# COMMENT

# **Open Access**

# First detection of porcine circovirus 4 (PCV-4) in Europe



Rocío Holgado-Martín<sup>1†</sup>, José Luís Arnal<sup>2†</sup>, Marina Sibila<sup>3,4,5</sup>, Giovanni Franzo<sup>6</sup>, Desireé Martín-Jurado<sup>2</sup>, David Risco<sup>1</sup>, Joaquim Segalés<sup>3,4,7\*†</sup> and Luís Gómez<sup>1\*†</sup>

# Abstract

Porcine circovirus 4 (PCV-4) is a novel virus recently discovered (2019) in domestic pigs from China, although several studies have proven its circulation since 2008. Later, PCV-4 was also detected in wild boar populations from China and domestic pigs from South Korea and Thailand. Currently, Asia is so far the only continent where this novel virus has been reported; few studies carried out in South America and Europe failed in the attempt to detect it. The objective of this Comment is to communicate the first detection of PCV-4 in Europe, specifically in wild boar and domestic pigs from Mid-South-Western Spain. A retrospective study was carried out on wild boar and domestic pigs, both extensively (Iberian breed) and intensively raised, from Spain and Italy, sampled between 1998 and 2022. PCV-4 genome detection was attempted using different conventional or quantitative real time PCR (qPCR) protocols and some positive results were confirmed through Sanger sequencing. A total of 57 out of 166 (34.3%) Spanish wild boar and 9 out of 223 (4%) Iberian pigs (both geographically located in the Mid-South-Western Spain) were qPCR positive, while the rest of tested animals from North-Eastern Spain and Italy were negative. Partial sequences of Rep or Cap genes of selected samples confirmed the presence of PCV-4. The relatively high prevalence in wild boar and the low one in Iberian pigs from the same areas suggests intra- and interspecific transmission, being the wild boar a potential viral reservoir. The epidemiological and clinical importance of these findings are currently unknown, but guarantees further research on this novel virus.

Keywords Porcine circovirus 4 (PCV-4), Wild boar, Iberian pig, Spain, Italy

 $^{\dagger}\text{Rocio}$  Holgado-Martín and José Luís Arnal equally contributing first authors.

<sup>+</sup>Joaquim Segalés and Luís Gómez equally contributing last authors and corresponding ones.

\*Correspondence: Joaquim Segalés joaquim.segales@irta.cat Luís Gómez luih@unex.es <sup>1</sup>Departamento de Medicina Animal, Unidad de Anatomía Patológica y

Anatomía Comparada, Facultad de Veterinaria de Cáceres, Universidad de Extremadura, Cáceres 10003, Spain

<sup>2</sup>Exopol, Autovaccines and Veterinary Diagnostics, Polígono Río Gállego D/14, San Mateo de Gállego, Zaragoza 50840, Spain
<sup>3</sup>Programa de Sanitat Animal, IRTA, Centre de Recerca en Sanitat Animal (CReSA), Universitat Autònoma de Barcelona (UAB), Campus, Bellaterra 08193, Spain
<sup>4</sup>Unitat Mixta d'Investigació IRTA-UAB en Sanitat Animal, Centre de Recerca en Sanitat Animal (CReSA), Campus de la Universitat Autónoma de Barcelona (UAB), Bellaterra 08193, Spain
<sup>5</sup>WOAH collaborating Centre for the Research and Control of Emerging and Re-Emerging Swine Diseases in Europe (IRTA-CReSA), Bellaterra 08193, Spain
<sup>6</sup>Department of Animal Medicine, Production and Health (MAPS), Padua University, Legnaro 35020, Italy
<sup>7</sup>Departament de Sanitat i Anatomia Animals, Facultat de Veterinària, Universitat Autònoma de Barcelona, Bellaterra 08193, Spain



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

## Background

Porcine circovirus 4 (PCV-4) is a novel virus from the Circoviridae family, which comprises small icosahedral and non-enveloped viruses with single-stranded and circular DNA genome. It was discovered in 2019 in domestic pigs (Sus scrofa domestica) in China [1]. However, retrospective studies have indicated that infection could be traced back to 2008 at least [2, 3]. Subsequently, it has also been found in wild boar (Sus scrofa) in China [4]. In addition, PCV-4 DNA has been detected in other domestic species such as dairy cows and dogs in China [5, 6]. Outside China, PCV-4 genome has been found only in Korea and Thailand [7, 8], although several surveys have been conducted in other continents, such as South America and Europe [9, 10]. Therefore, Asia is the only continent where the virus has been detected so far. The limited sequences available point out the existence of two different genotypes of PCV-4 (PCV-4a and PCV-4b). Both genotypes have been found in China, although only the PCV-4b one has been identified in Thailand and South Korea [7, 8].

PCV-4 DNA has been detected in domestic pigs affected by respiratory disorders and porcine dermatitis and nephropathy syndrome (PDNS), as well as in apparently healthy animals [1]. Detection of PCV-4 genome in wild boar has not been associated to any specific clinical condition [4]. Interestingly, PCV-4 has been detected in co-infection with other porcine circoviruses [4].

Most of the studies on PCV-4 have been based on conventional or real time quantitative PCR (qPCR) methods; only one used an in situ hybridization technique [8], although the obtained results were not conclusive.

# Main text

The incidence of pig infectious emerging and re-emerging diseases is continuously increasing [11]. These infections tend to spread easily because of the globalization of trade and the movement of people and animals worldwide [12]. In Spain, the porcine industry is mainly based on intensive farms; however, the Iberian pig production is still featured by a semi-extensive system specially in Mid-South-Western Spain, where pigs usually have contact with wild animals, such as wild boar [13]. Taking into account the economic and social importance of the swine industry in Spain as well as in Europe, the spread of emerging pathogens such as PCV-4 should be considered a matter of concern. Therefore, the objective of this Comment is to communicate the presence of PCV-4 in Europe, at least in Spain.

Table 1 summarizes the results of a collaborative and retrospective study carried out by several research groups from Spain and Italy using wild boar and domestic pig samples taken in different time periods between 1998 and 2023. Analyzed samples were collected from previously existing collections from epidemiological studies (in the case of wild boar), diagnostic cases or monitoring sampling.

Different PCR methodologies were used for the purpose, including techniques already published in the literature [1, 9] and new in-house conventional and real time PCR protocols targeting the Rep gene. Sanger sequencing was carried out on Rep or Cap gene from those samples whose Cq value was lower than 32, to maximize the likelihood of obtaining good-quality sequences [14]. The obtained nucleotide sequences were analysed and assembled using MEGA11 Molecular Evolutionary Genetics Analysis version 11 software [15].

A total of 57 different Spanish wild boar were positive for PCV-4 qPCR; specifically, 56 lymph nodes and 8 sera, with Cq values ranging 21-38 and 27-36, respectively. All positive serum samples were also positive to lymph node, except one. The Rep gene of 6 strains detected from positive lymph nodes was partially sequenced (GenBank accession numbers OR367318-OR367323) confirming the detection of PCV-4 genome. On the other hand, a total of 9 samples (seven intestines and two lungs) from different Iberian pig ranches were qPCR positive (Cq ranging from 31 to 37). The Cap gene from three samples was partially sequenced, confirming as well the detection of PCV-4 genome (GenBank accession numbers OR333699, OR333700 and OR359763). All qPCR positive animals were geographically located in Mid-South-Western Spain. Wild boar PCV-4 sequences were genetically identical to each other in the considered Rep region. A close relationship (identity>99.7) was detected with several other Chinese and South Korean strains. The only exception was represented by strain OR367323, which showed a p-distance=0.02 (i.e. 98% genetic identity) compared to the other Spanish wild boar sequences and a p-distance of 0.015 with a Chinese strain (OP497960) collected in 2022. A similar distance (p-distance=0.017) was detected with other Chinese strains sampled since 2017 (i.e. MT882411, MK986820, NC\_055580). The Cap sequences obtained from the Iberian pigs had the closest relationship with strains collected from China and South Korea since 2017 (p-distance=0.007). PCV-4 DNA was not detected in the other commercial pig samples from Spain (North-Eastern Spain) or in domestic swine (backyard and intensively raised) and wild boar from Italy (Table 1).

### Conclusions

To the best of our knowledge, the detection of PCV-4 in wild boar and Iberian pigs represents the first report of this virus in Europe. Interestingly, most of infected animals were wild boar, which may suggest their potential reservoir role. Moreover, analysed animals were located in the Mid-South-Western Spain (Fig. 1), which

Table 1	Wild boar and domestic pig samples tested by qPCR/PCR methods for the detection of PCV-4 in Sp	ain and Italy. Percentage
of qPCR/	PCR positive ones are given attending to the type of sample and/or clinical condition studied	

Country	Time period	Suidae species tested	Tested sample or clini- cal condition	Number of studied samples	Number of positive samples (%)
Spain	2011-2015	Wild boar*	Lymph nodes	166	56 (33.7%)
			Serum <sup>1</sup>	90	8 (8.9%)
	2021-2023	lberian pig*	Digestive <sup>2</sup>	99	7 (7%)
			Lung	57	2 (3.5%)
			Reproductive <sup>3</sup>	14	0 (0%)
			Others <sup>4</sup>	53	0 (0%)
	2018-2021	Commercial breed pig**	Digestive <sup>2</sup>	73	0 (0%)
			Lung	32	0 (0%)
			Reproductive <sup>2</sup>	66	0 (0%)
			Others <sup>4</sup>	13	0 (0%)
	1998–2020	Commercial breed pig**	PDNS cases <sup>5</sup>	100	0 (0%)
	2018-2020	Commercial breed pig**	Serum <sup>6</sup>	900	0 (0%)
Italy	2014-2015	Wild boar	Serum	187	0 (0%)
	2017-2018	Wild boar	Serum	29	0 (0%)
	2021-2022	Wild boar	Lung and lymph node homogenate	103	0 (0%)
		Commercial breed pig	Serum	45	0 (0%)
		Backyard pigs	Lung and lymph node homogenate	68	0 (0%)

\*Samples from animals located in Mid-South-Western Spain; \*\*Samples from animals located in North-Eastern Spain

<sup>1</sup>The 90 sera from Spanish wild boar belonged to the 166 ones from which lymph nodes were tested

<sup>2</sup>Digestive samples included intestines, feces and rectal swabs

<sup>3</sup>Reproductive samples included foetus homogenates, placenta and fetus stomach content

<sup>4</sup>Others included sera and oral fluids from monitoring analytics, joint swabs from poliarthritis illness and organs such as liver, spleen or brain from systemic disorders and sudden deaths

<sup>5</sup>Mixture of tissues (lymph nodes, tonsil, spleen and lung) of porcine dermatitis and nephropathy syndrome (PDNS) affected pigs

<sup>6</sup>Serum of 3 day-old healthy piglets

is the traditional area of Iberian pig rearing in outdoor conditions. Since the Iberian pig semi-extensive breeding system is a special ecosystem called "Dehesa", where domestic animals and wild fauna can interact directly or indirectly, it is tempting to speculate the potential interspecific transmission of pathogens from wild boar to Iberian pig [13, 16]. The lack of detection of PCV-4 infection in intensively reared pigs could be related to the higher biosecurity level in this type of farms and/or the lack of effective contact with the wild. In any case, the route of entry of PCV-4 in wild boar and Iberian pigs in Spain remains unknown.

A recent study carried out in Spain and Italy yielded negative results in the attempt to detect PCV-4 in serum and tissue homogenates collected from both wild boar and domestic pigs [9]. One possible explanation is that these previous surveys mainly included samples belonging to intensive breed pigs from Italy and North-Eastern Spain. Also, the previously tested wild boar were only from Italy, and novel results confirmed again negativity for PCV-4 in these populations. In addition, tested Italian backyard pigs were negative for the new virus. In consequence, PCV-4 seems to be still absent in pigs from these geographic areas.

In summary, this Comment confirms that PCV-4 has been retrospectively detected in Europe, specifically in wild boar and Iberian pigs reared in Mid-South-Western Spain. Interestingly, no evidence of infection has been found in domestic swine from North-Eastern Spain or in pigs and wild boar from Italy. If this epidemiological scenario means a limited geographical distribution of PCV-4 in Spain or in Europe requires further investigations. No clue on the potential pathogenicity of PCV-4 in domestic swine or wild boar can be drawn from the current exploratory study.



Fig. 1 Map of Spain showing the location (provinces) of PCV-4 PCR positive wild boar (red circles) and Iberian pigs (red squares). The green circles, squares and triangles show the provinces in which wild boar, Iberian pigs and intensive breeding pigs, respectively, yielded negative PCV-4 PCR negative results

#### Authors' contributions

RHM, JLA, MS, GF and DM performed the analysis of the samples and interpreted the data. DR, JLA, GF, JS and LG took the samples. All authors wrote, read and approved the final version of the manuscript.

#### Funding

This study was partly supported by the "Consejería de Economía, Ciencia y Agenda Digital, Junta de Extremadura" and by "European agricultural fund for rural development", "Fondo Europeo de Desarrollo Regional: Una manera de hacer Europa" (IB20027).

#### Declarations

#### Ethics approval and consent to participate

All studied samples used corresponded to sample submissions from previously existing collections from epidemiological studies (wild boar), diagnostic cases or monitoring sampling.

#### **Competing interests**

The authors declare that they have no competing interests.

Received: 3 August 2023 / Accepted: 8 September 2023 Published online: 10 October 2023

#### References

- Zhang HH, Hu WQ, Li JY, Liu TN, Zhou JY, Opriessnig T, Xiao CT. Novel circovirus species identified in farmed pigs designated as porcine circovirus 4, Hunan province, China. Transbound Emerg Dis. 2020;67(3):1057–61. https:// doi.org/10.1111/tbed.13446.
- Hou CY, Zhang LH, Zhang YH, Cui JT, Zhao L, Zheng LL, Chen HY. Phylogenetic analysis of porcine circovirus 4 in Henan Province of China: a retrospective study from 2011 to 2021. Transbound Emerg Dis. 2022;69(4):1890–901. https://doi.org/10.1111/tbed.14172.
- Ge M, Hu WQ, Ning KM, Li SY, Xiao CT. The seroprevalence of the newly identified porcine circovirus type 4 in China investigated by an enzymed-linked immunosorbent assay. Transbound Emerg Dis. 2021;68(6):2910–4. https://doi. org/10.1111/tbed.14184.
- Wu H, Hou C, Wang Z, Meng P, Chen H, Cao H. First complete genomic sequence analysis of porcine circovirus type 4 (PCV4) in wild boars. Vet Microbiol. 2022;273:109547. https://doi.org/10.1016/j.vetmic.2022.109547.
- Xu T, Chen L, Huang BZ, Zhu L, Sun XG, Lai SY, Ai YU, Zhou YC, Xu ZW. The first dog-origin porcine circovirus type 4 complete genomic sequence have high homology with that of pig-derived strains. Front Microbiol. 2023;14:1121177. https://doi.org/10.3389/fmicb.2023.1121177.
- Xu T, Chen XM, Fu Y, Ai Y, Wang DM, Wei ZY, Li XS, Zheng LL, Chen H. Y. Crossspecies transmission of an emerging porcine circovirus (PCV4): first molecular detection and retrospective investigation in dairy cows. Vet Microbiol. 2022;273:109528. https://doi.org/10.1016/j.vetmic.2022.109528.
- Nguyen VG, Do HQ, Huynh TML, Park YH, Park BK, Chung HC. Molecular-based detection, genetic characterization and phylogenetic analysis of porcine circovirus 4 from korean domestic swine farms. Transbound Emerg Dis. 2022;69(2):538–48. https://doi.org/10.1111/tbed.14017.
- 8. Sirisereewan C, Nguyen TC, Piewbang C, Jittimanee S, Kedkovid R, Thanawongnuwech R. Molecular detection and genetic characterization of porcine

circovirus 4 (PCV4) in Thailand during 2019–2020. Sci Rep. 2023;13(1):5168. https://doi.org/10.1038/s41598-023-32382-1.

- Franzo G, Ruiz A, Grassi L, Sibila M, Drigo M, Segalés J. Lack of porcine circovirus 4 genome detection in pig samples from Italy and Spain. Pathogens. 2020;9(6):433. https://doi.org/10.3390/pathogens9060433.
- Vargas-Bermudez DS, Mogollón JD, Jaime J. The prevalence and genetic diversity of PCV3 and PCV2 in Colombia and PCV4 Survey during 2015–2016 and 2018–2019. Pathogens. 2022;11(6):633. https://doi.org/10.3390/ pathogens11060633.
- Fournié G, Kearsley-Fleet L, Otte J, Pfeiffer DU. Spatiotemporal trends in the discovery of new swine infectious agents. Vet Res. 2015;46:1–9. https://doi. org/10.1186/s13567-015-0226-8.
- Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, Daszak P. Global trends in emerging infectious diseases. Nature. 2008;451(7181):990–3. https://doi.org/10.1038/nature06536.
- Kukielka E, Barasona JA, Cowie CE, Drewe JA, Gortazar C, Cotarelo I, Vicente J. Spatial and temporal interactions between livestock and wildlife in South Central Spain assessed by camera traps. Prev Vet Med. 2013;112(3–4):213–21. https://doi.org/10.1016/j.prevetmed.2013.08.008.

- 14. Men AE, Wilson P, Siemering K, Forrest S. Sanger DNA sequencing. Next Generation Genome Sequencing: Towards Personalized Medicine. 2008;1–11. https://doi.org/10.1002/9783527625130.ch1.
- Tamura K, Stecher G, Kumar S. MEGA11: Molecular Evolutionary Genetics Analysis Version 11. Mol Biol Evol. 2021;38(7):3022–7. https://doi.org/10.1093/ molbev/msab120.
- Rodríguez-Prieto V, Kukielka D, Martínez-López B, de las Heras Al, Barasona J, Gortázar C, Sánchez-Vizcaíno JM, Vicente J. Porcine reproductive and respiratory syndrome (PRRS) virus in wild boar and Iberian pigs in southcentral Spain. Euro J Wild Res. 2013;59:859–67. https://doi.org/10.1007/ s10344-013-0739-2.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.