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Entire genome sequence analysis of genotype IX Newcastle disease viruses reveals their early-genotype phylogenetic position and recent-genotype genome size

Xusheng Qiu^{1,2}, Qing Sun¹, Shuang Wu¹, Li Dong¹, Shunling Hu¹, Chunchun Meng², Yantao Wu¹, Xiufan Liu^{1*}

Abstract

Background: Six nucleotide (nt) insertion in the 5'-noncoding region (NCR) of the nucleoprotein (NP) gene of Newcastle disease virus (NDV) is considered to be a genetic marker for recent genotypes of NDV, which emerged after 1960. However, F48-like NDVs from China, identified a 6-nt insert in the NP gene, have been previously classified into genotype III or genotype IX.

Results: In order to clarify their phylogenetic position and explore the origin of NDVs with the 6-nt insert and its significance in NDV evolution, we determined the entire genome sequences of five F48-like viruses isolated in China between 1946 and 2002 by RT-PCR amplification of overlapping fragments of full-length genome and rapid amplification of cDNA ends. All the five NDV isolates shared the same genome size of 15,192-nt with the recent genotype V-VIII viruses whereas they had the highest homology with early genotype III and IV isolates.

Conclusions: The unique characteristic of the genome size and phylogenetic position of F48-like viruses warrants placing them in a separate geno-group, genotype IX. Results in this study also suggest that genotype IX viruses most likely originate from a genotype III virus by insertion of a 6-nt motif in the 5'-NCR of the NP gene which had occurred as early as in 1940 s, and might be the common origin of genotype V-VIII viruses.

Background

Newcastle disease (ND) is one of the most serious infectious diseases of birds causing major economic losses in poultry industry[1-3]. Its causative agent, virulent Newcastle disease virus (NDV), belongs to the genus *Avulavirus*, in the subfamily *Paramyxovirinae*, family *Paramyxoviridae*, order *Mononegavirales*[4,5]. NDVs have a negative-sense, single-stranded continuous RNA genome about 15,186-nt, 15,192-nt or 15,198-nt in length [6-8] that contains six genes in the order of 3'-NP-P-M-F-HN-L-5', encoding six viral proteins (nucleoprotein, phosphoprotein, matrix protein, fusion protein, haemagglutinin-neuraminidase and large protein, respectively)[9]. Two additional proteins, V and W, are

expressed by mRNAs derived from the P gene via RNA editing [10,11].

Phylogenetically, NDVs have been classified into two major divisions, class I and class II [8,12]. Class I NDVs with the genome size of 15,198-nt are occasionally isolated from wild aquatic birds and domestic poultry and all but one of them are avirulent [8,13-16]. Class II viruses include most virulent and some avirulent NDVs: genotypes I-IV viruses are early lineage before 1960 with the genome size of 15,186-nt; whereas genotypes V-VIII are recent lineage after 1960 with the genome size of 15,192-nt [4,7,8,17,18]. Genotype I of class II contains mainly avirulent isolates from wild waterfowl and poultry species of the world; genotype II consists of North American isolates, which display different virulence ranging from lentogenic, mesogenic to velogenic; genotypes III and IV viruses represent early isolates from the Far East and Europe respectively during the first pandemic from mid 1920 s to late 1950 s; NDV strains isolated

* Correspondence: xfliu@yzu.edu.cn

¹Key Laboratory of Animal Infectious Diseases, Yangzhou University, Yangzhou 225009, PR China

Full list of author information is available at the end of the article

from the second pandemic during 1960 s and 1970 s belong to new genotypes V and VI; subtype VIb viruses are responsible for the third pandemic of pigeon origin during the 1980 s; novel genotypes of VIII and VII (many subgenotypes) which result in the fourth and latest pandemic have emerged since late 1980 s in the Far East, Europe, and South Africa [8,19-21].

NDV strain F48 ("F48E8" or "F48E9" was used in previous publications in which E8 or E9 means the 8th or 9th egg-passage of the original virus) was isolated from a diseased chicken in Northern China in 1946 and has been used as standard challenge strain for vaccine evaluation in this country [21-23]. The phylogenetic grouping of F48-like viruses is controversial in the literature: genotype IX of class II by some researchers [14-16,21,23,24] while genotype III by others for their highest homology of F gene with genotype III viruses [8,25,26]. At all events, it is evident that genotype IX is a sister clade of genotype III isolates which emerged in 1930 s. On the other hand, F48-like viruses have the 6-nt insert in the 5'-NCR of NP gene, which is considered to be a genetic marker of NDV strains emerged after 1960 [7,8]. However, the full-length genome of F48-like

NDVs has not been determined. In order to clarify the phylogenetic position of F48-like viruses and explore the origin of NDVs with 6-nt insert and its significance in NDV evolution, five F48-like viruses isolated in China between 1946 and 2002 were characterized and sequenced.

Results

Analysis of genome size

To determine the exact genome size of F48-like NDV isolates, the full-length genome sequences were compiled from sequences of nine overlapping cDNA fragments along with the sequences of the GC-rich region of NP gene and both ends of the genomes. Those sequences were submitted to GenBank and the accession number was FJ436302 - FJ436306. The results of sequencing displayed that these F48-like NDVs carried 6-nt insert in the 5'-NCR of the NP gene, the same as that of genotypes V-VIII NDVs which emerged after 1960 s (see Figure 1). Besides, no other insert or deletion was found when compared with all known NDV isolates. Therefore, the genome size of all the five genotype IX isolates was 15,192-nt, just as predicted before.



Figure 1 Alignment of the 3'-terminal non-coding sequences of the NP gene in the region of 6 nt insertion. Sequences obtained from the current study are underlined. The position of gaps were filled with '-'. The insertion site was framed.

Moreover, those 5 viruses isolated during 1948-2002 shared 99% nucleotide sequence identity of their genomes and the same 6-nt insert motif CCCCCC.

Phylogenetic analysis

Phylogenetic analysis of the five F48-like NDV strains together with NDVs representing the established genotypes was first performed using the variable region sequences (nt 47-420) of the F gene (Figure 2). The tree consisted of two major divisions, class I and class II, the latter was further divided into two lineages, early and recent. The early lineage included five genotypes (I to IV and IX) while the recent lineage consisted of four genotypes (V to VIII). It is obvious that F48-like strains (genotype IX) were close to but diverged from the early genotypes III and IV strains, forming a separate subclade.

Table 1 shows the range of F gene sequence similarity of NDV strains within one genotype and between different genotypes. The sequence similarity of F gene between genotype IX and III was the highest, ranging from 91.2% to 94.3%. The sequence similarity of other genes between genotype IX and III NDVs was also the highest when compared with those between genotype IX and other genotypes (data not shown).

Indeed, no matter which gene was used, the phylogenetic trees indicated very similar relationship of genetic groups. Figure 3 is the phylogenetic tree based on the entire genome sequences of the five F48-like NDVs in this study and those of other NDVs representing genotypes I through VIII which are available from the GenBank. The phylogenetic position of the F48-like NDVs here is consistent with the tree in Figure 2. Genotype IX strains were also clustered into early lineage, closely related with but diverged from genotypes III and IV strains.

GC content of the genomic sequences

The GC content of the sequences of Newcastle disease virus is also an important molecular characteristic. In table 2, we calculated the GC content of different region from 25 strains of NDV, including the entire genome, 6 complete viral genes, and also the 5' NCR of NP gene in which extra 6 nt were detected. It was noted that the GC content of full-length genome of all the 25 NDV strains were similar, however, the GC content of 5' NCR of NP gene showed significant difference. The 5' NCR of NP gene of genotype IV-IX NDV strains displayed more than 60% GC content, while that of genotype I and II strains showed about 53% and 55% GC content. Interestingly, the GC content of Genotype III strains in the same region was about 58%, higher than genotype I and II NDVs but lower than genotype IV-IX NDVs.

Molecular characterization of F protein

All the genotype IX strains in this study as well as other genotype IX isolates whose F gene sequences are available in the Genbank displayed the F protein cleavage site motif as 112RRQRR↓F117, the same as that of genotype III-IV strains (Figure 4). This finding is coincident with the biological characteristics of F48, which is used as the standard challenge strain in China (ICPI, 1.99). Moreover, F protein of genotype IX NDVs also had six potential N-glycosylation sites which were highly conserved among NDV isolates. The transmembrane (TM) and cytoplasmic regions of genotype IX NDVs contained several conserved substitutions and a non-conserved N for D substitution at residue 545.

Molecular characterization of HN protein

NDV strains of different genotypes show differences in the size of the HN protein which is the major determinant for virulence. The HN protein of all genotype IX strains is 571 amino acids long, the same size as HN of genotypes III-VIII viruses. The HN proteins of genotype IX strains contained all the six sites N-linked potential glycosylation sites at position 119, 341, 433, 481, 508, and 538 [27,28]. In addition, the HN of genotype IX NDVs contained positions E401, R41 and Y526 associated with receptor binding, and residues R174, R416 and R498 involving in NA activity [29-31].

Alignment of untranslated region of NDV genome

Figure 5 shows the alignment of the leader (A) and trailer (B) sequences of genotype IX NDVs with those of other genotype strains. Genotype IX NDVs contained the same gene-start (GS) signal and gene-end (GE) signal which are highly conservative for all NDV strains. Besides, the NP-P intergenic region of only one nucleotide was G in most genotype I-II NDV strains, whereas it was A in genotype IX NDV strains as well as genotype III-VIII strains. Several unique nucleotide substitutions were found in trailer region of genotype IX viruses, for example, C15095, C15107, C15125, and C15151 (Figure 5B).

Discussion

The outbreaks of the genotype V-VIII NDVs were still an enigma in the history of NDV evolution. Where did those viruses come from? Did they evolve directly from genotype I, II or III? The genetic character that obviously differentiating those viruses from early genotypes was the six nucleotide (nt) insertion in the 5'-non-coding region (NCR) of the nucleoprotein (NP) gene.

In this study, the 5 viruses isolated from 1948 to 2002 displayed the genome size of 15,192 nt due to the same 6-nt insert CCCCCC in the NP gene. However, those NDV strains shared high identity with genotype III, and

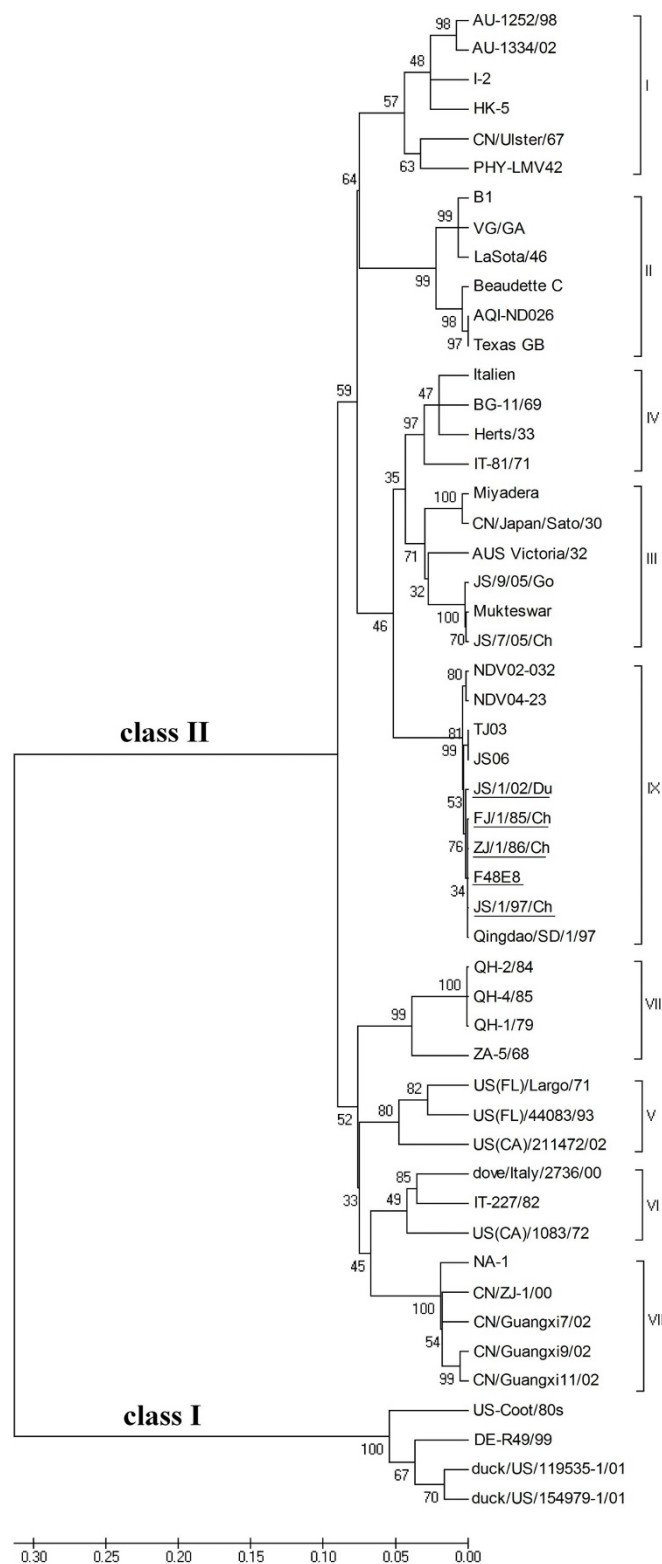


Figure 2 Phylogenetic tree of NDV strains. Tree construction was done using the Neighbor Joining method with the maximum composite likelihood substitution model for partial F gene (nt 47-420) by program MEGA version 4 (Tamura, Dudley, Nei, and Kumar 2007). Divisions and genotypes are indicated by roman numerals. Sequences obtained from the current study are underlined.

Table 1 Range of F gene nucleotide sequence similarity (%) of NDV strains within one genotype and between different genotypes

Genotype (No. of sequences)	I	II	III	IV	V	VI	VII	VIII	IX
I(23)	>91.8	87.0-91.8	88.6-93.3	86.2-92.5	81.9-86.3	83.4-87.9	82.6-92.2	85.8-89.7	89.3-91.7
II(32)	-	>89.1	87.1-91.2	85.1-90.0	81.2-85.3	82.3-86.4	80.9-90.1	84.8-88.0	88.5-89.9
III(6)	-	-	>92.4	87.8-94.6	83.5-88.3	83.7-90.1	83.2-89.1	87.1-91.6	91.2-94.3
IV(5)	-	-	-	>92.2	83.6-89.5	84.8-91.3	84.1-89.6	86.7-92.2	88.6-92.9
V(11)	-	-	-	-	>89.2	84.4-90.9	82.7-88.5	86.7-90.8	84.4-87.4
VI(16)	-	-	-	-	-	>91.4	84.8-92.4	88.4-92.4	84.8-88.7
VII(45)	-	-	-	-	-	-	>84.6	86.3-91.2	84.8-87.7
VIII(2)	-	-	-	-	-	-	-	>93.1	87.6-90.1
IX(7)	-	-	-	-	-	-	-	-	>99.3

The nucleotide sequence similarity of NDV strains were calculated by the MegAlign program in the Lasergene package (DNASTAR Inc. Madison, WI 53715, USA). 30 sequences of full-length genome together with more than one hundred sequences of complete F gene were involved.

were obviously clustered in a sister clade of genotype III in phylogenetic trees (see Figure 2, 3). It is well known that genotype III is a typical “early” NDV geno-group, while 6-nt insert of NP is characteristic of “recent” genotypes[8]. That is to say, two contradictive genetic features were both identified in F48-like NDV strains, suggesting the transitional role for those viruses in NDV evolution.

It is reasonable to infer that genotype IX viruses most likely originate from an early genotype III virus by the insertion of a 6-nt motif in the 5'-NCR of the NP gene, and recent V-VIII genotypes may come from genotype IX viruses, or evolve directly from genotype III or IV viruses in the same way. There are several evidence can be provided to support this hypothesis.

Firstly, it was noteworthy that a classic genotype III strain Australia-Victoria/32 shared the highest sequence similarity with F48 strains. The nucleotide sequence identity of the F, HN and L gene of Australia Victoria/32 (AV/32) with F48 strains was 94.3%, 93.7% and 94.4% respectively. In previous studies, early lineage viruses AV/32 (genotype III) and Herts/33 (genotype IV) have been positioned as the possible progenitor of recent virulent strains according to the sequence and phylogenetic analysis based on the HN, M-F, M, P and L gene sequences respectively and also their early date of isolation [32-36]. Results in this study indicated that F48-like viruses (genotype IX) invariably shared the closest homology with AV/32 viruses and displayed the genome size of 15192-nt.

Secondly, F48 which was isolated from an ND outbreak in Northern China in 1946 [22] is the earliest NDV isolate known to have the 6-nt insert in the 5'-NCR of NP gene, which suggested that NDV strains with 15,192-nt genome size emerged as early as 1940 s, rather than 1960 s when genotype V strains came out.

The recent genotypes V-VIII strains, as early as they were first isolated, have displayed wide genetic distances

and geographical distributions, which is indicative of a long period of evolution prior to the emergence of the recent viruses. On the other hand, the insertion of a 6-nt motif is an rare event in NDV evolution in view of the extremely low probability of nucleotide addition or deletion in the genome RNA of Paramyxoviruses [37,38]. Thus, it was most possible that genotype IX NDV was the common origin of genotype V-VIII.

Thirdly, it is noteworthy that all the recent viruses are virulent and their HN protein is exclusively 571 amino acids long, suggesting that their common progenitor must possess those genetic characteristics. As described below, genotype IX displayed the F protein cleavage site motif of ¹¹²RRQRR↓F¹¹⁷ and a HN protein of 571 amino acids.

At last, the GC content of the full-length and partial genome of Newcastle disease virus was compared in this study (table 2). It was noted that most region of genomic sequences of all class II NDV strains shared similar GC content, however, the 5' NCR of NP gene, where the 6-nt insert was found, showed significant difference in GC content. All the viruses with 15,192-nt genome displayed more than 60% GC content, while that of most 15,186-nt genome strains were no than 50% in this region. Interestingly, F48-like viruses showed high GC content in the 5' NCR of NP gene, the same as that of recent genotypes, suggesting the relationship between genotype IX and IV-VIII.

Moreover, the significance of 6-nt insert in NP gene for NDV has never been explored. A phenomenon has been observed in this study: genotype IX viruses were isolated ranging from 1940 s to 2000 s; In comparison, the early genotype III viruses such as AV/32 from Australia, Miyadera/51 and Sato/30 from Japan and Mukteswar from India are prevalent in Australia and Asia during the first pandemic of ND before 1960 but no longer detected thereafter with the exception of Mukteswar which is used as a vaccine virus in some Asian

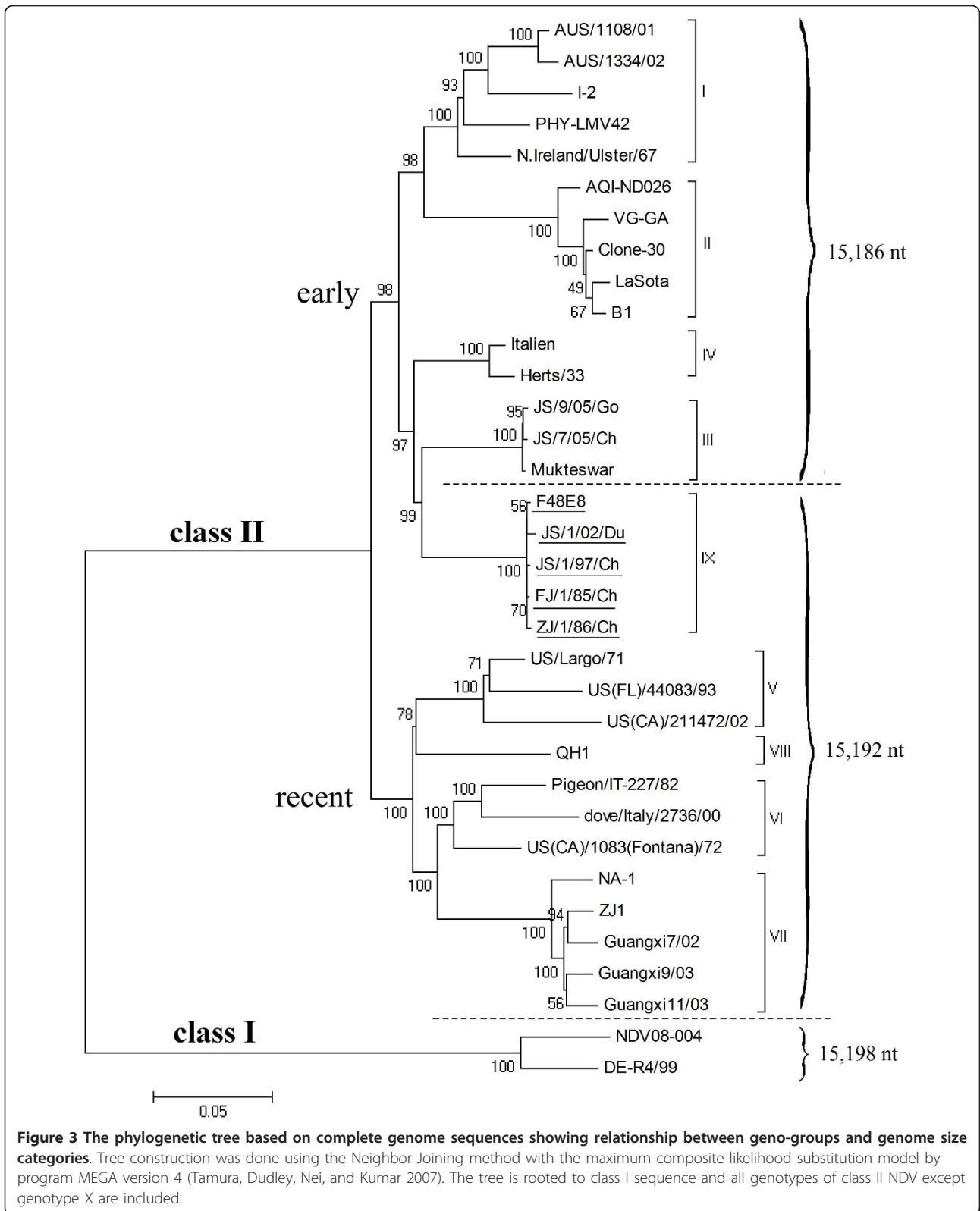


Figure 3 The phylogenetic tree based on complete genome sequences showing relationship between geno-groups and genome size categories. Tree construction was done using the Neighbor Joining method with the maximum composite likelihood substitution model by program MEGA version 4 (Tamura, Dudley, Nei, and Kumar 2007). The tree is rooted to class I sequence and all genotypes of class II NDV except genotype X are included.

Table 2 The GC contents in different regions of NDV genomes from 25 strains representing different genotypes

Genotype (No. of sequences)	Genome	NP gene	5'UTR of NP	P gene	M gene	F gene	HN gene	L gene
I(3)	46.56 ± 0.16	52.31 ± 0.28	55.55 ± 0.29	52.54 ± 0.84	49.05 ± 0.33	45.27 ± 0.09	46.91 ± 0.19	43.97 ± 0.37
II(3)	46.23 ± 0.05	52.25 ± 0.13	52.90 ± 0.28	52.58 ± 0.28	47.89 ± 0.16	44.82 ± 0.08	46.56 ± 0.03	43.77 ± 0.04
III(3)	46.58 ± 0.03	52.27 ± 0.00	58.21 ± 0.50	52.74 ± 0.07	48.59 ± 0.18	45.04 ± 0.03	47.51 ± 0.08	44.05 ± 0.02
IV(3)	46.68 ± 0.13	52.35 ± 0.35	62.85 ± 0.76	52.49 ± 0.62	49.30 ± 0.60	45.19 ± 0.13	46.32 ± 0.69	44.50 ± 0.15
IX(3)	46.85 ± 0.04	52.80 ± 0.03	62.16 ± 0.28	51.71 ± 0.13	49.48 ± 0.05	44.59 ± 0.03	48.09 ± 0.09	44.45 ± 0.06
V(3)	46.24 ± 0.30	52.12 ± 0.13	64.58 ± 2.48	52.46 ± 0.66	47.62 ± 0.17	44.95 ± 0.46	46.35 ± 0.86	44.04 ± 0.36
VI(3)	46.12 ± 0.34	51.66 ± 0.78	60.39 ± 0.84	53.04 ± 0.67	47.16 ± 0.59	45.40 ± 0.26	45.65 ± 0.33	43.80 ± 0.52
VII(3)	46.61 ± 0.13	51.45 ± 0.46	62.48 ± 2.83	53.60 ± 0.36	48.51 ± 0.20	45.12 ± 0.34	46.49 ± 0.76	44.45 ± 0.07
VIII(1)	46.29	51.92	61.06	53.64	47.32	44.84	46.29	43.94

The GC content of the full-length genome as well as the 6 viral genes was calculated by the EditSeq program in the Lasergene package (DNASTAR Inc. Madison, WI 53715, USA). Data are shown as the average of each genotype ± standard deviation.

countries[39]. Therefore, two different kinds of early genotypes NDVs, one with the genome size of 15,186-nt and the other with the genome size of 15,192 nt, exist simultaneously between 1940 s and 1960, while the former one is the predominant. In contrast, the recent genotypes V-IX viruses with genome size of 15,192-nt have emerged and predominated while early genotypes III and IV viruses have disappeared since 1960 s. It seems that NDVs with the insertion of 6-nt motif in the 5'NCR of NP gene might gain certain survive advantage in selective pressure of the ever-changing ecology.

Conclusion

Results in this study indicated that F48-like viruses are transitional class II NDVs with early-genotype phylogenetic position and recent-genotype genome size of 15,192-nt, which makes them to be a separate group, genotype IX; genotype IX viruses most likely originate from an early genotype III virus by the insertion of a 6-nt motif in the 5'-NCR of the NP gene, and recent V-VIII genotypes may come from genotype IX viruses, or evolve directly from genotype III or IV viruses in the same way; and this insertion is an important event in NDV evolution which had occurred as early as in 1940 s.

Materials and methods

Viruses

NDV strains for entire genome sequence analysis in this study are as follows: F48E8 (the 8th egg-passaged stock of F48) was isolated from chicken outbreak in Northern China in 1946 [22]; FJ/1/85/Ch, ZJ/1/86/Ch, and JS/1/97/Ch were isolated from chickens in Eastern China in 1985, 1986 and 1997 respectively; strain JS/1/02/Du was isolated from a healthy duck in our laboratory in 2002 [21]. All the five strains have been characterized as virulent NDVs and assigned to genotype IX previously (detailed data see Table 3). They were grown in 10-day-old embryonated specific-pathogen-free (SPF) chicken eggs and the allantoic fluids were harvested and stored in -70°C until use.

Preparation of viral RNA and RT-PCR

Viral genomic RNA was directly extracted from the allantoic fluid of each isolate using a Trizol RNA extraction kit (Invitrogen, Carlsbad, CA), according to the manufacturer's instructions. The cDNA was reverse transcribed from viral RNA with 6-nt random primer or a specific primer 5'-ACC AAA CAG AGA ATC-3' complementary to the 3' end of the NDV genomic RNA. A set of nine primer pairs specific for genotype III and IX

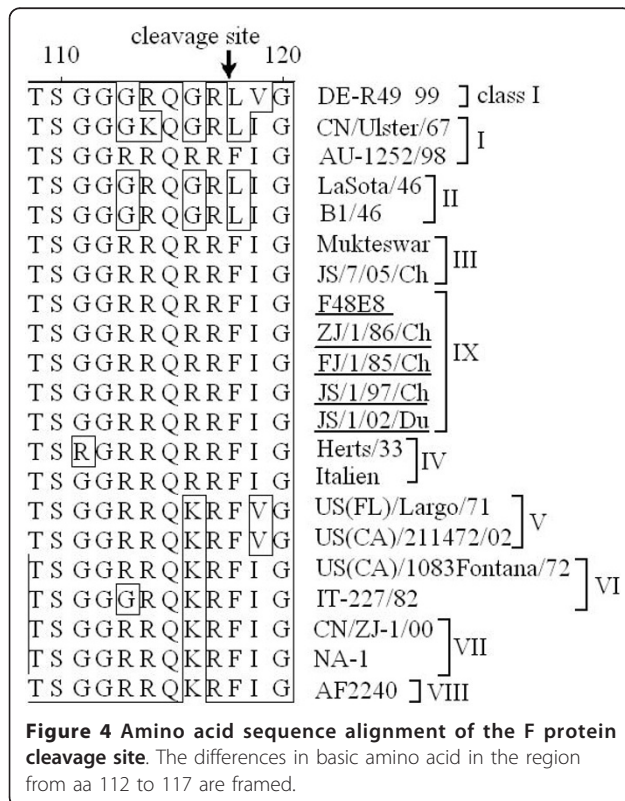


Table 4 Nine pairs of primers used to generate overlapping PCR fragments from genomes of F48-like NDV strains

Fragment designation		Primer sequence (5'-3')	Position	Expected product size (bp)
A	F ^a	ACCAAACAGAGAATCCGTGAG	1-21	2280
	R ^b	TGGACGATTTATTGCTAAGCTTG	2258-2280	
B	F	CAAGACTGGAGCAAGCAACT	2219-2238	2003
	R	GGAGAGGCATTTGCTATAGG	4202-4221	
C	F	GGGCTCAGTGATGTGCTCG	4100-4118	1964
	R	ATATAGGTAATGAGAGCAGATGTG	6040-6063	
D	F	AAATAATATGCGTGCCACCT	5434-5453	1661
	R	GAACGCAGAGTAGAAAAGAATA	7073-7094	
E	F	CAAGAACACCTGAATTTTATCCCG	6686-6909	2260
	R	TTAGATGCCTTTGGACCTGTTTTA	8922-8945	
F	F	TGGTTTCACTCAAATGGTCC	8876-8896	1223
	R	ATCCCTTCTGCCATTACCTG	10079-10098	
G	F	ACCCTTGAGTAC2TAAGAGATGA	9965-9987	2116
	R	TGTCCCCATAAGCCAGAT	12062-12080	
H	F	AATCTGATACCATAGAACCTTGT	11801-11823	2032
	R	GTCCGAAATGTCGCTGTG	13815-13832	
I	F	CTAGGAAGAGCCTTAATTTGAT	13350-13371	1838
	R	ACAAAGATTTGGTGAATGACA	15167-15187	

a: forward primer; b: reverse primer.

Madison, WI) using a specific primer 5TLF, 5'-GTC CAT TCT GTG CAG AGA GTT TAG TGA G-3', which was located from 14,502 nt to 14,527 nt in the viral genome (mentioned in the direction of 3' end to 5' end of genomic RNA). The reaction mixture was incubated at 42°C for 60 min, and then the cDNA was treated with an equal volume of 0.6 N NaOH for 20 min at 60°C to hydrolyze the mRNA and denature the first-strand cDNA. After purification by using PCR purification kit (Axygen, Union City, CA), the cDNA was ligated with adaptor CL+ by T4 RNA ligase according to procedures as described in 3' RACE. The resulting adaptor-ligated cDNA was amplified using primer 5TLF and anti-adaptor primer CL-. Heminested PCR reaction was then conducted using the primer CL- and specific primer 5TSF, 5'-CAA TAC TGG GTC TCA GAG TCA AAA ATC-3', which was located from 14724 nt to 14751 nt in the viral genome (mentioned in the direction of 3' to 5' of genomic RNA), and then 1 µl of 1:100 diluted primary PCR product was used as template.

Cloning and sequencing of the amplified products

RT-PCR products of overlapping fragments covering entire genome and GC-rich 5' NCR of NP gene and RT-PCR products by RACE were extracted from agarose gel, ligated into the TA cloning system (Promega, Madison, WI), then transferred into *E. coli* DH5α strain. At least four clones of each segment were sequenced in both directions using the ABI-3700-based (Applied Biosystems Inc.) fluorescent cycle sequencing technology by

Sangon Biotechnology (Shanghai, China), and then the correct sequences were determined.

Sequence analysis

Prediction of amino acid sequences, alignment of sequences and phylogenetic analysis were conducted using the MegAlign program (Windows 32, MegAlign 4.00) in the Lasergene package (DNASTAR Inc. Madison, WI 53715, USA). The sequences of overlapping DNA fragments were aligned and compiled into complete genome. Phylogenetic analysis was performed by using the Lasergene software package and MEGA version 4 (Tamura, Dudley, Nei, and Kumar 2007). Data and accession numbers of complete genome sequences of NDV strains in this study were presented in additional file 1 Table S1. Additional F gene sequences used in Figure 2 were taken from the EMBL/GenBank, the origins of which were described previously [8].

Additional material

Additional file 1: Table S1: Background information of NDV strains with complete genome sequences used in this study. The genotyping, accession number and references of those NDV strains are shown.

Abbreviations

APMV: avian paramyxovirus; CEFs: chicken embryo fibroblasts; NCR: Non-coding region; ND: Newcastle disease; NDV: Newcastle disease virus; RACE: rapid amplification of cDNA end; RT-PCR: reverse-transcription polymerase chain reaction.

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Author details

¹Key Laboratory of Animal Infectious Diseases, Yangzhou University, Yangzhou 225009, PR China. ²Shanghai Veterinary Institute, Chinese Academy of Agricultural Sciences, Shanghai 200241, PR China.

Authors' contributions

XQ, QS, SW and CM contributed for RT-PCR, sequence analysis and generation of phylogenetic tree. Shuang Wu isolated and collected viruses used in this study. LD performed the virus propagation in eggs. XQ and XL drafted the manuscript. XL, SH and YW coordinated overall planning and designed this study. All authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

- Alexander DJ: Newcastle disease, other avian paramyxoviruses, and pneumovirus infections. *Diseases of Poultry*. 11 edition. Ames, Iowa: Iowa State University Press; 2003, 63-69.
- Shimazu Y, Takao SJ, Irie T, Kiyotani K, Yoshida T, Sakaguchi T: Contribution of the leader sequence to homologous viral interference among Sendai virus strains. *Virology* 2008, **372**(1):64-71.
- Aldous EW, Alexander DJ: Detection and differentiation of Newcastle disease virus (avian paramyxovirus type 1). *Avian Pathol* 2001, **30**(2):117-128.
- de Leeuw O, Peeters B: Complete nucleotide sequence of Newcastle disease virus: evidence for the existence of a new genus within the subfamily Paramyxovirinae. *J Gen Virol* 1999, **80**(Pt 1):131-136.
- Mayo MA: A summary of taxonomic changes recently approved by ICTV. *Arch Virol* 2002, **147**(8):1655-1663.
- Krishnamurthy S, Samal SK: Nucleotide sequences of the trailer, nucleocapsid protein gene and intergenic regions of Newcastle disease virus strain Beaudette C and completion of the entire genome sequence. *J Gen Virol* 1998, **79**(Pt 10):2419-2424.
- Huang Y, Wan HQ, Liu HQ, Wu YT, Liu XF: Genomic sequence of an isolate of Newcastle disease virus isolated from an outbreak in geese: a novel six nucleotide insertion in the non-coding region of the nucleoprotein gene. *Brief Report. Arch Virol* 2004, **149**(7):1445-1457.
- Czeglédi A, Ujvári D, Somogyi E, Wehmann E, Werner O, Lomniczi B: Third genome size category of avian paramyxovirus serotype 1 (Newcastle disease virus) and evolutionary implications. *Virus Res* 2006, **120**(1-2):36-48.
- Millar NS, Chambers P, Emmerson PT: Nucleotide sequence of the fusion and haemagglutinin-neuraminidase glycoprotein genes of Newcastle disease virus, strain Ulster: molecular basis for variations in pathogenicity between strains. *J Gen Virol* 1988, **69**(Pt 3):613-620.
- Samson AC, Levesley I, Russell PH: The 36 K polypeptide synthesized in Newcastle disease virus-infected cells possesses properties predicted for the hypothesized 'V' protein. *J Gen Virol* 1991, **72**(Pt 7):1709-1713.
- Steward M, Vipond IB, Millar NS, Emmerson PT: RNA editing in Newcastle disease virus. *J Gen Virol* 1993, **74**(Pt 12):2539-2547.
- Gould AR, Hansson E, Selleck K, Kattenbelt JA, Mackenzie M, Della-Porta AJ: Newcastle disease virus fusion and haemagglutinin-neuraminidase gene motifs as markers for viral lineage. *Avian Pathol* 2003, **32**(4):361-373.
- Alexander DJ, Campbell G, Manvell RJ, Collins MS, Parsons G, McNulty MS: Characterisation of an antigenically unusual virus responsible for two outbreaks of Newcastle disease in the Republic of Ireland in 1990. *Vet Rec* 1992, **130**(4):65-68.
- Mia Kim L, King DJ, Suarez DL, Wong CW, Afonso CL: Characterization of class I Newcastle disease virus isolates from Hong Kong live bird markets and detection using real-time reverse transcription-PCR. *J Clin Microbiol* 2007, **45**(4):1310-1314.
- Bjarnadottir H, Gudmundsson B, Gudnason J, Jonsson JJ: Encapsidation determinants located downstream of the major splice donor in the maedi-visna virus leader region. *J Virol* 2006, **80**(23):11743-11755.
- Liu X, Wang X, Wu S, Hu S, Peng Y, Xue F: Surveillance for avirulent Newcastle disease viruses in domestic ducks (*Anas platyrhynchos* and *Cairina moschata*) at live bird markets in Eastern China and characterization of the viruses isolated. *Avian Pathol* 2009, **38**(5):377-391.
- Seal BS, Wise MG, Pedersen JC, Senne DA, Alvarez R, Scott MS, King DJ, Yu Q, Kapczynski DR: Genomic sequences of low-virulence avian paramyxovirus-1 (Newcastle disease virus) isolates obtained from live-bird markets in North America not related to commonly utilized commercial vaccine strains. *Vet Microbiol* 2005, **106**(1-2):7-16.
- Ujvári D, Wehmann E, Herczeg J, Lomniczi B: Identification and subgrouping of pigeon type Newcastle disease virus strains by restriction enzyme cleavage site analysis. *J Virol Methods* 2006, **131**(2):115-121.
- Lomniczi B, Wehmann E, Herczeg J, Ballagi-Pordany A, Kaleta EF, Werner O, Meulemans G, Jorgensen PH, Mante AP, Gielkens AL, et al: Newcastle disease outbreaks in recent years in western Europe were caused by an old (VI) and a novel genotype (VII). *Arch Virol* 1998, **143**(1):49-64.
- Herczeg J, Wehmann E, Bragg RR, Travassos Dias PM, Hadjiev G, Werner O, Lomniczi B: Two novel genetic groups (VIIb and VIII) responsible for recent Newcastle disease outbreaks in Southern Africa, one (VIIb) of which reached Southern Europe. *Arch Virol* 1999, **144**(11):2087-2099.
- Liu XF, Wan HQ, Ni XX, Wu YT, Liu WB: Pathotypical and genotypical characterization of strains of Newcastle disease virus isolated from outbreaks in chicken and goose flocks in some regions of China during 1985-2001. *Arch Virol* 2003, **148**(7):1387-1403.
- Liang Y, Ma WT: A fowl disease caused by a filterable virus. *Agricult Bull (in Chinese)* 1946, **12**:14-16.
- Liang R, Cao DJ, Li JQ, Chen J, Guo X, Zhuang FF, Duan MX: Newcastle disease outbreaks in western China were caused by the genotypes VIIa and VIII. *Vet Microbiol* 2002, **87**(3):193-203.
- Miller PJ, Decanini EL, Afonso CL: Newcastle disease: evolution of genotypes and the related diagnostic challenges. *Infect Genet Evol* 10(1):26-35.
- Yu L, Wang Z, Jiang Y, Chang L, Kwang J: Characterization of newly emerging Newcastle disease virus isolates from the People's Republic of China and Taiwan. *J Clin Microbiol* 2001, **39**(10):3512-3519.
- Qin ZM, Tan LT, Xu HY, Ma BC, Wang YL, Yuan XY, Liu WJ: Pathotypical characterization and molecular epidemiology of Newcastle disease virus isolates from different hosts in China from 1996 to 2005. *J Clin Microbiol* 2008, **46**(2):601-611.
- Ronin C, Bouchilloux S, Granier C, van Rietschoten J: Enzymatic N-glycosylation of synthetic Asn-X-Thr containing peptides. *FEBS Lett* 1978, **96**(1):179-182.
- Hart GW, Brew K, Grant GA, Bradshaw RA, Lennarz WJ: Primary structural requirements for the enzymatic formation of the N-glycosidic bond in glycoproteins. Studies with natural and synthetic peptides. *J Biol Chem* 1979, **254**(19):9747-9753.
- Connaris H, Takimoto T, Russell R, Crennell S, Moustafa I, Portner A, Taylor G: Probing the sialic acid binding site of the hemagglutinin-neuraminidase of Newcastle disease virus: identification of key amino acids involved in cell binding, catalysis, and fusion. *J Virol* 2002, **76**(4):1816-1824.
- Takimoto T, Taylor GL, Crennell SJ, Scroggs RA, Portner A: Crystallization of Newcastle disease virus hemagglutinin-neuraminidase glycoprotein. *Virology* 2000, **270**(1):208-214.
- Iorio RM, Field GM, Sauvion JM, Mirza AM, Deng R, Mahon PJ, Langedijk JP: Structural and functional relationship between the receptor recognition and neuraminidase activities of the Newcastle disease virus hemagglutinin-neuraminidase protein: receptor recognition is dependent on neuraminidase activity. *J Virol* 2001, **75**(4):1918-1927.
- Sakaguchi T, Toyoda T, Gotoh B, Inocencio NM, Kuma K, Miyata T, Nagai Y: Newcastle disease virus evolution. I. Multiple lineages defined by sequence variability of the hemagglutinin-neuraminidase gene. *Virology* 1989, **169**(2):260-272.
- Locke DP, Sellers HS, Crawford JM, Schultz-Cherry S, King DJ, Meinersmann RJ, Seal BS: Newcastle disease virus phosphoprotein gene analysis and transcriptional editing in avian cells. *Virus Res* 2000, **69**(1):55-68.

34. Seal BS, King DJ, Locke DP, Senne DA, Jackwood MW: **Phylogenetic relationships among highly virulent Newcastle disease virus isolates obtained from exotic birds and poultry from 1989 to 1996.** *J Clin Microbiol* 1998, **36**(4):1141-1145.
35. Seal BS, King DJ, Meinersmann RJ: **Molecular evolution of the Newcastle disease virus matrix protein gene and phylogenetic relationships among the paramyxoviridae.** *Virus Res* 2000, **66**(1):1-11.
36. Wise MG, Sellers HS, Alvarez R, Seal BS: **RNA-dependent RNA polymerase gene analysis of worldwide Newcastle disease virus isolates representing different virulence types and their phylogenetic relationship with other members of the paramyxoviridae.** *Virus Res* 2004, **104**(1):71-80.
37. Taniura N, Saito M, Okuwa T, Saito K, Ohara Y: **Different subcellular localization of Theiler's murine encephalomyelitis virus leader proteins of GDVII and DA strains in BHK-21 cells.** *J Virol* 2009, **83**(13):6624-6630.
38. Plumet S, Herschke F, Bourhis JM, Valentin H, Longhi S, Gerlier D: **Cytosolic 5'-triphosphate ended viral leader transcript of measles virus as activator of the RIG I-mediated interferon response.** *PLoS One* 2007, **2**(3): e279.
39. Qiu X, Sun Q, Yao C, Dong L, Wu Y, Hu S, Liu X: **[Full-length genome analysis of two genotype III velogenic Newcastle diseases virus strains reveals their close relationship with vaccine Mukteswar].** *Wei Sheng Wu Xue Bao* 2009, **49**(3):302-308.
40. Ricour C, Borghese F, Sorgeloos F, Hato SV, van Kuppeveld FJ, Michiels T: **Random mutagenesis defines a domain of Theiler's virus leader protein that is essential for antagonism of nucleocytoplasmic trafficking and cytokine gene expression.** *J Virol* 2009, **83**(21):11223-11232.

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