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Cytomegalovirus infection may be oncoprotective against neoplasms of B-lymphocyte lineage: single-institution experience and survey of global evidence

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Abstract

Background: Although cytomegalovirus (CMV) is not considered tumorigenic, there is evidence for its oncomodulatory effects and association with hematological neoplasms. Conversely, a number of experimental and clinical studies suggest its putative anti-tumour effect. We investigated the potential connection between chronic CMV infection in patients with B-lymphocyte (B-cell) malignancies in a retrospective single-center study and extracted relevant data on CMV prevalences and the incidences of B-cell cancers the world over.

Methods: In the clinical single-center study, prevalence of chronic CMV infection was compared between patients with B-cell leukemia/lymphoma and the healthy controls. Also, global data on CMV seroprevalences and the corresponding country-specific incidences of B- lineage neoplasms worldwide were investigated for potential correlations.

Results: Significantly higher CMV seropositivity was observed in control subjects than in patients with B-cell malignancies (p = 0.035). Moreover, an unexpected seroepidemiological evidence of highly significant inverse relationship between country-specific CMV prevalence and the annual incidence of B-cell neoplasms was noted across the populations worldwide (p = -0.625, p < 0.001).

Conclusions: We try to draw attention to an unreported interplay between CMV infection and B-cell lymphomagenesis in adults. A large-scale survey across > 70 countries disclosed a link between CMV and B-cell neoplasms. Our evidence hints at an antagonistic effect of chronic CMV infection against B-lymphoproliferation.

Keywords: Cytomegalovirus, B-cell malignancies, Global, Seroprevalence, Oncoprotection

Background

Although a benign infectious agent in the healthy, the human cytomegalovirus (CMV) is a notorious driver of morbidity and mortality in hematological patients with failed immunocompetence [1]. Cytomegalovirus infection is the most significant viral complication of allogeneic hematopoietic cell transplantation (HCT) [2, 3]. The virus is highly pervasive, with a widely varying seroprevalence due to different demographic factors including socioeconomic status (SES) of populaces and communities [4–6].

While not regarded as a *bona fide* tumorigenic virus, CMV boasts an array of features that imply its oncogenic potential. The genome of CMV carries two anti-apoptotic

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genes, upregulates p53 [7] and augments anaplasia in cancer cells and/or tumor-associated cells [8–10]. Also, CMV may contribute to cancer formation via a "hit-andrun" mechanism, as well [11–14]. Aditionally, recent studies have identified congenital CMV infection as a risk for developing childhood hematologic malignancy [15, 16].

In contrast, clinical evidence that favors an anti-limphoproliferative effect of CMV, recently came from de Carvalho Batista Éboli et al. (2022). They verified liver pretransplant positivity for CMV as a protective factor for posttransplant lymphoproliferative disorder (PTLD) in pediatric patients [17]. A possible virus-vs-leukemia phenomenon has also been described [18], along with inhibition of migration of tumor cells [19, 20]. Several experiments done with murine CMV documented apoptosis in tumor cells [21, 22]. In humans, patients experiencing CMV reactivation early after allogeneic HCT for acute leukemia and non-Hodgkin lymphomas (NHL) have reduced relapse rates [23–27].

Research on CMV infection, reactivation, and multiorgan sequelae preferentially focuses on T-lymphocyte (T-cell) immune response. Recent studies on humanized animal models make the case in favor of importance of anti-CMV antibodies as being produced by host B-cells [28–32].

We asked if CMV seroststus may relate to a possible oncomodulatory role played by chronic CMV infection in individuals afflicted by lymphoid neoplasias derived from a single histologic lineage. The current work provides evidence that chronic CMV infection protects against malignant diseases of B-lymphocyte origin.

Methods

Patient and control cohorts

Our retrospective study cohort (N=83; M/F=43/40) was monocentric and comprised patients treated at the Clinic of Hematology, University Clinical Center, Belgrade, Republic of Serbia. The median age was 49.45 years (M=52.3, range 20–73; F=48.1, range 21–73). Information on demographic markers, underlying B-cell disorders, and chemoradiation regimens administered was abstracted from patients' medical records. Principal patient characteristics, diagnoses and chemotherapy regimens are presented in Tables 1 and 2. Close relatedness of malignant diseases with B-lymphocyte ontogeny was considered to have a virological authority over the clinical diversity of B-cell neoplasms.

The control group (N=259; M/F=73/186) consisted of population-based pauci-symptomatic noninstitution-alized civilians (mean age: 41.79 years, range: 20–86). None among the controls has had a record of malignant

disease. Study cohorts differed substantially by age and gender (p < 0.001) requiring statistical matching (Table 3).

Sampling and data collection

Whole blood was a clinical source of samples collected between February and November 2017 by venipuncture using standardized clot-activator vacutainers. After clotting and centrifugation the serum fraction was screened for anti-CMV IgG and IgM antibodies at the Virology Laboratory of the Institute of Microbiology and Immunology, Faculty of Medicine, University of Belgrade. Antibody classes were determined by means of commercial anti-CMV ELISA IgG and IgM kits (EUROIMMUN AG, Lübeck, Germany), with antibody detection performed spectrophotometrically on an ELISA Reader 270 (bioMérieux, Marcy-l'Étoile, France).

Peripheral blood samples from control cohort members were profiled for the presence of anti-CMV antibodies at the Institute of Virology, Vaccines, and Sera "Torlak", Serbian National Reference Laboratory for Viruses. Commercial kits (Enzygnost, Marburg, Germany) and a Multiskan Ex ELISA Reader (Thermo Electron Corporation, Waltham, MA, USA) were used to detect IgG and IgM classes.

Prevalence of IgG seropositivity is a hallmark of past infections in a population [6]. Cytomegalovirus positivity was based on detection of either CMV specific IgG or both IgG and IgM in the serum indicating contact with the pathogen. All persons presenting with an antibody profile consistent with primoinfection were excluded from the study, as it was posited that there is not enough time for a B-cell malignancy to develop within a first-contact millieu.

Diseases herein studied belonged strictly to B-cell immunolymphoproliferative disorders. The diagnoses were established and morphologically code-specified according to the International Classification of the Diseases for Oncology by World Health Organization, ICD-0-3 [33] and the 2016 revision of the World Health Organization Classification of Lymphoid Neoplasms [34].

In order to compare our results with available data at a global level, we interrogated published information on CMV prevalences and burden of B-cell malignancies across the globe. The PubMed advanced search was used with the search keywords "cytomegalovirus", "CMV", "B lymphoma", "Hodgkin's disease", "non-Hodgkin lymphoma", "B acute lymphoblastic leukemia", "B chronic lymphocytic leukemia" and "myeloma", all being neoplasms of the B-cell lineage.

Incidences of B-cell malignancies were obtained from the World Health Organization Global Cancer Observatory (GLOBOCAN) [35] and compared to CMV prevalences from 74 countries for which this data was Janković et al. Virology Journal (2022) 19:155 Page 3 of 17

 Table 1
 Principal demographics, clinical characteristics, and CMV serology of the patient group

	IgG positive	IgG negative	<i>p</i> -value
Patients (N = 83)	75 (90.4%)	8 (9.6%)	N/A
Age (median, years)	50.05	43.75	0.070 ^a
Age categories (years)			
20–39	18 (85.7%)	3 (14.3%)	0.054 ^b
40–59	34 (89.5%)	4 (10.5%)	
>59	23 (95.8%)	1 (4.2%)	
Gender			
Male	37 (86%)	6 (14%)	0.266 ^c
Female	38 (95%)	2 (5%)	
Diagnose; ICD-O-3 code*			
Non-Hodgkin lymphoma; 9591/3	33 (94.3%)	2 (5.7%)	0.339 ^c
Hodgkin's disease; 9650/3	18 (81.8%)	4 (18.2%)	
B-chronic lymphocytic leukemia; 9823/3	9 (90%)	1 (10%)	
Waldenström macroglobulinemia; 9761/3	2 (100%)	0	
B-cell lymphoma, NOS; 9690/3	2 (100%)	0	
Plasma cell myeloma; 9732/3	2 (66.7%)	1 (33.7%)	
Hairy cell leukemia; 9940/3	2 (100%)	0	
B-cell acute lymphoblastic leukemia; 9811/3	7 (100%)	0	
Antiviral therapy (Acyclovir)**			
Yes	17 (81%)	4 (19%)	0.438 ^c
No	36 (89.5%)	4 (10.5)	
Chemotherapy [†]			
Yes	38 (92.7%)	3 (7.3%)	0.706 ^c
No	25 (83.3%)	5 (16.7%)	

^{*}Diagnostic recognition and technical signification of lymphoid B-cell dyscrasias listed were morphologically code-specified and comply with criteria of the Classification of Diseases for Oncology (World Health Organization) and the 2016 revision of lymphoid neoplasms [35, 69]

available [36]. Age-adjusted annual incidence rates ($/10^5$ population) of B-cell neoplasms (standardized to the year 2000 US Census Bureau million population by the direct method) were collected and presented as sums rather than separately and apart.

It is important to note that published reports do not always clearly discriminate between B-cell and T-cell disorders. Cases of B- and T-acute lymphoblastic leukemia (B-ALL and T-ALL) were frequently presented jointly as "ALL". B- and T-non-Hodgkin's lymphoma (B-NHL and T-NHL) were often jointly described as "NHL". Moreover, even if B- and T-cell components of lymphoproliferative diseases were reported, complementary information on the prevalence of CMV seropositivity for each component was often not reported. Crude annual incidence rates (/10⁵) of all B-cytopathies were summed-up in Tables 4 and 5. Merging

the rates enhanced their statistical power and ease of interpretation.

Clinical sampling was approved by University Clinical Centre of Serbia, University of Belgrade Ethical Review Board. The patients signed individually a document of informed consent.

Statistical analysis

Results are presented as count (percent) or median (min-max) depending on data type. Groups were compared using non-parametric tests, Fisher's exact test for frequencies, Mantel-Haenszel chi square test for trend and Mann-Whitney U test for numeric data with non-normal distribution. Propensity score matching was performed in order to find the best matching cases in control group by age and gender. Correlation between numerical variables was performed using Spearman correlation

^{**}Data missing in 22 patients

[†] Data missing in 12 patients

 $^{^{\}rm a}$ Mann–Whitney ${\it U}$ test

^b Mantel-Haenszel chi square test for trend

^c Fisher's Exact Test

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Table 2 Details on chemotherapy

Regimens*	Patients (N=83)
ABVD	4
BEAM	1
CHOP	4
COP	1
DHAP	10
R-DHAP	1
Endoxan	1
ESHAP	1
PAD	1
R-EPOCH	1
R-CHOP	8
HyperCVAD	8
Untreated	42

*ABVD—Doxorubicin hydrochloride (Adriamycin), Bleomycin sulfate, Vinblastine sulfate, and Dacarbazine; BEAM—Carmustine (BiCNU), Etoposide, Cytarabine (Ara-C, cytosine arabinoside), Melphalan; R-CHOP—Rituximab, Cyclophosphamide, Doxorubicin hydrochloride, Vincristine (Oncovin), Prednisolone; CTD—Cyclophosphamide (Endoxan), Thalidomide, Dexamethasone; DHAP—Dexamethasone, High-dose Ara-C, Platinol (cisplatin); ESHAP—Etoposide, Methylprednisolone, High-dose Ara-C, Cisplatin; PAD—Bortezomib, Doxorubicin, Dexamethasone; R-CD—Rituximab, Cyclophosphamide, Dexamethasone; P-CVP—Rituximab, Cyclophosphamide, Dexamethasone; R-EPOCH—Rituximab, Etoposide, Prednisone, Vincristine, Cyclophosphamide, Doxorubicin; HyperCVAD—Cytarbine, Vincristine, Cyclophosphamide, Doxorubicine, Dexamethasone

analysis. All *p*-values less than 0.05 were considered significant. All data were analyzed using SPSS 20.0 (IBM corp.) statistical software.

Results

General characteristics of the patient group are presented in Tables 1 and 2 and the comparisons between the study and control groups is presented in Table 3. Tests for IgG antibodies were successful in all patients.

CMV serostatus was relatively homogeneous across different B-cell neoplasms despite their glaring clinical diversity which ranged from acute B-ALL and aggressive B-NHL to mature B-chronic lymphocytic leukemia (B-CLL), low-grade B-NHL, and plasmocytoma (Table 1). Biological characteristics they share in common (immunophenotype and somatic mutation profiles) remain preserved in cancerogenesis such that clinical distinctiveness of B-cell neoplasms did not hamper the understanding of their virology.

Most IgG positives were patients with NHL (33/35, 94.3%) followed by B-CLL (8/9, 88.9%), and Hodgkin's disease (HD) (13/17, 76.5%). CMV was least pervasive in multiple myeloma (2/3, 66.7%) but the patients were too few. Low natural incidence of some B-cell disorders resulted in a low number of consecutive patients detected over a short interval of observation. All patients with hairy cell leukemia (2/2), Waldenström's macroglobulinemia (2/2), and non-specified B-cell lymphoma (2/2) were IgG positive. Their numbers were insufficient and were excluded from separate analyses. Positive CMV serology did not correlate among different B-lymphoproliferative diseases (p=0.339).

The study cohorts had markedly different (p<0.001) age and gender structure (Table 3). This required statistical matching to compensate for these discrepancies, after which there remained statistical variance for neither of variables (Table 3). Interestingly, a notable difference in CMV seropositivity emerged between the study group and normal populace after the gender/age matching was performed. The prevalence of CMV infection was significantly higher in the control group (p=0.035), compared to the patient group (Table 3).

Binary logistic regression with B-cell malignancy as dependent and CMV serostatus as independent variable demonstrated that subjects with positive serostatus

Table 3 Statistical information on patient and control groups prior and after matching for age and gender

Cohort characteristics	Unadjusted			Matched cases	i	
	Patients	Controls	<i>p</i> -value	Patients	Controls	<i>p</i> -value
Subjects (N)	83	259	N/A	83	77	N/A
Mean age (yr.) range	49.45 20-73	41.79 20–86	< 0.001*	49.45 20-73	48.05 22–86	0.551*
Gender (N, %)						
Male	43 (51.8%)	73 (28.2%)	< 0.001 [†]	43 (51.8%)	29 (37.7%)	0.072 [†]
Female	40 (48.2%)	186 (71.8%)		40 (48.2%)	48 (62.3%)	
CMV IgG (N, %)						
Yes	75 (90.4%)	214 (82.6%)	0.09 [†]	75 (90.4%)	76 (98.7%)	0.035^{\dagger}
No	8 (9.6%)	45 (17.4%)		8 (9.6%)	1 (1.3%)	

^{*}Mann-Whitney UTest

[†] Fisher's Exact Test

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Table 4 Provisional data on CMV seropositivity in summarized incidence rates of B-lymphoid neoplasms around the world

Population*	Representative CMV (%) seropositivity	B-neoplasms (merged rates yr ⁻¹ /10 ⁵)**	<i>p</i> -value [†]	Refs
The US	44	44.72	N/A	Chihara et al. [55] SEER Program [56]
US-born Asians	75	18.6	< 0.05	Clarke et al. [52] Li et al. [53]
Europe	70	25.44	< 0.05	Zuhair et al. [36]
Middle East [¶] (Iraq, Jordan, S. Arabia)	90	9.12	< 0.05	Yaqo et al. [57]
Sub-Saharan Africa	~92	7.2	< 0.05	Tomoka et al. [81]
Hong Kong	> 90	11.49	< 0.05	Bassig et al. [58]
East Asia (China, Japan)	~90	8.72	< 0.05	Chihara et al. [55]

^{*}Males and females were combined. Worldwide population means of CMV seroprevalence by the International Agency for Research on Cancer (IARC) [35] were relied on in comparisons with annual rates of B-cell maladies. We made use of standardized population-based cancer registry records in Chihara et al.[55]

Remark: The differences in study populations, high false discovery rates and dissimilar calendar periods of observation in the reports used affected the calculations. These should be viewed with caution

Table 5 Crude incidence rate estimates of key B-cell malignancies: aggregate rates by country, race, and ethnicity

Country (race/ethnicity)	Lymphoid ma (crude annua		nates/10 ⁵ populat	ion)		Summed crude aggregate
	B-NHL*	HD	B-ALL	B-CLL	PCM	estimates
The US	27.4	3.23	1.69	6.3	6.1	44.72
The US-born Asians	11.8	1.28	0.7	1.8	3.2	18.6
Europe (wgt. mean)	11.6	3.58	0.68	4.88	4.7	25.44
Middle East	4.9	1.45	0.56	0.71	1.5	9.12
Sub-Saharan Africa	5.5	0.7	0.35	0.36	0.29	7.2
Hong Kong	6.68	0.62	1.28	0.6	2.31	11.49
East Asia (China, Japan)	4.8	0.53	0.96	1.13	1.3	8.72

^{*}B-NHL B-cell non-Hodgkin Lymphoma; HD Hodgkin's disease; B-ALL B-cell acute lymphoblastic leukemia; B-CLL B-cell chronic lymphocytic leukemia; PCM plasma cell myeloma

were \sim 7 times less likely (OR, 0.067; 95% CI, 0.016 to 1.150) to have a B-cell malignancy relative to seronegatives. The difference was not significant (p=0.067), but near the conventional level of significance (0.05).

The results pointed to a potential protective effect that CMV may proffer against B-cell dyscrasia. In order to investigate our evidence on a much larger scale, we compared annual incidence rates of B-cell neoplasms to CMV prevalences in 74 countries for which these variables were available (Fig. 1A–D). Interestingly, a significant negative correlation between CMV pervasiveness and the incidence of all clinical types of B-cell malignancies was observed the world over (Fig. 1A; Spearman $\rho=-0.625,\ p<0.001$). Similarly, an inverse association was evidenced separately for three different B-cell malignancies: HD (Fig. 1B; Spearman $\rho=-0.618,\ p<0.001$), non-Hodgkin lymphomas (Fig. 1C; Spearman $\rho=-0.617,\ p<0.001$), and myeloma (Fig. 1D; Spearman $\rho=-0.633,\ p<0.001$), separately.

^{**}The crude numbers of extracted incidence rates of B-cell neoplasms were merged together in the middle column). Compounded incidences were preferred as a proviso for each world zone. Thus, rough rates depart from factual ones and should be taken as broadly consultative. Also, racial groups were collected under different protocols, excepting the Surveillance, Epidemiology, and End Results (SEER) 13 Incidence Database. Withal, unequal pathological categorization according to diagnosis (or cellular origin) and sub-classification criteria for B-cell leukemia/lymphoma in particular, varied from country to country introducing further biases. This hampered estimation of summated incidence rates. Consequently, workout of regional rates of B-cell neoplasms do not sum-up to estimates reported in the literature used

[†] P-values were two-sided using the t-test. They reflect coupling strength between CMV seropositivity and incidence rate of B-cell neoplasms in the US versus other countries

¹ Age-adjusted incidence rates refer to patients > 70 years of age. This age group had the longest exposure to CMV and could not be strictly compared to global SEER data

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These results support the reality of oncoprotection by the chronic CMV infection against B-lymphomagenesis irrespective of a clinical form of a B-cell neoplasm.

Discussion

This is the first study reporting on the current estimate of CMV infection in Serbian hemato-oncological patients and healthy controls. Also, our clinical results are supported by the worldwide survey of relevant data. Together, they offer the first insight into a possible connection between the chronic CMV infection and B-cell neoplasms, hinting at an oncoprotection conferred by this virus on its host.

CMV seroprevalences in patients with hematological malignancies

CMV seroprevalence varies in published studies on patients with hematological malignancies. Virus prevalence in our patient cohort (90.4%) places the Republic of Serbia among the most CMV-permeated populations in the world [37–44]. Much lower seroprevalence (70%) of anti-CMV IgG was reported in a multicenter cohort of Swedish patients (Re: Mission, NCT01347996, www.clinicaltrials.gov [45]. The *lowest* CMV infestation was reported in landmark studies from the US [46, 47], a highly developed country with one of the largest incidence rates of B-cell disorders.

In studies on HCT recipients [3], and B-CLL patients [48], females were significantly more CMV seropositive. Similar to our clinical population, in Brazilian patients with various hematologic disorders females were more CMV seropositive than males albeit not significantly [37]. On the contrary, Sudan females with leukemia were less seropositive for CMV than males [49]. Marchesi et al. [39] reported largest prevalence of CMV in patients with B-CLL, and multiple myeloma which is similar to the present findings.

Inverse association between CMV seroprevalence and incidence of B-cell neoplasms across the globe

There is a stark difference in annual incidences of B-lymphoid malignancies between Western and Eastern countries [50, 51]. We try to draw attention to an inverse association between the annual age-adjusted incidences of B-cell malignancies and the spread of CMV seropositivity at a global level (Tables 5 and 6, Fig. 1A–D). Seroprevalence in presumably epidemiologically unrelated

communities was frequently lower in patients with B-cell and even in other malignancies (acute myeloid leukemia, AML; chronic myeloid leukemia—CML) than that reported in voluntary blood/organ donors and in the general population [36]. This difference is explainable if chronic CMV infection conferred a degree of protection on its immunocompetent host against B-cell malignancies. This is consistent with the evidence in the current work where healthy controls were significantly more CMV seropositive (p=0.035; Table 3) than patients with B-cell malignancies. A potential explanation might be an increase in resistence against B-cell neoplasia fostered by primary CMV infection.

As the prevalence of CMV infection recedes across the populations, corresponding annual incidence of B-cell diseases tends to increase. For decades, incidence of lymphoid neoplasms has been globally increasing across age strata and sex. This may signify a gradual loss of protection provided by the latent CMV infection which is being globally eroded by steadily improving economic prowess and modern access to health care.

A racial/ethnic background is related to SES [52, 53]. The difference in incidence of B-lymphoid malignancies between the US and Japan is elevated, 2.5- to fivefold. The largest proportional difference between the US and Japan was in B-CLL (the US, 24.1%; Japan, 3.2%) [52]. Annual incidence rates of B-cell neoplasms in the US-born Asians/Pacific islanders are generally intermediate to those in the US whites and East Asians; exactly parallel trend is observed in their respective CMV seroprevalences. The incidence rates of B-cell neoplasms tend to negatively parallel the prevalence of CMV seropositivity in respective populations worldwide (Fig. 1A-D). HD and B-NHL showed the largest difference in annual incidences between the US and East Asian countries. The SES correlates with trends in age-standardized incidences of B-lymphoid disorders and is also associated with CMV infection around the world. Seroprevalence of CMV decreased in pregnant women in Ishikawa Prefecture (Japan) from 93.2% to 66.7% over the period between 1980 and 1998 and in parallel with the increase in SES [54]. Of note, age-adjusted incidence of lymphoid malignancies in Japan increased significantly as opposed to no significant annual percent change in the US (Japan, +2.4%; US, +0.1%) [55]. This may be a consequence of growing SES in Japan and the consequent drop in CMV infection there.

(See figure on next page.)

Fig. 1 The scatter charts present country specific CMV prevalence (mean) plotted against estimated age-standardized (world) annual incidence rates (per 100,000) of microscopically verified cases of B-cell types of cancer in 74 countries (blue circles) [35, 36, 69]. **A)** B-cell malignancies (all types) (Spearman $\rho = -0.625$, p < 0.001), **B)** Hodgkin's disease (Spearman $\rho = -0.618$, p < 0.001), **C)** non-Hodgkin lymphomas (Spearman $\rho = -0.618$, p < 0.001), and **D)** multiple myeloma (Spearman $\rho = -0.633$, p < 0.001) in 2020. The inverse relationship between viral pervasiveness and the annual incidence rate of hematologic malignancies is highly significant for all (**A**) and each individual B-cell cancer type (**C-D**)

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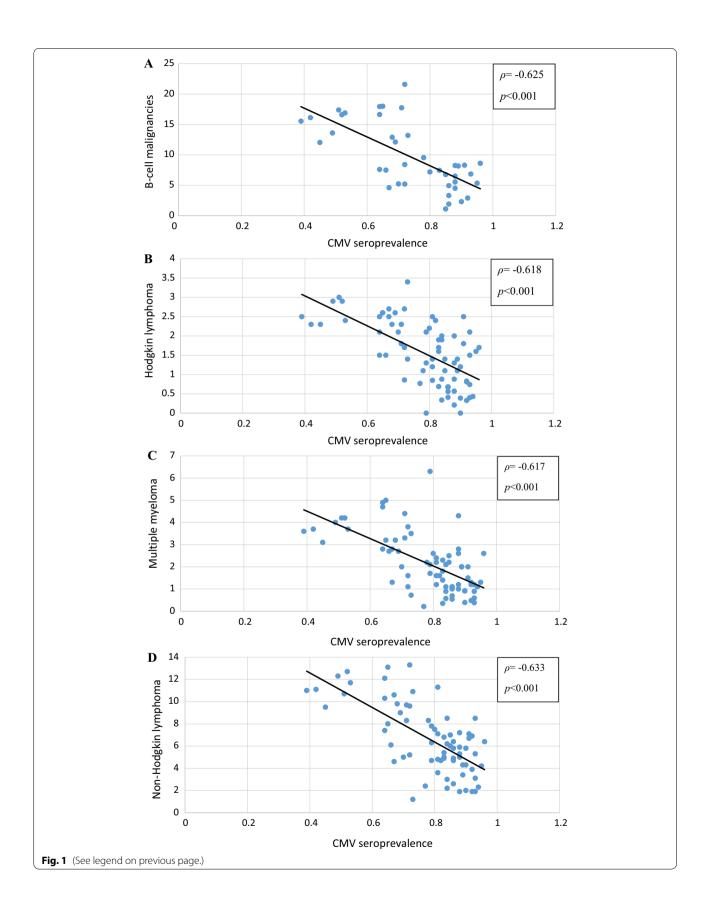


 Table 6
 Country-specific CMV seroprevalence in patient cohorts compared to matched blood/organ donors and healthy general populations

HD, MM, CLL HD, MHL, HD, NHL, Gic malignancies AT- cell types) ALL (B- & T-cell ALL (B- & T-cell Ogic malignancies					
ALL AL, NHL, HD, MM, CLL AL, NHL, HD, MM, CLL ALL ALL (children) Hematology patients (various) ALL, MM, HD, NHL, Hematolgic malignancies (various) ALL (B- & T- cell types) Hematologic malignancies	Cytomegalovirus seropositivity (CMV ⁺)	sitivity (CMV ⁺)		Transplant type and	References
ALL AL, NHL, HD, MM, CLL 835 AL, NHL, HD, MM, CLL 680 ALL (children) Hematology patients (various) ALL, MM, HD, NHL, (various) ALL, MM, HD, NHL, (various) ALL (B- & T- cell types) Hematologic malignancies 127 types) Hematologic malignancies 72 (4)	Patients N, (CMV ⁺ %) (unless otherwise noted)	Country specific blood/ organ donors CMV ⁺ , mean [36]	Country specific general popul. CMV ⁺ , mean [36]	support medication	
ALL AL, NHL, HD, MM, CLL 835 AL, NHL, HD, MM, CLL 952 AL (children) Hematology patients (various) ALL, MM, HD, NHL, 108 ALL (B- & T- cell types) 644 ALL (B- & T- cell types) 7564 ALL (B- & T- cell types) 7564 ALL (B- & T- cell types) 77 (ypes) ALL (B- & T- cell types) 78 ALL (B- & T- cell types) ALL (H- & T- cell types) 79 ALL (H- & T- cell types) 70 ALL (H- & T- cell types) 70 ALL (H- & T- cell types) 71 ALL (H- & T- cell types) 72 (ALL NHL, MM, CLL 74 ALL NHL, MM, CLL 75 ALL NHL, MM, CLL 76 ALL NHL, MM, CLL 76 ALL NHL, MM, CLL 77 ALL NHL, MM, CLL 76 ALL NHL, MM, CLL 76 ALL NHL, MM, CLL 77 ALL NHL, MM, CLL 76 ALL					
AL, NHL, HD, MM, CLL AL, CLL, NHL ALL (children) Hematology patients (various) ALL, MM, HD, NHL, (various) ALL, MM, HD, NHL, (various) ALL, MM, HD, NHL, (various) ALL (B- & T- cell types) ALL (H- & T- cell types) Hematologic malignancies ALL (H- & T- cell types) ALL (H- & T- cell types) Hematologic malignancies 72 (4)	1 359 (48%)	0.67	0.64	Allo-HCT, GvHD prophylaxis	Kollman et al. 2001 [46]
ALL, CLL, NHL 680 ALL (Children) Hematology patients 470 (various) ALL, MM, HD, NHL, 20 (various) ALL, MM, HD, NHL, 20 (various) ALL, MM, HD, NHL, 20 (various) ALL (B- & T- cell types) 564 ALL (B- & T- cell types) 188 ALL (B- & T- cell types) 188 Hematologic malignancies 72 (4		0.67	0.64	Allo-HCT, GvHD prophylaxis	Nichols et al. 2002 [47]
ALL, CLL, NHL AL (children) Hematology patients (various) ALL, MM, HD, NHL, 20 (4 Hematolgic malignancies (various) 33 ALL (B-&T- cell types) ALL (B-&T- cell types) ALL (B-&T- cell types) ALL (B-&T- cell types) Hematologic malignancies 72 (4 Hematologic malignancies	107 (57%)	0.67	0.64	Auto-HCT, BCNU, BEAM, melphalan alone	Fassas et al. 2001 [82]
ALL (B-&T-cell types)	680 (52.4%)	29.0	0.64	Chemo radiation therapy	Nguyen et al. 2001 [83]
AL (children) Hematology patients (various) ALL, MM, HD, NHL, Hematolgic malignancies (various) ALL (B- & T- cell types) ALL (H- & T- cell types)	952 (47.5%)	0.63	09.0	Allo-HCT, GvHD prophylaxis	Lee et al. 2007 [84]
Hematology patients (various) ALL, MM, HD, NHL, Hematolgic malignancies N/A (various) 33 ALL (B-&T-cell types) 564 ALL (B-&T-cell types) 188 Poor risk ALL (B-&T-cell Types) ALL NHL, MM, CLL 165 Hematologic malignancies 72 (4	144 (55.7%)	0.67	0.64	Allo-HCT, GvHD prophylaxis	Behrendt et al. 2009 [85]
ALL, MM, HD, NHL, Hematolgic malignancies (various) 3.3 ALL (B- & T- cell types) 564 ALL (B- & T- cell types) 188 Poor risk ALL (B- & T-cell types) ALL, NHL, MM, CLL Hematologic malignancies 72 (4		0.91	0.89	Z/A	De Matos et al. 2011 [37]
Hematolgic malignancies N/A (various) ALL (B- & T- cell types) 564. ALL (B- & T- cell types) 188. Poor risk ALL (B- & T-cell 127 types) ALL, NHL, MM, CLL 165 Hematologic malignancies 72 (4	20 (82.5%)	0.91	0.89	Allo-HCT, GvHD prophylaxis	Dieamant et al. 2011 [86]
33 ALL (B- & T- cell types) 564 ALL (B- & T- cell types) 188 Poor risk ALL (B- & T-cell 127 types) ALL, NHL, MM, CLL 165 Hematologic malignancies 72 (4		0.91	0.89	HCT, GvHD prophylaxis	Ferrés et al. 2012 [87]
33 ALL (B- & T- cell types) 564 ALL (B- & T- cell types) 188 Poor risk ALL (B- & T-cell 127 types) ALL, NHL, MM, CLL 165 Hematologic malignancies 72 (4					
ALL (B- & T- cell types) 188 Poor risk ALL (B- & T-cell 127 types) ALL, NHL, MM, CLL 165 Hematologic malignancies 72 (4		0.55 (ave.)	0.51		Duval et al. 2010 [88]
Poor risk ALL (B- & T-cell 127 types) ALL, NHL, MM, CLL 165 Hematologic malignancies 72 (4	5) 1 883 (59.7%)	0.62 (ave.)	0.59 (ave.)	Allo-HCT GvHD prophylaxis	Teira et al. 2016 [89]
ALL, NHL, MM, CLL Hematologic malignancies 72 (4		0.63	0.58	MUD-BMT	Cornelissen et al. 2001 [90]
Hematologic malignancies 72 (4	165 (60% donors)	0.66 (ave.)	0.68 (ave.)	Allo-HCT	Marty et al. 2017 [91]
(various)	ancies 72 (48%)	0.43	0.39	Allo-HCT (43 pts.) Chemotherapy (28 pts.)	Fleming et al. 2010 [92]
Sweden & Italy, Gothen- AML 81 (70 burg, Rome	81 (70%)	0.74	0.71	Chemoradio therapy	Bernson et al. [45]

Table 6 (continued)

		-	11.000		-	
Continent, country, or	Hematologic	Cytomegalovirus seropositivity (CMV)	Sitivity (CMV ')		I ransplant type and	Keterences
region (listed West-to- East)	maiignancy	Patients N, (CMV ⁺ %) (unless otherwise noted)	Country specific blood/ organ donors CMV ⁺ , mean [36]	Country specific general popul. CMV ⁺ , mean [36]	support medication	
Sweden, Germany, The Netherlands, Italy, France	ALL (B- &T- cell types)	3 539 (55%)	0.62 (ave.)	0.58 (ave.)	Allo-BMT & HCT	Ljungman et al. 2003 [2]
EBMT member centers (whole registry cohort)	ALL, AML, HD, NHL	40 306 (53.9%)	0.62 (ave.)	0.58 (ave.)	Allo- & auto-HCT	Ljungman & Brand 2007 [3]
Sweden, Spain, UK, France, Italy, The Netherlands, Poland, Germany	ALL, LY	31 669 (59.8%)	0.62 (ave.)	0.58 (ave.)	Allo-HCT	Ljungman et al. 2014 [93]
48 transpl. centers (Europe, MENA, South Africa)	ALL, NHL, HD	165 (81.4%)	0.77 (ave.)	0.75 (ave.)	Allo-BMT	Ljungman et al. 2002 [94]
US (CIBMTR), Canada, UK, Spain, Sweden, Saudi Ara- bia, New Zealand, Japan	Childhood ALL (B- & T-cell)	980 (53%)	69.0	0.65	MRD, URD, UCBT, PB-HCT from URD	Mehta et al. 2019 [95]
France, USA, UK, Germany, Czech Republic, Israel	AML (age > 50)	3 398 (65%)	0.60 (ave.)		MUD HCT	Rubio et al. 2016 [96]
11 European countries with Israel and Turkey	Childhood B-precursor ALL	140 (41%)	0.63 (ave.)	0.65 (ave.)	Allo-HCT	Dalle et al. 2018 [97]
France, Israel, Spain, Germany, Belgium, The Netherlands, UK, Poland (EBMT)	B-ALL (age > 60)	126 (59.3%)	0.61	0.57	RIC allo-HCT	Roth-Guepin et al. 2017 [98]
Belgium, Brussels	n.m (9 children, 7 adults)	16 (35%)	0.56	0.52	Allo-HCT	Debaugnies et al. [99]
Denmark, Copenhagen	Childhood ALL & adults	118 (54%)	09.0	N/A	Allo-HCT	Kielsen et al. 2018 [100]
The Netherlands, Utrecht	ALL, NHL, HD	101 (50.6%)	0.57	0.53	Allo-BMT	Meijer et al. 2002 [101]
The Netherlands, Rot- terdam	ALL, NHL, MM	47 (66%)	0.57	0.53	Allo-HCT (sibling)	Broers et al. 2000 [102]
Poland, Wroclaw	ALL, LY	26 (78%)	0.7	99.0	HCT	Jaskula et al. 2015 [103]
Czech Republic, Brno	Childhood & adolescent ALL, NHL, HD	104 (37.6%)	n.m	0.42 (healthy agematched control)	Conventional chemo- therapy	Michálek & Horvath 2002 [104]
Germany, Russian Federation, Hamburg, St. Petersburg	ALL, NHL	54 (39%)	0.62	0.57	Allo-HCT	Kröger et al. 2001 [105]
Russian Federation, Moscow	AML, Mantle cell lym- phoma	183 (45.9%)	0.74	0.7	Allo-HCT	Vdovin et al. 2016 [106]
Croatia, Zagreb	ALL, NHL, MM, HD, CLL	47 (77%)	0.83	0.8	Allo-HCT	Peric et al. 2018 [107]

Table 6 (continued)

Continent, country, or	Hematologic	Cytomegalovirus seropositivity (CMV ⁺)	sitivity (CMV ⁺)		Transplant type and	References
region (listed West-to- East)	maiignancy	Patients N, (CMV ⁺ %) (unless otherwise noted)	Country specific blood/ organ donors CMV ⁺ , mean [36]	Country specific general popul. CMV ⁺ , mean [36]	support medication	
Hungary, Szeged	Ιλ	224 (75%)	0.87	0.84	Auto-HCT Chemotherapy	Piukovics et al. 2017 [1]
Italy, Rome	AML	52 (93%)	0.76	0.73	Chemoradio therapy	Capria et al. 2010 [40]
Italy, Rome	\ <u></u>	327 (93%)	0.76	0.73	Auto-BMT & Auto-HCT	Marchesi et al. 2015 [39]
Italy, Milan, Udine, Ber- gamo, Ancona, Alessandria	B-cell lymphoma	265 (70%)	0.76	0.73	Allo-HCT	Mariotti et al. 2014 [108]
Germany, France, Finland	ALL (B- & T-cell)	5 158 (60%)	0.58	0.54	Allo-HCT	Schmidt-Hieber et al. 2013 [109]
Serbia, Belgrade-Srem- Šumadija	ALL, HD, WD, CLL, MM,	83 (88%)	N/A	0.9	Allo-HCT, Chemoradio therapy	this work
Spain, Barcelona	AL, NHL, CLL, MM, HD/ST	150 (66%)	0.72	69.0	Allo-HCT-RIC GvHD prophylaxis	Piñana et al. 2010 [110]
Middle East North Africa Region	on					
Kingdom of Saudi Arabia, Jeddah	ALL, NHL, HD, MM, CLL	1 252 (95.76%)	0.89	0.88	Chemothrapy HCT	Zaidi et al. 2019 [41]
Kingdom of Saudi Arabia, Jordan, Riyadh, Irbid	ALL (children)	82 (1.22%)	0.85	0.87	Allo-HCT UCBT GvHD prophylaxis	Al-Sweedan et al. 2017 [111]
Kingdom of Saudi Arabia, Riyadh	AL (children)	73 (68%)	0.89	0.88	Allo-cord blood HCT	Al-Hajjar et al. 2011 [112]
Jordan, Amman	AL (children)	72 (31%)	0.83	0.85	Auto-HCT	Hussein et al. 2015 [113] & Al Mana et al. 2019 [114]
Israel, Tel Aviv, Petah Tikva	AL, LY	121 (61%)	0.75	0.72	Allo-HCT GvHD prophylaxis	Cohen et al. 2015 [115]
Iran, Tehran, Rasht	ALL, AML, NHL, MM & vari- 126 (ous disease	126 (97.6%)	96:0	0.95	Allo-HCT GvHD prophylaxis	Valadkhani et al. 2016 [116]
Iran, Mashhad	Voluntary blood donors	1 000 (99.2%)	96:0	0.95	N/A	Safabakhsh et al. 2013 [117]
Iran, Urmia	End-stage renal disease (immunodeficiency)	65 (77.4%)	96.0	0.95	Pre-transplant hemodi- alysis	Sepehrvand et al. 2010 [118]
Iran, Shiraz, Tehran,	Leukemia (unspecified)	6 (100%)	96.0	0.95	Allo-BMT	Behzad-Behbahani et al. 2004 [119]
Iran, Tehran, Shiraz	AML, Thalassemia, CML, AA, ALL	26 (100%)	96:0	0.95	ВМТ	Ziyaeyan <i>et. al.</i> 2006 [120]
Tunisia, Sousse, Sfax	ALL	39 (90%)	0.94	0.93	Chemotherapy (different phases)	Handous et al. 2020 [121]

Table 6 (continued)

Continent, country, or	Hematologic	Cytomegalovirus seropositivity (CMV ⁺)	sitivity (CMV ⁺)		Transplant type and	References
region (listed West-to- East)	malignancy	Patients N, (CMV ⁺ %) (unless otherwise noted)	Country specific blood/ organ donors CMV ⁺ , mean [36]	Country specific general popul. CMV ⁺ , mean [36]	support medication	
Egypt, Alexandria	Voluntary blood donors	88 (96.6%)	0.94	0.93	N/A	Gawad et al. 2016 [122]
Egypt, Cairo	AML & ALL	28 (39%, active CMV)	0.94	0.93	BMT	Zekri et al. 2004 [123]
Egypt, Cairo	B-ALL (40) T-ALL (10) (children & adolesc.)	40 (36% CMV DNA/ serum)	30 (46.7% CMV DNA control)	0.93	Consolidation Tx Salvage Tx	Loutfy et al. 2017 [124]
Australia, India						
Australia, Victoria	AL, B-NHL, HD, CLL, MM	28 (88%)	69'0	0.65	Conventional-dose chemotherapy Auto-HCT	Ng et al. 2005 [38]
Australia, Sydney	ALL, NHL, MM	103 (63%)	69.0	0.65	Allo-HCT, MUD-HCT	George et al. 2010 [125]
India, Vellore	Malignant & non-malignant diseases Patients: Donors:	463 (97.4) 403 (84.8%)	0.88	0.86	Allo-HCT	Devasia et al. 2018 [126]
East Asia						
Malaysia, Australia. Kuala Lumpur, Melbourne	ALL $N = 71$, AML $N = 6$, med. age: 28 yr	77 (73%)	0.78 (ave.)	0.75 (ave.)	Chemoradiotherapy	Azanan et al. 2016 [127]
China, Guangzhou province	B-ALL, B-NHL	156 (86%)	0.92	0.9	Allo-HCT (intensified condit.)	Xuan et al. 2012 [42]
China, Beijing	ALL, CML, MDS, AA, NHL	60 (87%)	0.92	0.9	Allo-HCT	Du et al. 2007 [128]
Taiwan, Kaohsiung, Taipei	AL, NHL	117 (91.8%)	0.95	0.93	Allo-HCT	Liu et al. 2012 [43]
Japan, Tokyo	Childhood AL	184 (81%)	9.76	0.72	UCBT	Tomonari et al. 2008 [44]
Japan, Fukuoka	Childhood ALL	101 (72%)	9.76	0.72	Allo-HCT	Inagaki et al. 2015 [26]

donor; URD, unrelated donor; UCBT, umbilical cord blood transplantation; PB-HCT, peripheral blood HCT; GvHD, graft vs. host disease; BMT, bone marrow transplantation; MUD, matched unrelated donor; BCNU, BEAM, and T-cell); HD, Hodgkin's disease; MM, multiple (plasma cell) myeloma; CLL, chronic lymphocytic leukemia; LY, lymphoid neoplasms (HD, NHL of B- and T-cell types); WD, Waldenström's disease; myeloid neoplasms (AML; CML, chronic myeloid leukemia; MDS, myelodysplastic syndrome; MPN, myeloproliferative neoplasm); AA, aplastic anemia; allo-HCT, allogeneic hematopoietic cell transplant; auto-HCT, autologous HCT; MRD, matched related n.m. not mentioned; N/A not applicable; AL denotes acute leukemia (AML and B- and T-cell ALL together); AML, acute myeloblastic leukemia; ALL, acute lymphoblastic leukemia; NHL, non-Hodgkin lymphoma (B- and melphalan indicate myeloablative protocols; CIBMTR, Center for International Bone Marrow Transplant Research; MEN4, Middle East North Africa; RIC, reduced intensity chemotherapy HCT; ST, solid tumors

is presented West-to-East. In most studies with mixed disease settings the prevalence of CMV seropositivity in subsets of B-cell diseases was not available. This perturbed the estimates of CMV seropositivity of interest, point out the studies evidencing a reduced CMV protection against lymphoproliferation in the patients reported. CMV seroprevalence in patients with hematological diseases across racial and ethnic groups divisions Studies with significantly lower CMV seropositivities in auto- and allo-HCTed patients, as compared to healthy donors and country-specific CMV prevalences, have seropositivity values indicated in bold. This was to corresponding country means [36]. This may signal a certain degree of oncoprotection secured to chronic carriers of latent virus. Endemicity of CMV seems to depend on SES defined factors and correlates with the affecting the veracity of presented data. Notwithstanding partially inadequate representation of patient populations, the prevalence of CMV in hematologic malignancies shown remains mostly lesser than the incidence rate of malignant B-cell diseases across distant domains (Tables 4 and 5, Fig. 1 A-D) Janković et al. Virology Journal (2022) 19:155 Page 12 of 17

Global disease burden reports [36, 52, 53, 55-72] suggest a significant inverse correlation between overall estimates of CMV seropositivity and the age-standardized and population-based incidence rate of B-cell cancers (Tables 4 and 6, Fig. 1A-D). Cytomegalovirus infection decreases as contemporary economy improves and affluence is gained across societal strata. Reduced rates of CMV primoinfection in developed countries may be the cause of an increased risk of contracting a B-cell malignancy. By contrast, high CMV prevalence in countries with adverse economic conditions, appears to mitigate the risk of B-lymphoproliferative disease. In populations where the prevalence of CMV declines an oncoprotective effect of CMV subsides such that an increased annual incidence rate of B-cell cancer is observed worldwide (Spearman $\rho = -0.625$, p < 0.001). However, some other factor(s) may operate along with CMV infection influencing the global correlation between increasing incidence of B-cell malignancies, improving SES, and reduced country-specific prevalence of CMV infection.

Clinical and in vitro experimental evidence supporting oncoprotection by CMV

Cytomegalovirus seroprevalence was higher in the controls than in our patients with B-cell malignancies (Table 3; p = 0.035). This argues against the promotive contribution of CMV in B-cell lymphomagenesis.

Evidence in favor of viral repression of the transformation process in cancer cells has been reported [73]. CMV inhibits the migratory capacity of mesenchymal breast cancer cell lines MDA-MB-231 and SUM1315 [19]. Mice xenografted with CMV-infected HepG2 cells were reported to manifest limited to no tumor growth, as opposed to an unbridled tumor expansion in placebotreated mice [74]. A runaway tumor growth was inhibited by restricting STAT3 activation, as well as by activation of the intrinsic apoptotic pathway [74, 75]. Apoptosis was also registered in the lung tissue of xeno-engrafted mice where HepG2 cells infected with human CMV were administered subcutaneously [74]. Erlach et al. [21, 22] proposed an innate anti-tumor mechanism elicited by murine CMV infection involving apoptosis of a liveradapted clonal variant of B-cell lymphoma. The murine CMV infection had a highly suppressive effect on lymphoma cells even without infecting them, resulting in a significant survival benefit. Erkes et al. [76] also demonstrated clearance of tumors in a mouse melanoma model after CMV was inoculated into growing neoplasm. Also, an inhibiting effect of CMV glycoprotein B on breast cancer cell migration was recently documented by Yang et al. [20].

Anti-tumor effects of CMV infection were tentatively supported by reports of reduced relapse rates in patients

with CMV reactivation early after allogeneic HCT for acute leukemia and NHL [23–27]. Changes within the immune system caused by CMV suggest a possible virus-*vs*-leukemia phenomenon [18] analogous to graft-*vs*-leukemia effect in B-CLL [77].

A study which screened neonatal Guthrie blood spots for CMV did not find that the CMV positives contracted B-ALL more often later in life [78]. MacKenzie et al. have screened common ALL patients and controls for presence of various herpesviruses, but were in doubt that a herpesvirus is an etiological agent in B-ALL [79]. Another study analyzed herpesvirus DNA in Guthrie cards and found no trace of EBV or HHV-6 but CMV presence has not been assessed [80]. Evidence garnered from these studies substantiates the assumption that CMV may forestall initiation of B-cell neoplasms.

A major strength of the present exploration is the use of a nationally representative sample to estimate CMV seroprevalence in the Republic of Serbia. Noteworthy limitations of our work are its retrospective nature and an artefact from a small sample size. Furthermore, a passive take of donor's IgG antibodies cannot be entirely excluded. This drawback to the study was mitigated by lower CMV seropositivity among blood transfusion-treated patients as compared to healthy controls.

Conclusions

Conclusively, we present first set of data on CMV sero-prevalence based on a sample of B-cell derived malignancies in Serbia. Also, we provide evidence that prevalences of CMV are strongly inversely associated with the annual incidence rates of malignant B-cell disorders the world over. This is suggestive of a possible protective effect of CMV against the profligate B-cell growth. The cellular niche may be less favourable for initiation of B-lymphomagenesis in chronic carriers of CMV. Prospective work with a larger study size of cell lineage-specific patient cohorts across clinical and histological lymphoma subtypes may be helpful in clarifying dilemmas regarding anti/pro tumoral activity of CMV.

Abbreviations

AA: Aplastic anemia; AL: Acute leukemia; ALL: Acute lymphoblastic leukemia; Allo-HCT: Allogeneic hematopoietic cell transplantation; AML: Acute myeloid leukemia; Auto-HCT: Autologous hematopoietic cell transplantation; BMT: Bone marrow transplant; CIBMTR: Center for International Bone Marrow Transplant Research; CLL: Chronic lymphocytic leukemia; CML: Chronic myeloid leukemia; CMV: Cytomegalovirus; GLOBOCAN: Global Cancer Observatory; GVHD: Graft-vs-host disease; HCT: Hematopoietic cell transplantation; HD: Hodgkin's disease; IARC: International Agency for Research on Cancer; LY: Lymphoid neoplasms; MDS: Myelodysplastic syndrome; MENA: Middle East North Africa; MM: Multiple myeloma; MPN: Myeloproliferative neoplasm; MRD: Matched related donor; MUD: Matched unrelated donor; n.m.: Not mentioned; N/A: Not applicable; NHL: Non-Hodgkin lymphoma; PB-HCT: Peripheral blood HCT; PCM: Plasma cell myeloma; PTLD: Posttransplant lymphoproliferative disorder; RIC: Reduced intensity chemotherapy; SEER: Surveillance, Epidemiology, and

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End Results; SES: Socioeconomic status; ST: Solid tumors; UCBT: Umbilical cord blood transplantation; URD: Unrelated donor; WD: Waldenström's disease.

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Author contributions

All authors contributed to the study conception and design. The first draft was written by MJ. MJ and AK conducted laboratory investigation, performed the experiments, collected laboratory information and, interpreted and analyzed the data. Coordination and supervision of data and sample collection was performed by MT, BM, IĐ and VS. IS analyzed and interpreted the data. TJ designed the methodology, managed resource acquisition, project administration and overall supervision, and was responsible for the research activity planning and execution. MB, JP, NM and SFV collected patient record data and performed laboratory experiments. All authors read, critically revised, and approved the final manuscript.

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Availability of data and materials

The datasets generated and/or analysed during the current study are not publicly available due to safeguarding patient anonymity, but are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

This work was performed in accordance with principles of 1964 Helsinki Declaration and its later amendments. Approval was granted by the University Clinical Centre of Serbia, University of Belgrade Ethical Review Board and Faculty of Medicine. Guidelines of Good Clinical Practice were observed. Document of informed approval was obtained from all consenting participants included in the study. The security and privacy of patient's health was not violated in this study.

Consent for publication

Not applicable.

Competing interests

The authors declare that research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Piukovics K, Terhes G, Gurbity-Pálfi T, Bereczki Á, Rárosi F, Deák J, et al. Cytomegalovirus infection in patients with haematological diseases and after autologous stem cell transplantation as consolidation: a single-centre study. Ann Hematol. 2017;96:125–31. https://doi.org/10. 1007/s00277-016-2831-7.
- Ljungman P, Brand R, Einsele H, Frassoni F, Niederwieser D, Cordonnier C. Donor CMV serologic status and outcome of CMV-seropositive recipients after unrelated donor stem cell transplantation: an EBMT megafile analysis. Blood. 2003;102(13):4255–60. https://doi.org/10.1182/blood-2002-10-3263.
- Ljungman P, Brand R. Factors influencing cytomegalovirus seropositivity in stem cell transplant patients and donors. Haematologica. 2007;92:1139–42. https://doi.org/10.3324/haematol.11061.
- Lachmann R, Loenenbach A, Waterboer T, Brenner N, Pawlita M, Michel A, et al. Cytomegalovirus (CMV) seroprevalence in the adult population of Germany. PLoS ONE. 2018;13(7):e0200267. https://doi.org/10.1371/ journal.pone.0200267.
- Cannon MJ, Schmid DS, Hyde TB. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. Rev Med Virol. 2010;20:202–13. https://doi.org/10.1002/rmv.655.
- Marshall GS, Rabalais GP, Stewart JA, Dobbins JG. Cytomegalovirus seroprevalence in women bearing children in Jefferson County. Kentucky Am J Med Sci. 1993;305:292–6. https://doi.org/10.1097/00000 441-199305000-00005.
- Cheeran MC-J, Lokensgard JR, Schleiss MR. Neuropathogenesis of congenital cytomegalovirus infection: disease mechanisms and prospects for intervention. Clin Microbiol Rev. 2009;22(1):99–126. https://doi.org/10.1128/CMR.00023-08.
- Michaelis M, Doerr HW, Cinatl J. The story of human cytomegalovirus and cancer: increasing evidence and open questions. Neoplasia. 2009;11(1):1–9. https://doi.org/10.1593/neo.81178.
- Cinatl J Jr, Cinatl J, Vogel JU, Rabenau H, Kornhuber B, Doerr HW. Modulatory effects of human cytomegalovirus infection on malignant properties of cancer cells. Intervirology. 1996;39(4):259–69. https://doi. org/10.1159/000150527.
- Cinatl J, Vogel JU, Cinatl J, et al. Long-term productive human cytomegalovirus infection of a human neuroblastoma cell line. Int J Cancer. 1996;65:90–6. https://doi.org/10.1002/(SICI)1097-0215(19960103)65:1% 3c90:AID-IJC16%3e3.0.CO;2-M.
- Nelson JA, Fleckenstein B, Jahn G, et al. Structure of the transforming region of human cytomegalovirus AD169. J Virol. 1984;49:109–15. https://doi.org/10.1128/JVI.49.1.109-115.1984.
- Boldogh I, Huang ES, Rady P, et al. Alteration in the coding potential and expression of H-ras in human cytomegalovirus–transformed cells. Intervirology. 1994;37:321–9. https://doi.org/10.1159/000150396.
- Shen Y, Zhu H, Shenk T. Human cytomagalovirus IE1 and IE2 proteins are mutagenic and mediate "hit-and-run" oncogenic transformation in cooperation with the adenovirus E1A proteins. Proc Natl Acad Sci USA. 1997;94:3341–5. https://doi.org/10.1073/pnas.94.7.3341.
- Doniger J, Muralidhar S, Rosenthal LJ. Human cytomegalovirus and human herpesvirus 6 genes that transform and transactivate. Clin Microbiol Rev. 1999;12(3):367–82 (PMID: 10398670).
- Francis SS, Wallace AD, Wendt GA, et al. In utero cytomegalovirus infection and development of childhood acute lymphoblastic leukemia. Blood. 2017;129:1680–4. https://doi.org/10.1182/ blood-2016-07-723148.
- Wiemels JL, Talbäck M, Francis S, Feychting M. Early infection with cytomegalovirus and risk of childhood hematologic malignancies. Cancer Epidemiol Biomarkers Prev. 2019;28(6):1024–7. https://doi.org/10.1158/ 1055-9965.EPI-19-0044.
- 17. Éboli LPDCB, Tannuri ACA, Tannuri U. Seropositivity for cytomegalovirus and PCR-EBV monitoring: Protective factors for posttransplant lymphoproliferative disorder in pediatric liver transplant. Pediatr Transplant. 2022;2022:e14226. https://doi.org/10.1111/petr.14226.

- Koldehoff M, Lindemann M, Opalka B, et al. Cytomegalovirus induces apoptosis in acute leukemia cells as a virus-versus-leukemia function. Leuk Lymphoma. 2015;56:3189–97. https://doi.org/10.3109/10428194. 2015.1032968.
- Oberstein A, Shenk T. Cellular responses to human cytomegalovirus infection: Induction of a mesenchymal-to-epithelial transition (MET) phenotype. Proc Natl Acad Sci USA. 2017;114:E8244–53. https://doi.org/ 10.1073/pnas.1710799114.
- Yang R, Liang J, Xu GX, et al. Human cytomegalovirus glycoprotein B inhibits migration of breast cancer MDA-MB-231 cells and impairs TGF-β/Smad2/3 expression. Oncol Lett. 2018;15(5):7730–8. https://doi. org/10.3892/ol.2018.8344.
- Erlach KC, Böhm V, Seckert CK, et al. Lymphoma cell apoptosis in the liver induced by distant murine cytomegalovirus infection. J Virol. 2006;80:4801–19. https://doi.org/10.1128/JVI.80.10.4801-4819.2006.
- Erlach KC, Podlech J, Rojan A, Reddehase MJ. Tumor control in a model of bone marrow transplantation and acute liver-infiltrating B-cell lymphoma: an unpredicted novel function of cytomegalovirus. J Virol. 2002;76:2857–70. https://doi.org/10.1128/jvi.76.6.2857-2870.2002.
- Elmaagacli AH, Steckel NK, Koldehoff M, et al. Early human cytomegalovirus replication after transplantation is associated with a decreased relapse risk: evidence for a putative virus-versus-leukemia effect in acute myeloid leukemia patients. Blood. 2011;118:1402–12. https://doi. org/10.1182/blood-2010-08-304121.
- Green ML, Leisenring WM, Xie H, et al. CMV reactivation after allogeneic HCT and relapse risk: evidence for early protection in acute myeloid leukemia. Blood. 2013;122:1316–24. https://doi.org/10.1182/ blood-2013-02-487074.
- Koldehoff M, Ross SR, Dührsen U, et al. Early CMV-replication after allogeneic stem cell transplantation is associated with a reduced relapse risk in lymphoma. Leuk Lymphoma. 2017;58:822–33. https://doi.org/10.1080/10428194.2016.1217524 (PMID: 27687578).
- Inagaki J, Noguchi M, Kurauchi K, et al. Effect of cytomegalovirus reactivation on relapse after allogeneic hematopoietic stem cell transplantation in pediatric acute leukemia. Biol Blood Marrow Transplant. 2016;22:300–6. https://doi.org/10.1016/j.bbmt.2015.09.006.
- 27. Litjens NHR, van der Wagen L, Kuball J, Kwekkeboom J. Potential beneficial effects of cytomegalovirus infection after transplantation. Front Immunol. 2018;9:389. https://doi.org/10.3389/fimmu.2018.00389.
- 28. Lilleri D, Gerna G, Furione M, Zavattoni M, Spinillo A. Neutralizing and ELISA IgG antibodies to human cytomegalovirus glycoprotein complexes may help date the onset of primary infection in pregnancy. J Clin Virol. 2016;81:16–24. https://doi.org/10.1016/j.jcv.2016.05.007.
- Theobald SJ, Khailaie S, Meyer-Herman M, Volk V, Olbrich H, Danisch S. Signatures of T and B cell development, functional responses and PD-1 upregulation after HCMV latent infection and reactivations in Nod.Rag. Gamma Mice humanized with cord blood CD34+ cells. Front Immunol. 2018;9:2734. https://doi.org/10.3389/fimmu.2018.02734.
- Martins JP, Andoniou CE, Fleming P, Kuns RD, Schuster IS, Voigt V, et al. Strain-specific antibody therapy prevents cytomegalovirus reactivation after transplantation. Science. 2019;363(6424):288–93. https://doi.org/ 10.1126/science.aat0066.
- Gerna G, Lilleri D. Human cytomegalovirus (HCMV) infection/reinfection: development of a protective HCMV vaccine. New Microbiol. 2019;42(1):1–20 (PMID: 30671581).
- Theobald SJ, Kreer C, Khailaie S, Bonifacius A, Eiz-Vesper B, Figueiredo C, et al. Repertoire characterization and validation of gB-specific human IgG3 directly cloned from humanized mice vaccinated with dendritic cells and protected against HCMV. PLoS Pathog. 2020;16(7):e1008560. https://doi.org/10.1371/journal.ppat.1008560.
- 33. Fritz A, Percy C, Jack A, et al editors. International classification of diseases for oncology. 3rd ed. Malta: World Health Organization; 2013.
- Swerdlow SH, Campo E, Pilleri SA, Harris NL, Stein H, Siebert R, et al. The 2016 revision of the World Health Organizartion classification of lymphoid neoplasms. Blood. 2016;127:2375–90. https://doi.org/10.1182/ blood-2016-01-643569.
- 35. https://gco.iarc.fr/
- Zuhair M, Smit GSA, Wallis G, Jabbar F, Smith C, Devleesschauwer B, et al. Estimation of the worldwide seroprevalence of cytomegalovirus: a systematic review and meta-analysis. Rev Med Virol. 2019;29(3):e2034. https://doi.org/10.1002/rmv.2034.

- De Matos SB, Meyer R, Lima FW. Seroprevalence and serum profile of cytomegalovirus infection among patients with hematologic disorders in Bahia State. Brazil J Med Virol. 2011;83:298–304. https://doi.org/10. 1002/jmv.21965.
- Ng AP, Worth L, Chen L, Seymour JF, Prince HM, Slavin M, et al. Cytomegalovirus DNAemia and disease: incidence, natural history and management in settings other than allogeneic stem cell transplantation. Haematologica. 2005;90(12):1672–9 (PMID: 16330442).
- Marchesi F, Pimpinelli F, Gumenyuk S, Renzi D, Palombi F, Pisani F, et al. Cytomegalovirus reactivation after autologous stem cell transplantation in myeloma and lymphoma patients: a single-center study. World J Transplant. 2015;5:129–36. https://doi.org/10.5500/wjt.v5.i3.129.
- Capria S, Gentile G, Capobianchi A, Cardarelli L, Gianfelici V, Trisolini SM, et al. Prospective cytomegalovirus monitoring during first-line chemotherapy in patients with acute myeloid leukemia. J Med Virol. 2010;82(7):1201–7. https://doi.org/10.1002/jmv.21779.
- Zaidi ARZ, Al-Ammari MO, Al-Naamani M, Altaf SY, AlShehry N, Tailor IK, et al. Very high seroprevalence of cmv and ebv among a large series of patients with hematological malignancies at a tertiary care center in Saudi Arabia—a case for investigating cooperativity of viruses in carcinogenesis? Blood. 2019;134(Suppl 1):5818. https://doi.org/10.1182/ blood-2019-131323.
- 42. Xuan L, Huang F, Fan Z, Zhou H, Zhang X, Yu G, et al. Effects of intensified conditioning on Epstein-Barr virus and cytomegalovirus infections in allogeneic hematopoietic stem cell transplantation for hematological malignancies. J Hematol Oncol. 2012;5:46. https://doi.org/10.1186/1756-8722-5-46
- 43. Liu Y-C, Lu P-L, Hsiao H-H, Chang C-S, Liu T-C, Yang W-C, et al. Cytomegalovirus infection and disease after allogeneic hematopoietic stem cell transplantation: experience in a center with a high seroprevalence of both CMV and hepatitis B virus. Ann Hematol. 2012;91:587–95. https://doi.org/10.1007/s00277-011-1351-8.
- Tomonari A, Takahashi S, Ooi J, Tsukada N, Konuma T, Kato S, et al. Imapact of cytomgalovirus serostatus on outcome of unrelated cord blood transplantation for adults: a single-institute experience in Japan. Eur J Haematol. 2008;80(3):251–7. https://doi.org/10.1111/j.1600-0609.2007.
- Bernson E, Hallner A, Sander FE, Nicklasson M, Nilsson MS, Christenson K, et al. Cytomegalovirus serostatus affects autoreactive NK cells and outcomes of IL2-based immunotherapy in acute myeloid leukemia. Cancer Immunol Res. 2018;6:1110–9. https://doi.org/10.1158/2326-6066.CIR-17-0711.
- Kollman C, Howe CW, Anasetti C, Antin JH, Davies SM, Filipovich AH, et al. Donor characteristics as risk factors in recipients after transplantation of bone marrow from unrelated donors: the effect of donor age. Blood. 2001;98:2043–51. https://doi.org/10.1182/blood.V98.7.2043.
- Nichols WG, Corey L, Gooley T, Davis C, Boeckh M. High risk of death due to bacterial and fungal infection among cytomegalovirus (CMV)seronegative recipients of stem cell transplants from seropositive donors: evidence for indirect effects of primary CMV infection. J Infect Dis. 2002;185:273–82. https://doi.org/10.1086/338624.
- Pourgheysari B, Bruton R, Parry H, Billingham L, Fegan C, Murray J, et al. The number of cytomegalovirus-specific CD4+ T cells is markedly expanded in patients with B-cell chronic lymphocytic leukemia and determines the total CD4+ T-cell repertoire. Blood. 2010;116:2968–74. https://doi.org/10.1182/blood-2009-12-257147.
- Dafalla ABY, Elnil YFH, Gorish BMT. Seroprevalence of cytomegalovirus infection among leukemic patients in Khartoum state. Virol Mycol. 2018;7:2. https://doi.org/10.4172/2161-0517.1000183.
- Torres HA, Kontoyannis DP, Aguilera EA, Younes A, Luna MA, Tarrand JJ, et al. Cytomegalovirus infection in patients with lymphoma: An important cause of morbidity and mortality. Clin Lymphoma Myeloma. 2006;6(5):393–8. https://doi.org/10.3816/CLM.2006.n.016.
- Armitage JO, Gascoyne RD, Lunning MA, Cavalli F. Non-Hodgkin lymphoma. Lancet. 2017;390(10091):298–310. https://doi.org/10.1016/ S0140-6736(16)32407-2.
- Clarke CA, Glaser SL, Gomez SI, Wang SS, Keegan TH, Yang J, et al. Lymphoid malignancies in US Asians: incidence rate differences by birthplace and acculturation. Cancer Epidemiol Biomarkers Prev. 2011;20(6):1064–77. https://doi.org/10.1158/1055-9965.EPI-11-0038.

- 53. Li Y, Wang Z, Yi D, Ma S. Racial differences in three mayor NHL subtypes: descriptive epidemiology. Cancer Epidemiol. 2015;39(1):8–13. https://doi.org/10.1016/j.canep.2014.12.001.
- Hoshiba T, Asamoto A, Yabuki Y. Decreasing seropositivity of cytomegalovirus of pregnant women in Japan. Nihon Rinsho. 1998;56(1):193–6 (PMID: 9465689).
- Chihara D, Ito H, Matsuda T, Shibata A, Katsumi A, Nakamura S, et al. Differences in incidence and trends of haematological malignancies in Japan and the United States. Br J Haematol. 2014;164(4):536–45. https://doi.org/10.1111/bjh.12659.
- Howlader N, Noone AM, Krapcho M, et al. (Eds.). SEER Cancer Statistics Review, 1975–2014. National Cancer Institute, Bethesda, MD (https://seer.cancer.gov/csr/1975_2014/. Non-Hodgkin Lymphoma (Tab. 19.5), Hodgkin Lymphoma (Tab. 9.5), Leukemia (Tab. 13.5), Myeloma (Tab. 18.5), Age-Adjusted SEER Incidence Rates (Tab. 1.26, per 100,000 and age-adjusted to the 2000 US Std. Population).
- Yaqo RT, Jalal SD, Ghafour KJ, Hassan HA, Hughson MD. Non-Hodgkin lymphoma in the Middle East is characterized by low incidence rates with advanced age. J Glob Oncol. 2019;5:1–10. https://doi.org/10.1200/ JGO.18.00241.
- Bassig BA, Au X-Y, Mang Q, Ngan R, Morton LM, Dennis KM, et al. Subtype-specific incidence rates of lymphoid malignancies in Hong Kong compared to the United States, 2001–2010. Cancer Epidemiol. 2016;42:15–23. https://doi.org/10.1016/j.canep2016.02.007.
- Monga N, Nastoupil L, Garside J, Quigley J, Hudson M, O'Donovan P, et al. Burden of illness of follicular lymphoma and marginal zone lymphoma. Ann Hematol. 2019;98(1):175–83. https://doi.org/10.1007/ s00277-018-3501-8.
- Zhou L, Deng Y, Li N, Zheng Y, Tian T, Zhai Z, et al. Global, regional, and national burden of Hodgkin lymphoma from 1990 to 2017: estimates from the 2017 Global Burden of Disease study. J Hematol Oncol. 2019;12:107. https://doi.org/10.1186/s13045-019-0799-1.
- 61. Fitzmaurice C, Allen C, Barber RM, Barregard L, Bhutta ZA, Brenner H, Global Burden of Disease Cancer Collaboration, et al. Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 32 cancer groups, 1990 to 2015: a systematic analysis for the global burden of disease study. JAMA Oncol. 2017;3(4):524–48. https://doi.org/10.1001/jamaoncol.2016.5688.
- Liu W, Liu J, Song Y, Wang X, Zhou M, Wang L, et al. Union for China Lymphoma Investigators of the Chinese Society of Clinical Oncology Mortality of lymphoma and myeloma in China, 2004–2017: an observational study. J Hematol Oncol. 2019;12:22. https://doi.org/10.1186/s13045-019-0706-9.
- Liu J, Liu W, Mi L, Zeng X, Cai C, Ma J, et al. Union for China Leukemia Investigators of the Chinese Society of Clinical Oncology Incidence and mortality of multiple myeloma in China, 2006–2016: an analysis of the Global Burden of Disease Study 2016. J Hematol Oncol. 2019;12:136. https://doi.org/10.1186/s13045-019-0807-5.
- 64. Turesson I, Bjorkholm M, Hveding Blimark C, Kristinsson S, Velez R, Landgren O. Rapidly changing myeloma epidemiology in the general population: increased incidence, older patients, and longer survival. Eur J Haematol. 2018;101(2):237–44. https://doi.org/10.1111/ejh.13083.
- Kazandjian D. Multiple myeloma epidemiology and survival, a unique malignancy. Semin Oncol. 2016;43(6):676–81. https://doi.org/10.1053/j. seminoncol.2016.11.004.
- Winter JR, Taylor GS, Thomas OG, Jackson C, Lewis JEA, Stagg HR. Factors associated with cytomegalovirus serostatus in young people in England: a cross-sectional study. BMC Infect Dis. 2020;20(1):875. https://doi.org/10.1186/s12879-020-05572-9.
- 67. Lin J, Lin W, Mi L, Zeng X, Cai C, Ma J, et al. Incidence and mortality of multiple myeloma in China, 2006–2016: an analysis of the Global Burden of Disease Study 2016. J Haematol Oncol. 2019;12:136. https://doi.org/10.1186/s13045-019-0807-5.
- Dong Y, Shi O, Zeng Q, Lu X, Wang W, Li Y, et al. Leukemia incidence trends at the global, regional, and national level between 1990 and 2017. Exp Hematol Oncol. 2020;9:14. https://doi.org/10.1186/ s40164-020-00170-6.
- American Cancer Society, Inc.: Global Cancer Facts and Figures (4th ed., Source: GLOBOCAN 2018), Atlanta, GA, the US, American Cancer Society, Tables 4, 7, pp. 28, 59 (2018).

- Cowan AJ, Allen C, Barac A, Basaleem H, Bensenor I, Curado MP, et al. Global burden of multiple myeloma: a systematic analysis for the global burden of disease study 2016. JAMA Oncol. 2018;4(9):1221–7. https:// doi.org/10.1001/jamaoncol.20182128.
- Lee SJ, Tien HF, Park HJ, Kim JA, Lee DS. Gradual increase of chronic lymphocytic leukemia incidence in Korea, 1999–2010: comparison to plasma cell myeloma. Leuk Lymphoma. 2016;57(3):585–9. https://doi. org/10.3109/10428194.2015.1068307.
- Wu SJ, Huang SY, Lin CT, Lin YJ, Chang CJ, Tien HF. The incidence of chronic lymphocytic leukemia in Taiwan, 1986–2005: a distinct increasing trend with birth-control effect. Blood. 2010;116(22):4430–5. https:// doi.org/10.1182/blood-2010-05-285221.
- Herbein G. The human cytomegalovirus, from oncomodulation to oncogenesis. Viruses. 2018;10(8):408. https://doi.org/10.3390/v1008 0408.
- Kumar A, Coquard L, Pasquereau S, Russo L, Valmary-Degano S, Borg C, et al. Tumor control by human cytomegalovirus in a murine model of hepatocellular carcinoma. Mol Ther Oncolytics. 2016;3:16012. https:// doi.org/10.1038/mto.2016.12.
- Jurak I, Brune W. Induction of apoptosis limits cytomegalovirus crossspecies infection. EMBO J. 2006;25:2634–42. https://doi.org/10.1038/sj. emboi.7601133.
- Erkes DA, Wilski NA, Snyder CM. Intratumoral infection by CMV may change the tumor environment by directly interacting with tumorassociated macrophages to promote cancer immunity. Hum Vaccin Immunother. 2017;13:1778–85. https://doi.org/10.1080/21645515.2017. 1331795.
- Ben-Bassat I, Raanani P, Gale RP. Graft-versus-leukemia in chronic lymphocytic leukemia. Bone Marrow Transplant. 2007;39:441–6. https://doi. org/10.1038/sj.bmt.1705619.
- 78. Gustafsson B, Jernberg AG, Priftakis P, Bogdanovic G. No CMV DNA in Guthrie cards from children who later developed ALL. Pediatr Hematol Oncol. 2006;23(3):199–205. https://doi.org/10.1080/08880010500506677.
- MacKenzie J, Gallagher A, Clayton RA, Perry J, Eden OB, Ford AM, et al. Screening for herpesvirus genomes in common acute lymphoblastic leukemia. Leukemia. 2001;15(3):415–42. https://doi.org/10.1038/sj.leu. 2402049.
- Bogdanovic G, Jernberg AG, Priftakis P, Grillner L, Gustafsson B. Human herpes virus 6 or Epstein-Barr virus were not detected in Guthrie cards from children who later developed leukaemia. Br J Cancer. 2004;91(5):913–5. https://doi.org/10.1038/sj.bjc.6602099.
- 81. Tomoka T, Montgomery ND, Eric Powers E, Dhungel MB, Morgan EA, Mulenga M, et al. Lymphoma and pathology in Sub-saharan Africa: current approaches and future directions. Clin Lab Med. 2018;38(1):91–100. https://doi.org/10.1016/j.cll.2017.10.007.
- Fassas AB-T, Bolaños-Meade J, Buddharaju LN, Rapoport A, Cottler-Fox M, Chen T, et al. Cytomegalovirus infection and non-neutropenic fever after autologous stem cell transplantation: high rates of reactivation in patients with multiple myeloma and lymphoma. Br J Haematol. 2001;112:237–41. https://doi.org/10.1046/j.1365-2141.2001.02487.x.
- Nguyen Q, Estey E, Raad I, Rolston K, Kantarjian H, Jacobson K, et al. Cytomegalovirus pneumonia in adults with leukemia: an emerging problem. Clin Infect Dis. 2001;32(4):539–45. https://doi.org/10.1086/ 318721.
- Lee SJ, Klein J, Haagenson M, Baxter-Lowe LA, Confer DL, Eapen M, et al. High-resolution donor-recipient HLAmatching contributes to the success of unrelated donor marrow transplantation. Blood. 2007;110(13):4576–83. https://doi.org/10.1182/blood-2007-06-097386.
- Behrendt CE, Rosenthal J, Bolotin E, Nakamura R, Zaia J, Forman SJ.
 Donor and recipient cmv serostatus and outcome of pediatric allogeneic HSCT for acute leukemia in the era of CMVpreemptive therapy. Biol Blood Marrow Transplant. 2009;15(1):54–60. https://doi.org/10.1016/j.bbmt.2008.10.023.
- Dieamant DC, Bonon SHA, Peres RMB, Costa CRC, Albuquerque DM, Miranda ECM, et al. Cytomegalovirus (CMV) genotype in allogeneic hematopoietic stem cell transplantation. BMC Infect Dis. 2013;13:310. https://doi.org/10.1186/1471-2334-13-310.
- Ferrés M, Nervi B, Ramírez P. Prophylaxis against cytomegalovirus infection in pediatric and adult patients undergoing solid organ

- and hematopoietic stem cells transplantation. Rev Chilena Infectol. 2012;29(Suppl. 1):23–8. https://doi.org/10.4067/S0716-1018201200 0500004
- Duval M, Klein JP, He W, Cahn J-Y, Cairo M, Camitta BM, et al. Hematopoietic stem-cell transplantation for acute leukemia in relapse or primary induction failure. J Clin Oncol. 2010;28(23):3730–8. https://doi. org/10.1200/JCO.2010.28.8852.
- Teira P, Battiwalla M, Ramanathan M, Barrett MA, Ahn WA, Chen M, et al. Early cytomegalovirus reactivation remains associated with increased transplant-related mortality in the current era: a CIBMTR analysis. Blood. 2016;127(20):2427–38. https://doi.org/10.1182/blood-2015-11-679639.
- Cornelissen JJ, Carston M, Kollman C, King R, Dekker AW, Löwenberg B, et al. Unrelated marrow transplantation for adult patients with poor-risk acute lymphoblastic leukemia: strong graft-versus-leukemia effect and risk factors determining outcome. Blood. 2001;97(6):1572–7. https://doi. org/10.1182/blood.v97.6.1572.
- Marty FM, Ljungman P, Chemaly RF, Maertens J, Dadwal SS, Duarte RF, et al. Letermovir prophylaxis for cytomegalovirus in hematopoietic-cell transplantation. New Engl J Med. 2017;377(25):2433–44. https://doi. org/10.1056/NEJMoa1706640.
- Fleming T, Dunne J, Crowley B. Ex vivo monitoring of human cytomegalovirus-specific CD8(+) T-Cell responses using the QuantiFERON-CMV assay in allogeneic hematopoietic stem cell transplant recipients attending an Irish hospital. J Med Virol. 2010;82(3):433–40. https://doi. org/10.1002/jmv.21727.
- Ljungman P, Brand R, Hoek J, de la Camara R, Cordonnier C, Einsele H, et al. For the Infectious Diseases Working Party of the European Group for Blood and Marrow Transplantation. Donor cytomegalovirus status influences the outcome of allogeneic stem cell transplant: a study by the European group for blood and marrow transplantation. Clin Infect Dis. 2014;59(4):473–81. https://doi.org/10.1093/cid/ciu364.
- Ljungman P, de la Camara R, Milpied N, Volin L, Russell CA, Crisp A, et al.
 The Valacyclovir International Bone Marrow Transplant Study Group.
 Randomized study of valacyclovir as prophylaxis against cytomegalovirus reactivation in recipients of allogeneic bone marrow transplants.
 Blood. 2002;99(8):3050–6. https://doi.org/10.1182/blood.v99.8.3050.
- Mehta RS, Holtan SG, Wang T, Hemmer MT, Spellman SR, Arora M, et al. GRFS and CRFS in alternative donor hematopoietic cell transplantation for pediatric patients with acute leukemia. Blood Adv. 2019;3(9):1441–9. https://doi.org/10.1182/bloodadvances.2018030171.
- Rubio MT, Savani BN, Labopin M, Polge E, Niederwieser D, Ganser A, et al. The impact of HLA-matching on reduced intensity conditioning regimen unrelated donor allogeneic stem cell transplantation for acute myeloid leukemia in patients above 50 years—a report from the EBMT acute leukemia working party. J Hematol Oncol. 2016;9(1):65. https:// doi.org/10.1186/s13045-016-0295-9.
- Dalle J-H, Balduzzi A, Bader P, Lankester A, Yaniv I, Wachowiak J, et al. Allogeneic stem cell transplantation from HLA-mismatched donors for pediatric patients with acute lymphoblastic leukemia treated according to the 2003 BFM and 2007 International BFM Studies: impact of disease risk on outcomes. Biol Blood Marrow Transplant. 2018;24(9):1848–55. https://doi.org/10.1016/j.bbmt.2018.05.009.
- Roth-Guepin G, Canaani J, Ruggeri A, Labopin M, Finke J, Cornelissen JJ, et al. Allogeneic stem cell transplantation in acute lymphoblastic leukemia patients older than 60 years: a survey from the acute leukemia working party of EBMT. Oncotarget. 2017;8(68):112972–9. https://doi. org/10.18632/oncotarget.22934.
- Debaugnies F, Busson L, Ferster A, Lewalle P, Azzi N, Aoun M, et al. Detection of *Herpesviridae* in whole blood by multiplex PCR DNAbased-microarray analysis after hematopoietic stem cell transplantation. J Clin Microb. 2014;52(7):2552–6. https://doi.org/10.1128/JCM.00061-14.
- 100. Kielsen K, Enevold C, Heilmann C, Sengeløv H, Pedersen AE, Ryder LP, et al. Donor genotype in the interleukin-7 receptor α-chain predicts risk of graft-versus-host disease and cytomegalovirus infection after allogeneic hematopoietic stem cell transplantation. Front Immunol. 2018;9:109. https://doi.org/10.3389/fimmu.2018.00109.
- 101. Meijer E, Dekker AW, Rozenberg-Arska M, Weersink AJL, Verdonck LF. Influence of cytomegalovirus seropositivity on outcome after T cell-depleted bone marrow transplantation: contrasting results between recipients of grafts from related and unrelated donors. Clin Infecti Dis. 2002;35:703–12. https://doi.org/10.1086/342332.

- 102. Broers AEC, van der Holt R, van Esser JWJ, Gratama J-W, Henzen-Logmans S, Kuenen-Boumeester V, et al. Increased transplant-related morbidity and mortality in CMV-seropositive patients despite highly effective prevention of CMVdisease after allogeneic T-cell-depleted stem cell transplantation. Blood. 2000;95(7):2240–5. https://doi.org/10. 1182/blood.V95.7.2240.
- Jaskula E, Dlubek D, Tarnowska A, Lange J, Mordak-Domagala M, Suchnicki K, et al. Anti-CMV-IgG positivity of donors is beneficial for alloHSCT recipients with respect to the better short-term immunological recovery and high level of CD4+CD25high lymphocytes. Viruses. 2015;7(3):1391–408. https://doi.org/10.3390/v7031391.
- Michálek J, Horvath R. High incidence of Epstein-Barr virus, cytomegalovirus and human herpesvirus 6 infections in children with cancer. BMC Pediatr. 2002;2:1. https://doi.org/10.1186/1471-2431-2-1.
- 105. Kröger N, Zabelina T, Krüger W, Renges H, Stute N, Schrum J, et al. Patient cytomegalovirus seropositivity with or without reactivation is the most important prognostic factor for survival and treatment-related mortality in stem cell transplantation from unrelated donors using pretransplant in vivo T-cell depletion with anti-thymocyte globulin. Br J Haematol. 2001;113(4):1060–71. https://doi.org/10.1046/j.1365-2141.
- 106. Vdovin AS, Filkin SY, Yefimova PR, Sheetikov SA, Kapranov NM, Davydova YO, et al. Recombinant MHC tetramers for isolation of virus-specific CD8 + cells from healthy donors: potential approach for cell therapy of posttransplant cytomegalovirus infection. Biochemistry (Mosc). 2016;81(11):1371–83. https://doi.org/10.1134/S0006297916110146.
- 107. Peric Z, Wilson J, Durakovic N, Ostojic A, Desnica L, Rezo Vranjes V, et al. Early human cytomegalovirus reactivation is associated with lower incidence of relapse of myeloproliferative disorders after allogeneic hematopoietic stem cell transplantation. Bone Marrow Transpl. 2018;53(11):1450–6. https://doi.org/10.1038/s41409-018-0172-y.
- 108. Mariotti J, Maura F, Spina F, Roncari L, Dodero A, Farina L, et al. Impact of cytomegalovirus replication and cytomegalovirus serostatus on the outcome of patients with B cell lymphoma after allogeneic stem cell transplantation. Biol Blood Marrow Transplant. 2014;20(6):885–90. https://doi.org/10.1016/j.bbmt.2014.02.015.
- Schmidt-Hieber M, Labopin M, Beelen D, Volin L, Ehninger G, Finke J, et al. CMV serostatus still has an important prognostic impact in de novo acute leukemia patients after allogeneic stem cell transplantation: a report. Blood. 2013;122(19):3359–64. https://doi.org/10.1182/ blood-2013-05-499830.
- Piñana JL, Martino P, Barba P, Margall N, Roig MC, Valcárcel D, et al. Cytomegalovirus infection and disease after reduced intensity conditioning allogeneic stem cell transplantation: single-centre experience. Bone Marrow Transplant. 2010;45:534–42. https://doi.org/10.1038/bmt.2009. 180.
- Al-Sweedan S, Al-Seraihy A, Al-Ahmari A, Al-Jefri A, Mohammed V, Jafri R, et al. Factors determining the outcome of hematopoietic stem cell transplantation in patients with acute lymphoblastic leukemia at King Faisal Specialist Hospital and Research Center, Riyadh. Saudi Arabia J Pediatr Hematol/Oncol. 2017;39(1):33–7. https://doi.org/10.1097/MPH. 0000000000000679.
- 112. Al-Hajjar S, Al Seraihi A, Al Muhsen S, Ayas M, Al Jumaah S, Al Jefri A, et al. Cytomegalovirus infections in unrelated cord blood transplantation in pediatric patients: incidence, risk factors, and outcomes. Hematol Oncol Stem Cell Ther. 2011;4(2):67–72. https://doi.org/10.5144/1658-3876.2011.67.
- Hussein AA, Al-Antary ET, Najjar R, Al-Hamdan DS, Al-Zaben A, Frangoul H. Incidence and risk factors for cytomegalovirus (CMV) reactivation following autologous hematopoietic stem cell transplantation in children. Pediatr Blood Cancer. 2015;62(6):1099–101. https://doi.org/10.1002/ pbc.25292.
- 114. Al Mana H, Yassine HM, Younes NN, Al-Mohannadi A, Al-Sadeq DW, Alhababi D, et al. The current status of cytomegalovirus (CMV) prevalence in the MENA region: a systematic review. Pathogens. 2019;8(4):213. https://doi.org/10.3390/pathogens8040213.
- Cohen L, Yeshurun M, Shpilberg O, Ram R. Risk factors and prognostic scale for cytomegalovirus (CMV) infection in CMV-seropositive patients after allogeneic hematopoietic cell transplantation. Transpl Infect Dis. 2015;17(4):510–7. https://doi.org/10.1111/tid.12398.

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- Valadkhani B, Kargar M, Ashouri A, Hadjibabaie M, Gholami K, Ghavamzadeh A. The risk factors for cytomegalovirus reactivation following stem cell transplantation. J Res Pharm Pract. 2016;5(1):63–9. https://doi.org/10.4103/2279-042X.176554.
- Safabakhsh H, Tehranian F, Tehranian B, Hatami H, Karimi G, Shahabi M. Prevalence of anti-CMV antibodies in blood donors in Mashhad. Iran Iran J Epidemiol. 2013;9(1):52–7.
- Sepehrvand N, Khameneh ZR, Eslamloo HRF. Survey the seroprevalence of CMV among hemodialysis patients in Urmia, Iran. Saudi J Kidney Dis Transpl. 2010;21(2):363–7 (PMID: 20228534).
- Behzad-Behbahani A, Ehsanipour F, Alborzi A, Nourani H, Ramzi M, Rasoli M. Qualitative detection of human cytomegalovirus DNA in the plasma of bone marrow transplant recipients: Value as a predictor of disease progression. Exp Clin Transplant. 2004;2(1):196–200 (PMID: 15859928).
- Ziyaeyan M, Sabahi F, Alborzi A, Mahboudi F, Kazemnejad A, Ramzi M, et al. Diagnosis and monitoring of human cytomegalovirus infection in bone marrow transplant recipients by quantitative competitive PCR. Exp Clin Transplant. 2006;4(1):470–4 (PMID: 16827646).
- Handous I, Achour B, Marzouk M, Rouis S, Hazgui O, Brini I, et al. Coinfections of human herpesviruses (CMV, HHV-6, HHV-7 and EBV) in nontransplant acute leukemia patients undergoing chemotherapy. Virol J. 2020;17:37. https://doi.org/10.1186/s12985-020-01302-4.
- 122. Gawad AA, Hashish M, Abaza A, El-Kayal A. Cytomegalovirus immunoglobulin G avidity index among blood donors in Alexandria. Egypt Cent Eur J Public Health. 2016;24(4):314–20. https://doi.org/10.21101/cejph. a4157.
- 123. Zekri ARN, Mohamed WS, Samra MA, Sherif GM, El-Shehaby AM, El-Sayed MH. Risk factors for cytomegalovirus, hepatitis B and C virus reactivation after bone marrow transplantation. Transpl Immunol. 2004;13(4):305–11. https://doi.org/10.1016/j.trim.2004.10.001.
- 124. Loutfy SA, Abo-Shadi MA, Fawzy M, El-Wakil M, Metwally SA, Moneer MM, et al. Epstein-Barr virus and cytomegalovirus infections and their clinical relevance in Egyptian leukemic pediatric patients. Virol J. 2017;14(1):46. https://doi.org/10.1186/s12985-017-0715-7.
- 125. George B, Pati N, Gilroy N, Ratnamohan M, Huang G, Kerridge I, et al. Pre-transplant cytomegalovirus (CMV) serostatus remains the most important determinant of CMV reactivation after allogeneic hematopoietic stem cell transplantation in the era of surveillance and preemptive therapy. Transpl Infect Dis. 2010;12(4):322–9. https://doi. org/10.1111/j.1399-3062.2010.00504.x.
- 126. Devasia AJ, Mammen S, Korula A, Abraham A, Fouzia NA, Lakshmi KM, et al. A low incidence of cytomegalo virus infection following allogeneic hematopoietic stem cell transplantation despite a high sero-prevalence. Indian J Hematol Blood Transfus. 2018;34(4):636–42. https://doi.org/10.1007/s12288-018-0960-y.
- Azanan MS, Abdullah NK, Chua LL, Lum SH, Ghafar SSA, Kamarulzaman A, et al. Immunity in young adult survivors of childhood leukemia is similar to the elderly rather than age-matched controls: role of cytomegalovirus. Eur J Immunol. 2016;6(7):1715–26. https://doi.org/10.1002/eii.201646356.
- 128. Du J, Liu J, Gu J, Zhu P. HLA-DRB1*09 is associated with increased incidence of cytomegalovirus infection and disease after allogeneic hematopoietic stem cell transplantation. Biol Blood Marrow Transplant. 2007;13(12):1417–21. https://doi.org/10.1016/j.bbmt.2007.09.003.

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