To further investigate the molecular foundation of the antigenic difference of the 10 viruses, amino acid mutations of HA protein were analyzed. A total of 53 amino acid substitutions appeared in the HA protein of the selected 10 viruses (table 2). According to the distribution of the mutated amino acids, the 53 substitutions could be divided into four different patterns. In pattern 1, the substitutions mainly happened in virus xfy/04, and the other 9 viruses were basically homologous; in pattern 2, substitutions happened in viruses xg/07, xn/07 and zg/07, which were isolated in 2007; in pattern 3, substitutions happened in all the 5 viruses isolated in 2006–2007; and in pattern 4, substitutions happened sporadically in all the viruses (table 2).

2.5 Distributions of the mutated amino acids in the HA crystal structure

The precise sites of the mutated amino acids were mapped to the crystal structure of AIV HA molecule, 3GBM. The model displayed that 36 of the 53 mutations located in the globular head of HA1 proteins. And it could find that amino acid sites 119, 120, 138, 139, 140, 141, 161 and 162 were located at site A epitope of the HA protein; amino acid sites 124, 129, 151, 154, 155, 156, 184, 188 and 189 were located at site B epitope; amino acid sites 40, 45, 277 and 282 at site C epitope; and amino acids 94, 226, 227 at site D epitope; amino acids 84, 263, 269 at site E epitope (figure 3).

Discussion

The H5N1 AIVs continuously outbreak in East Asia countries after 2005 and form a new phylogenetic clade, gaining some new characterizations, such as neurovirulence and the ability to infect poultry and migratory birds [7,9]. Meanwhile, most of the subsequent viruses isolated from the infected human in China prove to be clade 2.3 [10]. H5N1 HP AIVs cause a constant increase

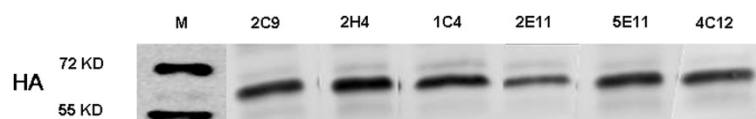


Figure 1 Reactivity of the Mabs to the HA protein of virus dw/04. Due to the different Mabs were used to perform the western blot, the six reactions were performed separately and the image was resembled together. Same HA protein loading and exposure time were used in the western blot assay.

Table 1 Comparison of antigen city of the 10 viruses in NT and HI assays using 6 HA protein Mabs

H5N1 strains	2 C9		2 H4		1 C4		5E12		2E11		4 C12	
	NT	HI	NT	HI	NT	HI	NT	HI	NT	HI	NT	HI
zfe/04	30720	128	9486	1024	30720	16384	30720	16384	378	128	11943	64
dw/04	35481	32	12800	512	30720	16384	30720	8192	672	64	6400	64
xfy/04	102400	32	32359	512	102400	16384	102400	16384	724	64	27542	32
xfj/04	74131	256	436	64	102400	16384	102400	16384	977	64	31623	64
ewhc/04	102400	<16	45709	256	102400	16384	102400	16384	12800	32	4467	32
hm/06	30720	64	192	256	156	256	1503	512	<60	<16	<60	<16
d16/06	30720	<16	168	128	2799	1024	13095	1024	<60	<16	405	<16
xn/07	28840	32	<200	<16	<200	64	2754	256	<200	<16	<200	<16
zg/07	27542	<16	<200	<16	1738	1024	16596	4096	<200	<16	457	<16
xg/07	20283	256	<60	<16	<60	32	600	512	<60	<16	<60	<16

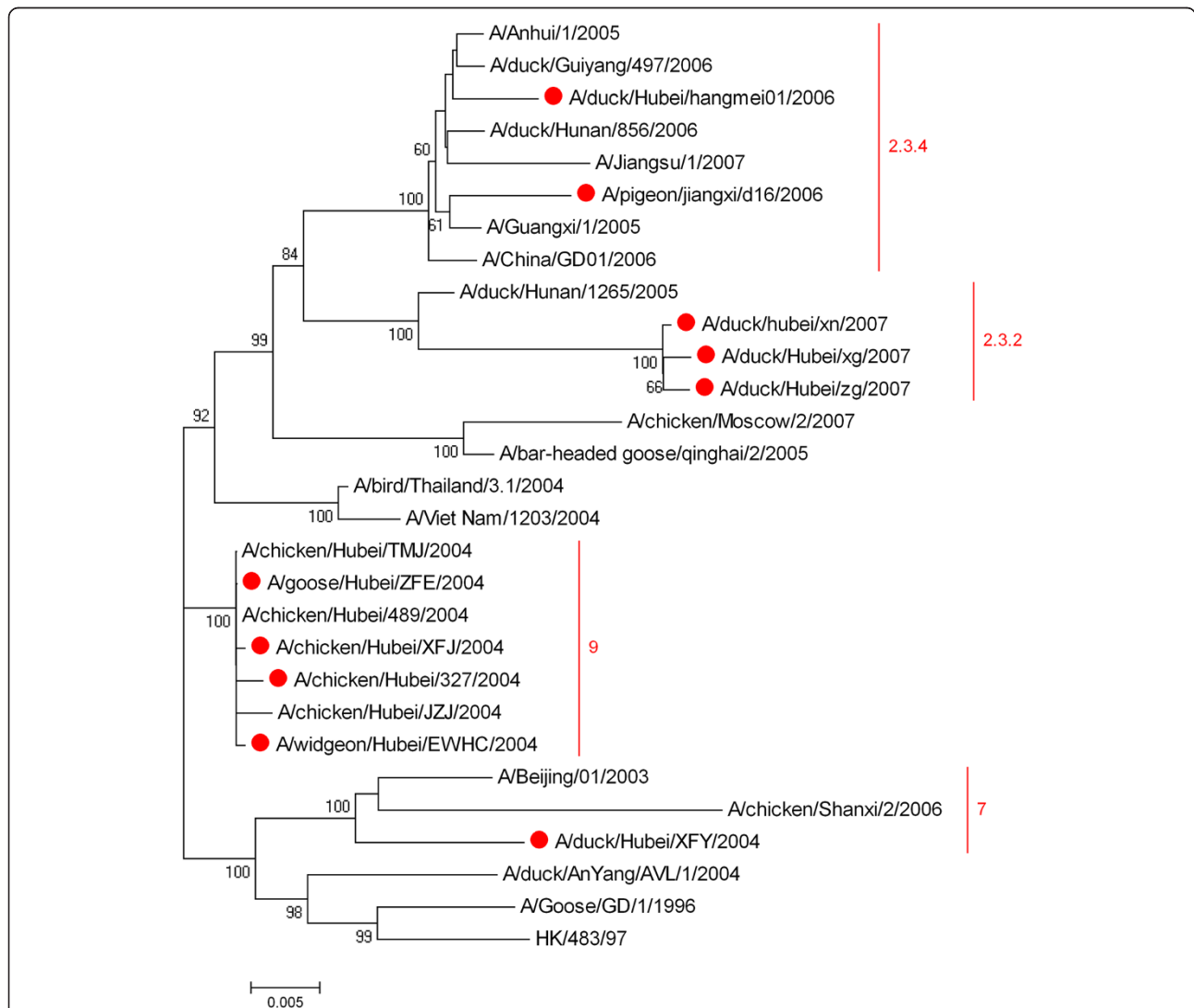


Figure 2 Phylogenetic analysis of HA gene of the 10 viruses. The phylogenetic tree was build with the program MEGA 4.1. Clades of H5N1 virus were classified according to nomenclature recommended by WHO evolution working group. The viruses used in present study were labeled by red spot.

Table 2 Mutated amino acid sites of the deduced HA protein of the 10 viruses

sites	zfe	dw	xfy	xfj	ewhc	hm	d16	xn	zg	xg ^{a)}	note ^{b)}
21	E	E	<u>V</u> ^{c)}	E	E	E	E	E	E	E	Pattern 1
114	I	I	<u>V</u>	I	I	I	I	I	I	I	
119	K	K	<u>R</u>	K	K	K	K	K	K	K	
188	T	T	<u>I</u>	T	T	T	T	T	T	T	
189	R	R	<u>K</u>	R	R	R	K	R	R	R	
282	M	M	<u>V</u>	M	M	<i>I</i>	<i>I</i>	M	M	M	
320	S	S	<u>A</u>	S	S	S	S	S	S	S	
325	R	R	<u>G</u>	R	R	R	R	R	R	R	
418	F	F	<u>S</u>	F	F	F	F	F	F	F	
450	D	D	<u>E</u>	D	D	D	D	D	D	D	
483	R	R	<u>K</u>	R	R	R	R	R	R	R	
499	K	K	<u>N</u>	K	K	K	K	K	K	K	
511	I	I	<u>M</u>	I	I	I	M	I	I	I	
513	T	T	<u>I</u>	T	T	T	T	T	T	T	
2	Q	Q	Q	Q	Q	Q	Q	H	H	H	Pattern 2
40	K	K	K	K	K	K	K	R	R	R	
45	D	D	D	D	D	D	D	N	N	N	
53	R	R	K	R	R	R	R	K	K	K	
120	S	S	S	S	S	S	S	D	D	D	
129	S	S	S	S	S	S	S	L	L	L	
154	N	N	N	N	N	N	N	D	D	D	
156	T	T	T	T	T	T	T	A	A	A	
162	R	R	R	R	R	R	R	K	K	K	
184	A	A	A	A	A	A	A	E	E	E	
200	V	V	V	V	V	V	V	I	I	I	
226	M	M	M	M	T	M	M	I	I	I	
263	A	A	A	A	A	A	A	T	T	T	
277	K	K	K	K	K	K	K	R	R	R	
528	A	A	A	A	A	A	A	V	V	V	
533	V	V	V	V	V	V	V	M	M	M	
84	S	S	S	S	S	N	N	N	N	N	Pattern 3
94	D	D	D	D	D	N	N	N	N	N	
124	N	N	N	N	N	D	D	D	G	D	
138	L	L	L	L	L	Q	Q	Q	Q	Q	
155	S	S	S	S	S	N	N	N	N	N	
227	E	E	E	E	E	D	D	D	D	D	
269	L	L	L	L	L	V	V	V	V	V	
310	R	R	R	R	R	K	K	K	K	K	
35	K	K	K	K	K	K	K	R	K	K	Pattern 4
139	G	<i>R</i>	G	G	G	G	G	G	G	G	
140	K	K	R	K	K	<i>T</i>	<i>T</i>	N	N	N	
141	S	S	<i>P</i>	S	S	<i>P</i>	<i>P</i>	S	S	S	
151	I	I	I	I	I	I	I	I	I	T	

Table 2 Mutated amino acid sites of the deduced HA protein of the 10 viruses (Continued)

161	K	K	K	K	K	<i>R</i>	<i>K</i>	<i>K</i>	<i>K</i>	<i>K</i>
174	V	V	V	V	V	<i>I</i>	<i>I</i>	<i>V</i>	<i>V</i>	<i>V</i>
181	P	P	P	P	P	<i>S</i>	<i>S</i>	<i>P</i>	<i>P</i>	<i>P</i>
209	L	<i>F</i>	L	L	L	<i>L</i>	<i>L</i>	<i>L</i>	<i>L</i>	<i>L</i>
240	N	<i>D</i>	N	N	N	<i>N</i>	<i>N</i>	<i>N</i>	<i>N</i>	<i>N</i>
313	L	L	L	L	L	<i>P</i>	<i>L</i>	<i>L</i>	<i>L</i>	<i>L</i>
322	Q	Q	Q	Q	Q	<i>L</i>	<i>L</i>	<i>Q</i>	<i>Q</i>	<i>Q</i>
354	Y	Y	Y	Y	Y	<i>Y</i>	<i>F</i>	<i>Y</i>	<i>Y</i>	<i>Y</i>
366	A	A	A	T	A	<i>A</i>	<i>A</i>	<i>A</i>	<i>A</i>	<i>A</i>
467	C	C	C	C	C	<i>R</i>	<i>C</i>	<i>C</i>	<i>C</i>	<i>C</i>

Note:

a): abbreviations of the 10 viruses: zfe/04, dw/04, xfy/04, x fj/04, ewhc/04, hm/06, d16/06, xn/07, zg/07 and xg/07.

b): the four patterns mutated amino acids according to mutated situation showed in the text.

c): residues in bold plus italic plus underline were mainly pattern 1 mutation happened in virus xfy/04; residues in bold plus italic were mainly pattern 2 and 3 mutations happened in viruses isolated in 2006–2007; residues in italic were sporadic mutations in the 10 viruses.

of human infection cases and nearly 60% of the infected people die (http://www.who.int/influenza/human_animal_interface/H5N1_cumulative_table_archives/en/index.html). Meanwhile, H5N1 HP AIVs are raging all over the world in poultry as well as migrated birds and putting significant threat to human health and poultry breeding.

Further analysis reveal that the HA gene of these viruses belong to Clade 2.3. The properties of the sequences, virulence and even the transmission property of the viruses in this clade are investigated profoundly in many reports [7,11-15]. Wu et.al show that viruses isolated from 2002 to 2007 have formed four distinct antigenic groups, and imply that a major antigenic drift might happen in the viruses isolated after 2005 (clade 2.3) [16]. But the antigenicity of the viruses in this clade is not fully studied. Investigation about the antigenicity of H5N1 AIVs isolated in different time and geographical areas after 2005 is beneficial to clarifying the changed biological properties of the emerged viruses.

In the present study, combining the HI and the NT activity of the Mabs to the 10 selected viruses isolated in central China, we conclude that a significant antigenic drift have occurred in the viruses isolated in the two periods. To reveal the molecular mechanism of the antigenic change, phylogenic analysis is performed and displays that HA genes of the viruses isolated in two periods obviously locate in different branches. The deduced amino acids demonstrate that the 53 substitutions in the HA protein of the 10 viruses are divided into four patterns. The pattern 1 mutations are only observed in virus xfy/04, the same sites in other 9 viruses are identical. Our previous study had demonstrated that the virus

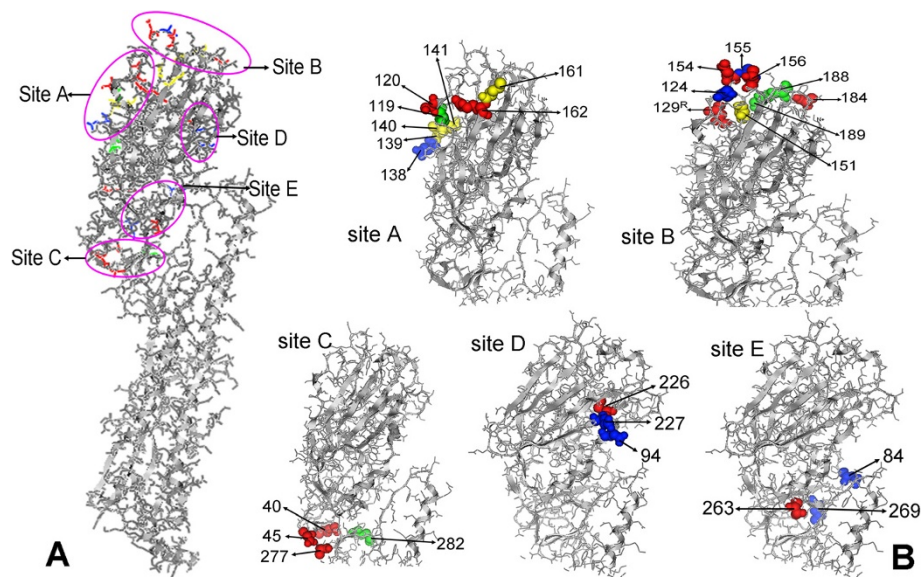


Figure 3 Mutated sites in the crystal structure of HA protein. The crystal model is HA protein of A/Vietnam/1203/2005, built by the program PyMOL program (V1.4). **A** is the intact model of HA protein displayed the mutated amino acids in the five antigenic epitope sites. **B** is the detail profile of the five antigenic epitope sites A, B, C, D and E. The mutated amino acid sites in the 10 avian influenza viruses HA proteins located in the five epitopes were labeled. Amino acids labeled with green were pattern 1 mutation; amino acids labeled with red were pattern 2 mutation; amino acids labeled with blue were pattern 3 mutation; amino acids labeled with orange were pattern 4 mutation (table 2).

xfy/04 showed low pathogenicity in mice, chickens and pigeons, while the other 9 viruses showed high pathogenicity in animal experiments or in field [7,17]. Therefore, pattern 1 mutations might be related to the low pathogenicity of virus xfy/04.

The amino acids in pattern 3 mutations are different in viruses isolated in 2004 and 2006–2007, indicating that this pattern mutation might happen from 2005 to 2006. However, the mutations in pattern 2 are only observed in the viruses isolated in 2007, indicating that this pattern mutation might happen from 2006 to 2007. This information reveals that the antigenicity of H5N1 AIV is constantly changing in the process of spread. On the other hand, viruses isolated in 2006 (hm/06 and d16/06) belong to clade 2.3.4, while viruses isolated in 2007 (xn/07, zg/07 and xg/07) belong to clade 2.3.2. Meanwhile, viruses in clade 2.3.2 have gradually replaced the viruses in clade 2.3.4 in these years and become the most prevalent viruses in China. This might suggest that viruses undergoing antigenicity alteration have a great advantage in prevalence and spread.

Conclusions

The present study reveals that the antigenic epitopes of the HA protein have changed in the prevalent AIVs in central China. The mutations in pattern 2 and 3 might be responsible for the differences of HI and NT activity and the distinct antigenic features of the viruses isolated

in the two periods. Particularly, most mutations in pattern 2 and 3 locate in the globular head and distribute at the five epitope sites, and even represent the trend of antigenic drift in 2004–2007. This phenomenon might demonstrate that those sites actually expose to the selected pressure and easily cause antigenic drift. Generally, the H5N1 AIVs are undergoing further antigenic drift and persistent monitoring the epidemiology and antigenic evolution of H5N1 AIV is imperative.

Materials and Methods

4.1 Virus propagation, purification and titration

The H5N1 AIV used in this study included: A/Chicken/Hubei/327/2004 (dw/04), A/Goose/Hubei/ZFE/2004 (zfe/04), A/Chicken/Hubei/XFJ/2004 (xfj/04), A/Duck/Hubei/XFY/2004 (xfy/04), A/widgeon/Hubei/EWHC/2004(ewhc/04), A/duck/Hubei/hangmei01/2006 (hm/06), A/pigeon/jiangxi/d16/2006 (d16/06), A/duck/Hubei/xn/2007 (xn/07), A/duck/Hubei/zg/2007 (zg/07), and A/duck/Hubei/xg/2007 (xg/07). All H5N1 strains were propagated in 9-day-old SPF embryonated eggs. Allantoic fluids were harvested and stored at -80°C . TCID₅₀ titrations of all ten AIV isolates were performed according to Reed and Muench in MDCK cultures [18].

4.2 Production of monoclonal antibodies against dw/04

The panel of Mabs (2 C9, 2 H4, 1 C4, 5E12, 2E11 and 4 C12) against HA protein of dw/04 was stored at -80°C

in our lab. The protocol of the Mab production had been previously described [8]. The antigenic characterization and neutralization assay of the viruses isolated in 2004 had been characterized in previous study [8].

4.3 Western blot assays

The Mabs activities were analyzed by western blot assay with HA protein of dw/04. 5 µg purified HA protein was dissolved in the loading buffer and applied onto the gels. Following the gel electrophoresis, the separated proteins were transferred to nitrocellulose membrane (Boster, China) using Trans-Biorad SD (Bio-Rad, USA). Each membrane was washed once with TBS-T buffer for 10 mins and blocked with 5% skimmed milk at 4°C overnight and then incubated with the Mab at 37°C for 1 h. Finally, after triple washes, the membrane was incubated with horseradish peroxidase-conjugated anti-mouse IgG (Boster, China) at 37°C for 1 h and visualized with ECL western-blotting detection system (Amersham, USA).

4.4 HI assay

HI assay of the Mabs were performed as described previously, using chicken erythrocytes for agglutination and four hemagglutination units of the selected 10 viruses [19].

4.5 Virus neutralization assays

Two-fold serial dilutions of the Mabs were mixed with the virus suspensions (100 TCID₅₀ per well) and incubated together for 1 h at 37°C. Then 100µL mixture was added onto washed Madin-Darby canine kidney (MDCK) cell in a flat bottom 96-well cell culture plate (Corning Incorporated Costar, USA). Cytopathogenic effect (CPE) was observed after a 3-day incubation at 37°C in the presence of 5% CO₂. Neutralizing titers were expressed as the highest dilution of the virus which could result in 50% neutralization of the CPE.

4.6 Phylogenetic analysis of the HA genes of the viruses

Total RNA was extracted from the allantoic fluid using Rneasy mini kit (Qiagen, California, USA), and reverse transcribed into cDNA. All the HA genes of selected viruses were amplified by the specific HA oligonucleotide primers. RT-PCR and sequencing of the HA genes of the viruses were repeated to confirm that the sequence data were identical. The other HA gene sequences for phylogenetic analysis were obtained from the NCBI Database. Sequences of HA gene were edited and aligned by ClustalX (Version1.81) software and phylogenetic tree was produced by MEGA 4.0.2 software.

4.7 Analysis of the mutated amino acid sites in the HA crystal model

The deduced amino acids were aligned to further analyzed the mutated sites of the HA protein of the 10 viruses (DNASar, Madison, WI). The crystal structure data of HA protein of the virus A/Vietnam/1203/2004 was download from PDB data bank (<http://www.rcsb.org/pdb/home/home.do>) and the PDB ID was 3GBM. All the mutated amino acids in HA proteins of the 10 viruses were mapped to the crystal model of protein 3GBM with the program PyMOL program (V1.4). The 5 antigenic sites of H5N1 AIV referenced previous reports or blasted the HA genes to the H3 subtype AIV whose epitopes had mapped to the crystal model of HA protein [2-5,8,16,20-22].

Abbreviations

AIV: avian influenza virus; CPE: cytopathogenic effect; HI: hemagglutination inhibition; HPAIV: highly pathogenic avian influenza virus; Mab: monoclonal antibody; MDCK: Madin-Darby canine kidney; NT: neutralization assay; TCID50: 50% tissue culture infection dose.

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

W Z and M J designed the experiments; W Z, J K and J Z performed the experiments; J K, J Z, H Z sequenced the HA genes of the viruses, performed the phylogenetic analysis and contributed the reagents and materials; W Z and M J wrote the manuscript. All authors read and approved the final manuscript.

Acknowledgment

This work was supported by National 973 program (2011CB505004, 2010CB534002), National S&T major project (2009ZX10004-109). We thanked Professor Yanxiu Liu of Foreign Language College, Huazhong Agriculture University for her help in English.

Received: 21 January 2012 Accepted: 25 July 2012

Published: 6 August 2012

Reference

1. Xu R, Ekiert DC, Krause JC, Hai R, Crowe JE Jr, Wilson IA: **Structural basis of preexisting immunity to the 2009 H1N1 pandemic influenza virus.** *Science* 2010, **328**:357-360.
2. Wiley DC, Wilson IA, Skehel JJ: **Structural identification of the antibody-binding sites of Hong Kong influenza haemagglutinin and their involvement in antigenic variation.** *Nature* 1981, **289**:373-378.
3. Wilson IA, Skehel JJ, Wiley DC: **Structure of the haemagglutinin membrane glycoprotein of influenza virus at 3 Å resolution.** *Nature* 1981, **289**:366-373.
4. Kaverin NV, Rudneva IA, Ilyushina NA, Varich NL, Lipatov AS, Smirnov YA, Govorkova EA, Gitelman AK, Lvov DK, Webster RG: **Structure of antigenic sites on the haemagglutinin molecule of H5 avian influenza virus and phenotypic variation of escape mutants.** *J Gen Virol* 2002, **83**:2497-2505.
5. Philpott M, Easterday BC, Hinshaw VS: **Neutralizing epitopes of the H5 hemagglutinin from a virulent avian influenza virus and their relationship to pathogenicity.** *J Virol* 1989, **63**:3453-3458.
6. Philpott M, Hioe C, Sheerar M, Hinshaw VS: **Hemagglutinin mutations related to attenuation and altered cell tropism of a virulent avian influenza A virus.** *J Virol* 1990, **64**:2941-2947.
7. Zou W, Yu Z, Zhou H, Tu J, Jin M: **Genetic characterization of an H5N1 avian influenza virus with neurovirulence in ducks.** *Virus Genes* 2009, **38**:263-268.
8. Huang H, Dan H, Zhou Y, Yu Z, Fan H, Tong T, Jin M, Chen H: **Different neutralization efficiency of neutralizing monoclonal antibodies against**

- avian influenza H5N1 virus to virus strains from different hosts. *Mol Immunol* 2007, **44**:1052–1055.
9. Sturm-Ramirez KM, Ellis T, Bousfield B, Bissett L, Dyrting K, Rehg JE, Poon L, Guan Y, Peiris M, Webster RG: Reemerging H5N1 influenza viruses in Hong Kong in 2002 are highly pathogenic to ducks. *J Virol* 2004, **78**:4892–4901.
 10. Shu Y, Yu H, Li D: Lethal avian influenza A (H5N1) infection in a pregnant woman in Anhui Province, China. *N Engl J Med* 2006, **354**:1421–1422.
 11. Neumann G, Chen H, Gao GF, Shu Y, Kawaoka Y: H5N1 influenza viruses: outbreaks and biological properties. *Cell Res* 2010, **20**:51–61.
 12. Li Y, Shi J, Zhong G, Deng G, Tian G, Ge J, Zeng X, Song J, Zhao D, Liu L, et al: Continued evolution of H5N1 influenza viruses in wild birds, domestic poultry, and humans in China from 2004 to 2009. *J Virol* 2010, **84**:8389–8397.
 13. Smith GJ, Vijaykrishna D, Ellis TM, Dyrting KC, Leung YH, Bahl J, Wong CW, Kai H, Chow MK, Duan L, et al: Characterization of avian influenza viruses A (H5N1) from wild birds, Hong Kong, 2004–2008. *Emerg Infect Dis* 2009, **15**:402–407.
 14. Yen HL, Lipatov AS, Ilyushina NA, Govorkova EA, Franks J, Yilmaz N, Douglas A, Hay A, Krauss S, Rehg JE, et al: Inefficient transmission of H5N1 influenza viruses in a ferret contact model. *J Virol* 2007, **81**:6890–6898.
 15. Smith GJ, Fan XH, Wang J, Li KS, Qin K, Zhang JX, Vijaykrishna D, Cheung CL, Huang K, Rayner JM, et al: Emergence and predominance of an H5N1 influenza variant in China. *Proc Natl Acad Sci U S A* 2006, **103**:16936–16941.
 16. Wu WL, Chen Y, Wang P, Song W, Lau SY, Rayner JM, Smith GJ, Webster RG, Peiris JS, Lin T, et al: Antigenic profile of avian H5N1 viruses in Asia from 2002 to 2007. *J Virol* 2008, **82**:1798–1807.
 17. Yu Z, Song Y, Zhou H, Xu X, Hu Q, Wu H, Zhang A, Zhou Y, Chen J, Dan H, et al: Avian influenza (H5N1) virus in waterfowl and chickens, central China. *Emerg Infect Dis* 2007, **13**:772–775.
 18. Zou W, Ke J, Zhang A, Zhou M, Liao Y, Zhu J, Zhou H, Tu J, Chen H, Jin M: Proteomics analysis of differential expression of chicken brain tissue proteins in response to the neurovirulent H5N1 avian influenza virus infection. *J Proteome Res* 2010, **9**:3789–3798.
 19. Masurel N, Ophof P, de Jong P: Antibody response to immunization with influenza A/USSR/77 (H1N1) virus in young individuals primed or unprimed for A/New Jersey/76 (H1N1) virus. *J Hyg (Lond)* 1981, **87**:201–209.
 20. Chen Y, Qin K, Wu WL, Li G, Zhang J, Du H, Ng MH, Shih JW, Peiris JS, Guan Y, et al: Broad cross-protection against H5N1 avian influenza virus infection by means of monoclonal antibodies that map to conserved viral epitopes. *J Infect Dis* 2009, **199**:49–58.
 21. Kaverin NV, Rudneva IA, Govorkova EA, Timofeeva TA, Shilov AA, Kochergin-Nikitsky KS, Krylov PS, Webster RG: Epitope mapping of the hemagglutinin molecule of a highly pathogenic H5N1 influenza virus by using monoclonal antibodies. *J Virol* 2007, **81**:12911–12917.
 22. Rudneva IA, Kushch AA, Masalova OV, Timofeeva TA, Klimova RR, Shilov AA, Ignatieva AV, Krylov PS, Kaverin NV: Antigenic epitopes in the hemagglutinin of Qinghai-type influenza H5N1 virus. *Viral Immunol* 2010, **23**:181–187.

doi:10.1186/1743-422X-9-148

Cite this article as: Zou et al.: The antigenic property of the H5N1 avian influenza viruses isolated in central China. *Virology Journal* 2012 **9**:148.

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