SHORT REPORT



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Sequence analysis of fusion protein gene of Newcastle disease virus isolated from outbreaks in Egypt during 2006

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Abstract

Background: Newcastle disease virus represents APMV-1 and is the most characterized among all APMV types. The F protein cleavage site sequence is a well-characterized determinant of NDV pathogenicity in chickens. In this study, the sequences of fusion protein (F) gene of three Newcastle disease virus (NDV) strains isolated from outbreak in chickens in the Al-Sharkia province of Egypt in 2006 were determined.

Findings: The viral genomic RNAs were extracted from the infective allantoic fluid and F gene is amplified using primer sets designed from the available sequences of NDV strains from GenBank. The pathogenicity of NDV strains was determined by three internationally recognized tests mean death time, intracerebral pathogenicity index, and intravenous pathogenicity index. The phylogenetic analysis showed that the Egypt isolates are closely related with the genotype II of class II NDV strains.

Conclusions: The sequences of the F genes of the 2006 Egypt isolates are closely related to that of the 2005 Egypt isolate from the same province suggesting that these strains are probably circulating in the vaccinated bird population in Egypt until development of an outbreak.

Keywords: Newcastle disease virus, African strain, Chickens, Sequence

Findings

Newcastle disease (ND) is a highly contagious and fatal disease of chickens. In many developing countries ND is endemic and the disease has the greatest impact on villages where the livelihood of people depends on poultry farming. The causative agent Newcastle disease virus (NDV), is a member of the genus *Avulavirus* in the family *Paramyxoviridae*. NDV isolates display a spectrum of virulence in chickens ranging from inapparent to fatal infection. Based on their pathogenicity in chickens, NDV isolates are categorized into three main pathotypes: lentogenic (low virulence), mesogenic (intermediate virulence) and velogenic (high virulence). It may appear that continents having warm climates are reservoirs of virulent NDV strains [1]. NDV is a well-studied paramyxovirus and complete genome sequences



ND outbreaks occur frequently in Egypt and the source of the virulent NDV in these outbreaks are not known. We have reported the complete genome sequence of a NDV strain isolated from an outbreak at a poultry farm in Al-Sharkia province, Egypt in 2005 (chicken/Egypt/1/2005) [5]. In this study, we have determined the fusion (F) gene sequences of three NDV strains isolated from outbreaks on poultry farms in Al-Sharkia province, Egypt in 2006. The infected birds showed severe neurological and/or respiratory symptoms. The viruses were confirmed as NDV by hemagglutination inhibition assay using a known NDV antiserum. The pathogenicity of NDV strains was determined by three internationally recognized tests mean death time (MDT), intracerebral pathogenicity index (ICPI), and intravenous pathogenicity index (IVPI) [6].



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Table 1 Pathogenicity index tests of Egyptian strains of NDV.

Strains	IVPI	ICPI	MDT	
NDV/chicken/Egypt/2/2006	2.1 ± 0.24	1.6 ± 0.15	60 ± 1.5	
NDV/chicken/Egypt/3/2006	2.2 ± 0.29	1.8 ± 0.19	50 ± 1.8	
NDV/chicken/Egypt/4/2006	2.25 ± 0.23	1.7 ± 0.18	55 ± 2.3	
NDV/LaSota	ND	0.0	102 ± 2.5	

Intravenous pathogenicity index (IVPI) was done in 6-week-old chickens, intracerebral pathogenicity index (ICPI) test in 1-day-old chicks inoculated intracerebrally and mean death time (MDT) in 9-day-old eggs inoculated through allantoic cavity following standard protocol.

The NDV strains were designated as NDV/chicken/ Egypt/2/2006, NDV/chicken/Egypt/3/2006 and NDV/ chicken/Egypt/4/2006. NDV strain LaSota was used as a positive control for all pathogenicity studies. The results of the pathogenicity tests showed that all three NDV strains were velogenic (Table 1).

The F gene was chosen for sequencing because this gene is a major determinant of virulence and NDV isolates are grouped into genotypes based on the sequences of this gene [7-9]. To determine the F gene sequence of all the Egyptian strains, the viruses were grown in the allantoic cavities of 9-day-old embryonated specific pathogen free (SPF) chicken eggs. Plaque purification of the viruses was not performed to avoid the genetic selection, which may not represent the actual viral genome. The viral genomic RNAs were extracted from the infective allantoic fluid using RNeasy mini kit (QIA-GEN). Primer sets were designed from the available sequences of NDV strains Beaudette C (accession numbers X04719) and Texas GB (accession numbers GU978777). Three primers namely F-1for 5' ACGGG-TAGAAGATTCTG 3', F-655for 5'GTTGACTAAGT-TAGGTG 3'and F-1780rev 5' CTCTCCGAATTG-ACAGAC 3' (number corresponds to NDV F gene nucleotide sequence) were used to sequence the entire F gene. Reverse transcription and PCR were done using virus specific primers by Superscript-II reverse transcriptase and high fidelity Platinum Pfx polymerase (both from Invitrogen), respectively. The PCR-amplified products were directly sequenced using BigDye terminator v 3.1 matrix standard kit and 3130*xl* genetic analyzer data collection software v3.0 (Applied Biosystems Inc). The entire F gene was sequenced at least three times from independent RNA preparations to ensure a consensus sequence. All experiments were carried out in an enhanced BSL3 containment facility certified by the USDA to work with highly virulent NDV strains with the investigators wearing appropriate protective equipment and compliant with all protocols approved by the Institutional Animal Care and Use Committee (IACUC) of the University of Maryland and under Animal Welfare Association (AWA) regulations.

The F gene of all three Egyptian strains of NDV is 1792 nt in length and encodes a predicted F protein of 553 amino acids (GenBank under accession numbers FJ969393, FJ969394 and FJ969395). Sequence analysis of the F protein of Egyptian strains showed 94% to 99% amino acid identity with that of strain BC (Table 2). Phylogenetic analysis showed that strains chicken/Egypt/ 2/2006 and chicken/Egypt/4/2006 are closely related with the virulent NDV strain Texas GB, while strain chicken/Egypt/3/2006 stands out from other NDV strains in genotype II of class II viruses (Figure 1). The F protein cleavage site sequence is a well-characterized determinant of NDV pathogenicity in chickens [8-10]. Virulent NDV strains typically contain a polybasic cleavage site $(R-X-K/R-R\downarrow F)$, which is recognized by intracellular proteases present in most cell types. The cleavage site of all Egyptian strains contained four basic amino acids at positions 112-116 (112 R-R-Q-K-R \downarrow F-I 118), corresponding to those of virulent NDV strains similar to the one that was isolated in 2005 from the same province [5]. In addition, the presence of the phenylalanine (F) residue at position 117 has been described as being a possible contributor to the neurological effects [11,12]. Interestingly, the F cleavage site of NDV strains isolated from Egypt was identical to those of NDV strains isolated from chickens and guinea fowl in the Mopti and Sikasso regions of Mali in 2008 and many other NDV strains from China [5,13,14].

Our results indicate that high levels of nucleotide and amino acid sequence identity exist between the African

Table 2 Percent amino acid sequence identity of the Egyptian strains with other strains of NDV.

NDV strains	Egypt/2/2006	Egypt/3/2006	Egypt/4/2006	Egypt/1/2005	LaSota	BC	Fontana
Egypt/2/2006		94.4	98.9	98.4	97.5	99.1	90.4
Egypt/3/2006			94.0	93.3	93.1	93.9	92.6
Egypt/4/2006				97.6	98.2	98.0	90.1
Egypt/1/2005					97.5	99.1	90.8
LaSota						97.8	90.4
BC							91.3
Fontana							

DNA pair-wise alignment was done using MegAlign (clustalW) in a Lasergene 8 software package.



and North American NDV strains. It is possible that NDV strains isolated from Egypt may represent the strains circulating in the poultry population in that region. These results also suggest that more than one genotype of NDV is circulating in the African subcontinents. However, determination of the complete genome sequences of Egyptian strains isolated in different geographic regions of Africa is necessary to understand the genetic relatedness among NDV strains circulating in different parts of the world.

The close genetic relatedness of the NDV strains isolated in 2005 and 2006 in the same province in Egypt suggest that these strains are endemic in the bird population in this province. Furthermore, these results suggest that NDV vaccine used in this province may not be very effective in stopping viral shedding, which will allow unnoticed circulation of the virulent virus in the vaccinated bird population until development of an outbreak. Therefore, it may be necessary to evaluate the effectiveness of the current vaccine used in the Al-Sharkia province against circulating NDV strains.

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Authors' contributions

Conceived and designed the experiments: MHM and SKS. Performed the experiments: MHM, SK and AP. Analyzed the data: MHM, SK and AP. Contributed reagents/materials/analysis tools: MHM and SKS. Wrote the paper: MHM, SK and SKS. All the authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

- Herczeg J, Wehmann E, Bragg RR, Travassos Dias PM, Hadjiev G, Werner O, Lomniczi B: Two novel genetic groups (VIIb and VIII) responsible for recent Newcastle disease outbreaks in Southern Africa, one (VIIb) of which reached Southern Europe. Arch Virol 1999, 144:2087-2099.
- Snoeck CJ, Ducatez MF, Owoade AA, Faleke OO, Alkali BR, Tahita MC, Tarnagda Z, Ouedraogo JB, Maikano I, Mbah PO, et al: Newcastle disease virus in West Africa: new virulent strains identified in non-commercial farms. Arch Virol 2009, 154:47-54.
- Maminiaina OF, Gil P, Briand FX, Albina E, Keita D, Andriamanivo HR, Chevalier V, Lancelot R, Martinez D, Rakotondravao R, et al: Newcastle disease virus in Madagascar: identification of an original genotype possibly deriving from a died out ancestor of genotype IV. PLoS One 2010, 5:e13987.
- Cattoli G, Fusaro A, Monne I, Molia S, Le Menach A, Maregeya B, Nchare A, Bangana I, Maina AG, Koffi JN, *et al*: Emergence of a new genetic lineage of Newcastle disease virus in West and Central Africa–implications for diagnosis and control. *Vet Microbiol* 2010, 142:168-176.
- Mohamed MH, Kumar S, Paldurai A, Megahed MM, Ghanem IA, Lebdah MA, Samal SK: Complete genome sequence of a virulent Newcastle disease virus isolated from an outbreak in chickens in Egypt. Virus Genes 2009, 39:234-7.
- Alexander DJ: Newcastle, disease: OIE Terrestrial Manual 2009. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2009, Chapter 2:3.
- Aldous EW, Mynn JK, Banks J, Alexander DJ: A molecular epidemiological study of avian paramyxovirus type 1 (Newcastle disease virus) isolates by phylogenetic analysis of a partial nucleotide sequence of the fusion protein gene. Avian Pathol 2003, 32:239-256.
- Panda A, Huang Z, Elankumaran S, Rockemann DD, Samal SK: Role of fusion protein cleavage site in the virulence of Newcastle disease virus. *Microb Pathog* 2004, 36:1-10.
- de Leeuw OS, Hartog L, Koch G, Peeters BP: Effect of fusion protein cleavage site mutations on virulence of Newcastle disease virus: nonvirulent cleavage site mutants revert to virulence after one passage in chicken brain. J Gen Virol 2003, 84:475-484.
- Li Z, Sergel T, Razvi E, Morrison T: Effect of cleavage mutants on syncytium formation directed by the wild-type fusion protein of Newcastle disease virus. J Virol 1998, 72:3789-3795.
- Lamb R, Parks G: Paramyxoviridae: the viruses and their replication. In Fields Virology.. 5 edition. Edited by: Knipe DM, Howley PM, Griffin DE, Lamb RA, Martin MA, Roizman B, Straus SE. Philadelphia: Lippincott Williams 2007:1449-1496.
- 12. Kattenbelt JA, Stevens MP, Gould AR: Sequence variation in the Newcastle disease virus genome. *Virus Res* 2006, 116:168-184.
- Servan de Almeida R, Maminiaina OF, Gil P, Hammoumi S, Molia S, Chevalier V, Koko M, Andriamanivo HR, Traore A, Samake K, et al: Africa, a reservoir of new virulent strains of Newcastle disease virus? *Vaccine* 2009, 27:3127-3129.
- Abolnik C, Horner RF, Bisschop SP, Parker ME, Romito M, Viljoen GJ: A phylogenetic study of South African Newcastle disease virus strains isolated between 1990 and 2002 suggests epidemiological origins in the Far East. Arch Virol 2004, 149:603-619.

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